



REVIEW ARTICLE OPEN

Epigenetics-targeted drugs: current paradigms and future challenges

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Epigenetics governs a chromatin state regulatory system through five key mechanisms: DNA modification, histone modification, RNA modification, chromatin remodeling, and non-coding RNA regulation. These mechanisms and their associated enzymes convey genetic information independently of DNA base sequences, playing essential roles in organismal development and homeostasis. Conversely, disruptions in epigenetic landscapes critically influence the pathogenesis of various human diseases. This understanding has laid a robust theoretical groundwork for developing drugs that target epigenetics-modifying enzymes in pathological conditions. Over the past two decades, a growing array of small molecule drugs targeting epigenetic enzymes such as DNA methyltransferase, histone deacetylase, isocitrate dehydrogenase, and enhancer of zeste homolog 2, have been thoroughly investigated and implemented as therapeutic options, particularly in oncology. Additionally, numerous epigenetics-targeted drugs are undergoing clinical trials, offering promising prospects for clinical benefits. This review delineates the roles of epigenetics in physiological and pathological contexts and underscores pioneering studies on the discovery and clinical implementation of epigenetics-targeted drugs. These include inhibitors, agonists, degraders, and multitarget agents, aiming to identify practical challenges and promising avenues for future research. Ultimately, this review aims to deepen the understanding of epigenetics-oriented therapeutic strategies and their further application in clinical settings.

Signal Transduction and Targeted Therapy (2024)9:332

; <https://doi.org/10.1038/s41392-024-02039-0>

INTRODUCTION

From a historical perspective, the term “epigenetics” was first introduced by Conrad Waddington in 1942 to describe heritable changes in gene function that do not involve alterations to the DNA sequence, leading to changes in biological phenotypes. Following nearly a century of rigorous research, a diverse array of epigenetic-modifying enzymes has been identified, and the elucidation of distinct molecular mechanisms has established epigenetics as a robust discipline.¹ Presently, epigenetics is defined as a chromatin state regulatory system comprised of five principal mechanisms: DNA modifications,² histone modifications,³ RNA modifications,⁴ chromatin remodeling,⁵ and the regulation based on non-coding RNA (ncRNA).⁶ These mechanisms independently transmit genetic information from the DNA sequence, enabling the activation or repression of specific genome regions in response to physiological or pathological signals (Fig. 1).

Enzymes that regulate epigenetic modifications are categorized into “writers,” “erasers,” “readers,” and “remodelers” based on their functions.^{7–9} Writers modify specific bases or amino acids, whereas erasers remove these modifications, exerting reciprocal

effects on gene expression. For instance, DNA methyltransferase (DNMT) catalyzes the addition of methyl groups to form 5-methylcytosine (m5C) in DNA bases,¹⁰ whereas the ten-eleven translocation (TET) enzymes initiate DNA demethylation, converting m5C into derivatives, such as 5-hydroxymethylcytosine (5hmC), 5-formylcytosine, and 5-carboxycytosine.¹¹ Typically, genes expressed at higher levels exhibit lower methylation, whereas genes with lower expression levels tend to be more heavily methylated.¹² Readers are proteins that contain specific motifs to recognize and bind these modifications, such as the methyl-CpG-binding domain (MBD) responsible for recognizing 5mC.¹³ These proteins influence chromatin status and recruit or collaborate with other enzymes to regulate gene expression.^{13,14} Remodelers are crucial in chromatin remodeling, moving or removing nucleosomes at vital regulatory elements like enhancers and promoters to modify chromatin accessibility.¹⁵ Furthermore, as unique epigenetic regulators distinct from epigenetic-modifying enzymes, ncRNAs directly bind to various genomic regions or specific RNA sequences to modulate gene expression.¹⁶ Variations in the given ncRNA may regulate the interactions or functions of its interactor partners, including proteins, RNAs, DNAs,

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Received: 2 August 2024 Revised: 14 October 2024 Accepted: 29 October 2024

Published online: 26 November 2024

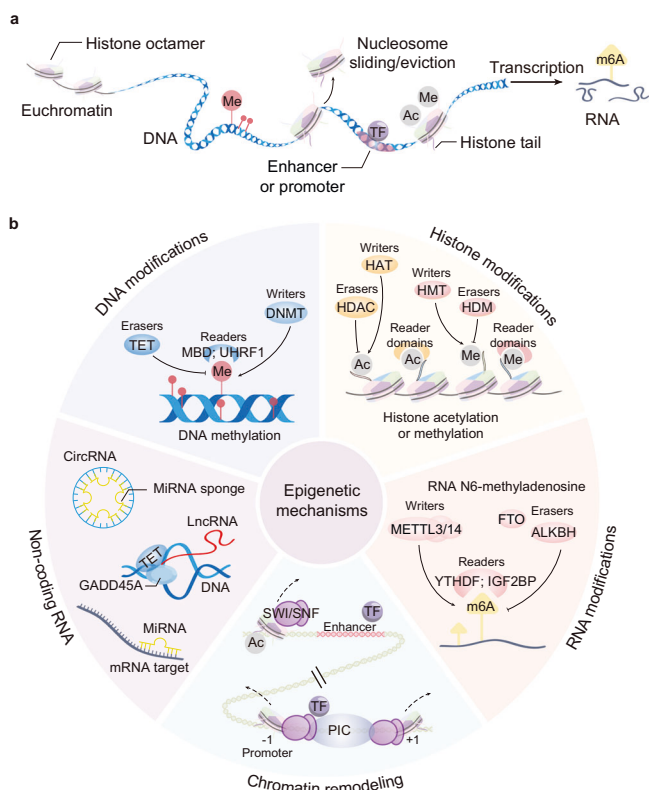


Fig. 1 Epigenetic mechanisms and key examples of widely studied modifications and their modifying enzymes. **a** DNA modifications, histone modifications, RNA modifications, chromatin remodeling, and the regulation based on non-coding RNA constitute the core content of epigenetics, being responsible for passing on heritable variations of genetic information independently of the DNA sequence. **b** Epigenetic modifications are reversible progress catalyzed by functionally complementary modifying enzymes, which provide targets for disease therapeutics

and lipids, thereby influencing various biological cellular processes or pathological phenotypes.¹⁷

The discovery of functionally complementary epigenetic-modifying enzymes has underscored the reversibility of most known epigenetic modifications. This insight supports the development of strategies to modulate gene expression via targeted regulation of these enzymes, providing a strong theoretical basis for creating novel therapeutic approaches from an epigenetic perspective. To date, four categories of epigenetics-targeted drugs have received the Food and Drug Administration (FDA) approval for clinical use, with numerous clinical trials ongoing to refine their applications. A timeline of significant milestones in epigenetic research is depicted in Fig. 2.

Over the past few decades, numerous studies have underscored that abnormalities in the expression and function of epigenetic-regulating enzymes are crucial in the onset and progression of various diseases. Epigenetics-targeted drugs, therefore, have emerged as pivotal topics due to their significant physiological and pathological implications. The development of drug screening models rooted in epigenetic principles is anticipated to substantially expand therapeutic options in clinical settings. Moreover, advancements in epigenetic analysis and molecular modification techniques have accelerated the clinical adoption of these targeted drugs. Despite these developments, there remains a gap in comprehensive reviews that address epigenetic regulations in physiological and disease contexts and detail the latest advancements in drug

development targeting these mechanisms. This review aims to fill that void by summarizing the current understanding of epigenetic regulations and clinical trials of targeted drugs, thereby outlining the future application of these promising agents. We begin with an overview of epigenetic mechanisms and their crucial roles in health and disease, followed by an in-depth discussion on the exploration and application of marketed epigenetic drugs. We then provide a systematic account of recent progress in developing potential therapeutic agents targeting various epigenetic enzymes, highlighting emerging research trends. Finally, we present the breakthroughs and challenges in epigenetic drugs, particularly the benefits of combining them with traditional therapies such as radiotherapy, chemotherapy, and targeted therapy, to underscore their potential in translational medicine.

BIOLOGICAL AND PATHOLOGICAL ROLES OF EPIGENETICS

Epigenetic modifications are a fundamental mechanism regulating gene expression, crucial for various cellular functions. Dysregulated epigenetic regulators, whether overexpressed or underactive, compromise normal functions and contribute to disease onset. Thus, epigenetic modifications hold significant potential for disease treatment and biotechnological applications, driving the development of targeted therapeutic drugs.

Epigenetics and early embryonic development

Epigenetic landscapes undergo substantial changes to ensure the coordinated progression of embryogenesis and subsequent development throughout an individual's life.¹⁸ Mutations in epigenetic-modifying enzymes, whether heterozygous or hemizygous, are commonly associated with congenital conditions, such as Rubinstein-Taybi syndrome, linked to mutations in the cyclic adenosine monophosphate-responsive element-binding protein (CREB)-binding protein (CBP) and its paralog, E1A-binding protein (P300),¹⁹ immunodeficiency-centromeric instability-facial anomalies syndrome related to DNMT3B mutations,²⁰ and Kabuki syndrome due to mutations in lysine methyltransferase 2D (KMT2D).²¹ DNA methylation reprogramming, a pivotal aspect of epigenetic modification in early embryonic stages, involves genome-wide removal of epigenetic marks through extensive DNA demethylation, followed by remethylation.²² This process, integral to mammalian development, has only been fully understood with the advent of whole-genome bisulfite sequencing, which allows for single-base resolution analysis of DNA methylation kinetics.^{23,24} Advances in precise assays for assessing DNA methylation at specific genetic loci have led to significant insights into these epigenomic reprogramming processes. This reprogramming results in global hypomethylation and significant loss of genetic memory, which is foundational for acquiring pluripotency and redetermining cell fate.²⁵ Following fertilization, methylation patterns evolve progressively, enabling cells to differentiate and contribute to the development of various biological systems. The dynamic regulation of DNA methylation, including reprogramming, is indispensable for mammalian development and differentiation. Another vital mechanism, histone modification, plays a critical role during zygotic genome activation (ZGA), which involves the transition of the zygotic genome from a state of silence to active transcription.²⁶ Notably, the de novo establishment of histone 3 lysine 14 acetylation (H3K14ac) and histone 3 lysine 9 trimethylation (H3K9me3) following fertilization is crucial for the timely activation of ZGA genes during development.^{27,28} The SWI/SNF complex also plays a significant role in the precise activation and repression of tissue-specific transcription factors, functioning as a chromatin remodeler that orchestrates the coordinated differentiation of multiple cell lineages during

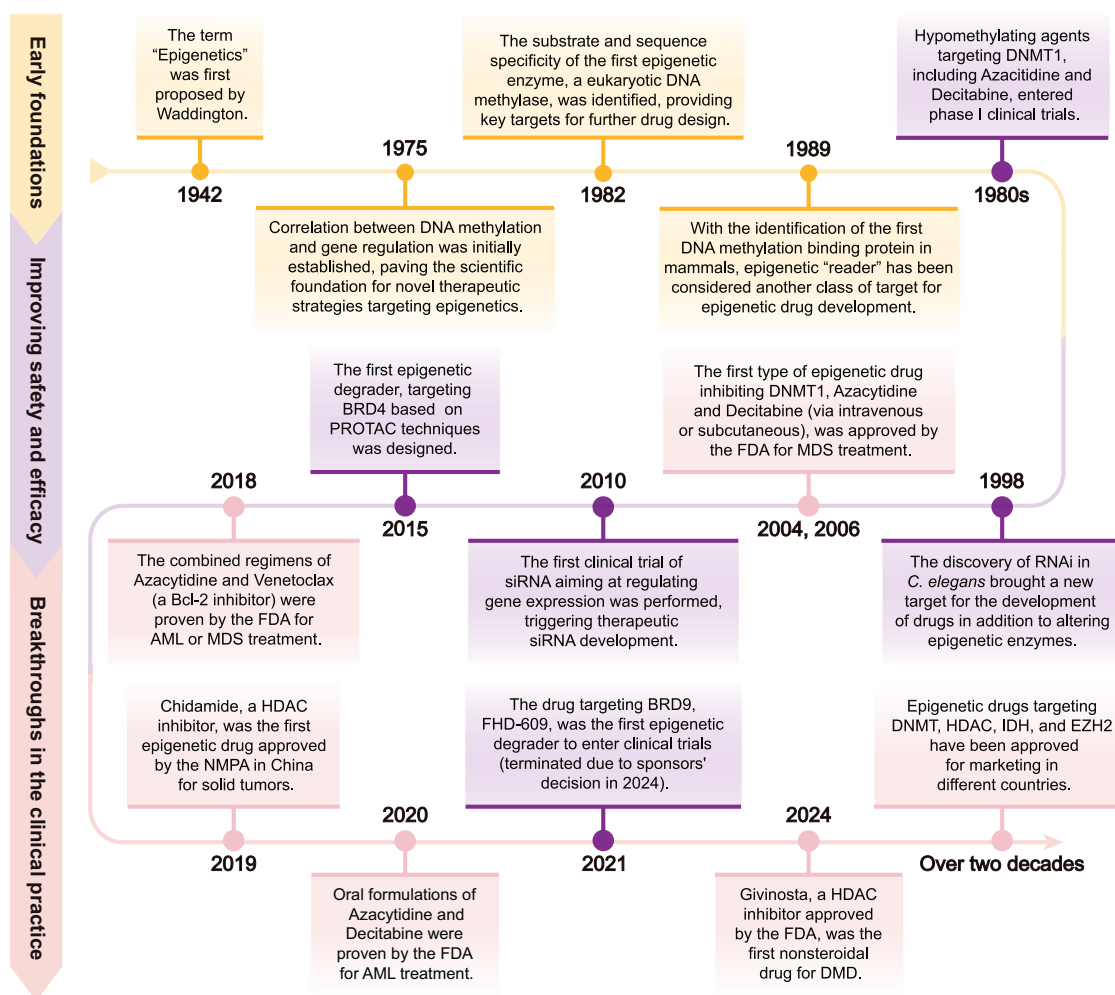


Fig. 2 Timeline of major discoveries and advances in epigenetic research. The significant discoveries and advances are depicted in the illustrator and displayed as primarily "Early foundations" (yellow boxes) on the top, "Improving safety and efficacy" (purple boxes) in the middle, and "Breakthroughs in the clinical practice" (pink boxes) at the bottom

development.^{29,30} Additionally, the role of RNA modifications in embryo development is increasingly recognized and summarized.^{31,32} Recent studies indicate that deficiencies in methyltransferase-like proteins (METTLs) and their associated RNA N6-methyladenosine (m6A) levels can induce G1/S cell cycle arrest in hematopoietic stem and progenitor cells in model organisms.³³ Furthermore, aberrant RNA modification patterns are integrated into the regulatory networks of other epigenetic mechanisms, such as histone deubiquitylation and DNA methylation, playing critical roles in nuclear reprogramming.^{34,35} Recently, preliminary evidence of ncRNAs being engaged in embryo development has been proposed according to the reported variation among ncRNAs contents during different stages of early embryonic development in mouse models.^{36–38} As an indicator of developmental competence, ncRNA plays an irreplaceable role in the continuous stages of pre-implantation development, embryo implantation, and post-implantation development.³⁹ Aberrant levels of certain ncRNAs may disturb the transition of fertilized oocytes to pluripotent blastocysts, and may even affect the differentiation of epiblast stem cells.^{40,41} Notably, ncRNAs may also act as regulatory factors for other epigenetic mechanisms. For example, during mouse ZGA, the negative regulation of Dnmt3a/3b expression by microRNA-29b (miR-29b) helps maintain proper DNA demethylation to establish the imprinting of genes.⁴² However, considering that most of the current understanding has only been validated in animal

models, much work is still required to explore the role of ncRNAs during embryonic and fetal development in humans.

Epigenetics and aging

Since the late 1990s and early 21st century, researchers have observed that epigenetic changes accompany aging based on data derived from cellular experiments.⁴³ Initially, it was unclear whether these epigenetic alterations were a cause or a consequence of aging. Recent work by Yang et al.⁴⁴ has successfully dissociated epigenetic dysregulation from genetic changes, confirming that the collapse of epigenetic modifications is a potent driver of aging. DNA methylation, a central epigenetic mechanism, regulates both development and aging. Notably, global DNA methylation levels in most regeneratively capable tissues tend to decrease with age.⁴⁵ Beyond global changes, studies increasingly report high frequencies of age-related alterations in DNA methylation accumulated in specific cellular regions. These differentially methylated regions associated with aging lead to either the upregulation or repression of downstream genes. For example, age-related hypermethylation within the promoter regions of tumor suppressor and metabolic genes may partially explain the increased susceptibility of the elderly to tumors and various metabolic disorders.⁴⁶ Conversely, DNMT inhibitor decitabine can reverse hypermethylation in tumor suppressors, enhancing their expression and inducing a senescence-like phenotype in tumor cell lines.⁴⁷ Other

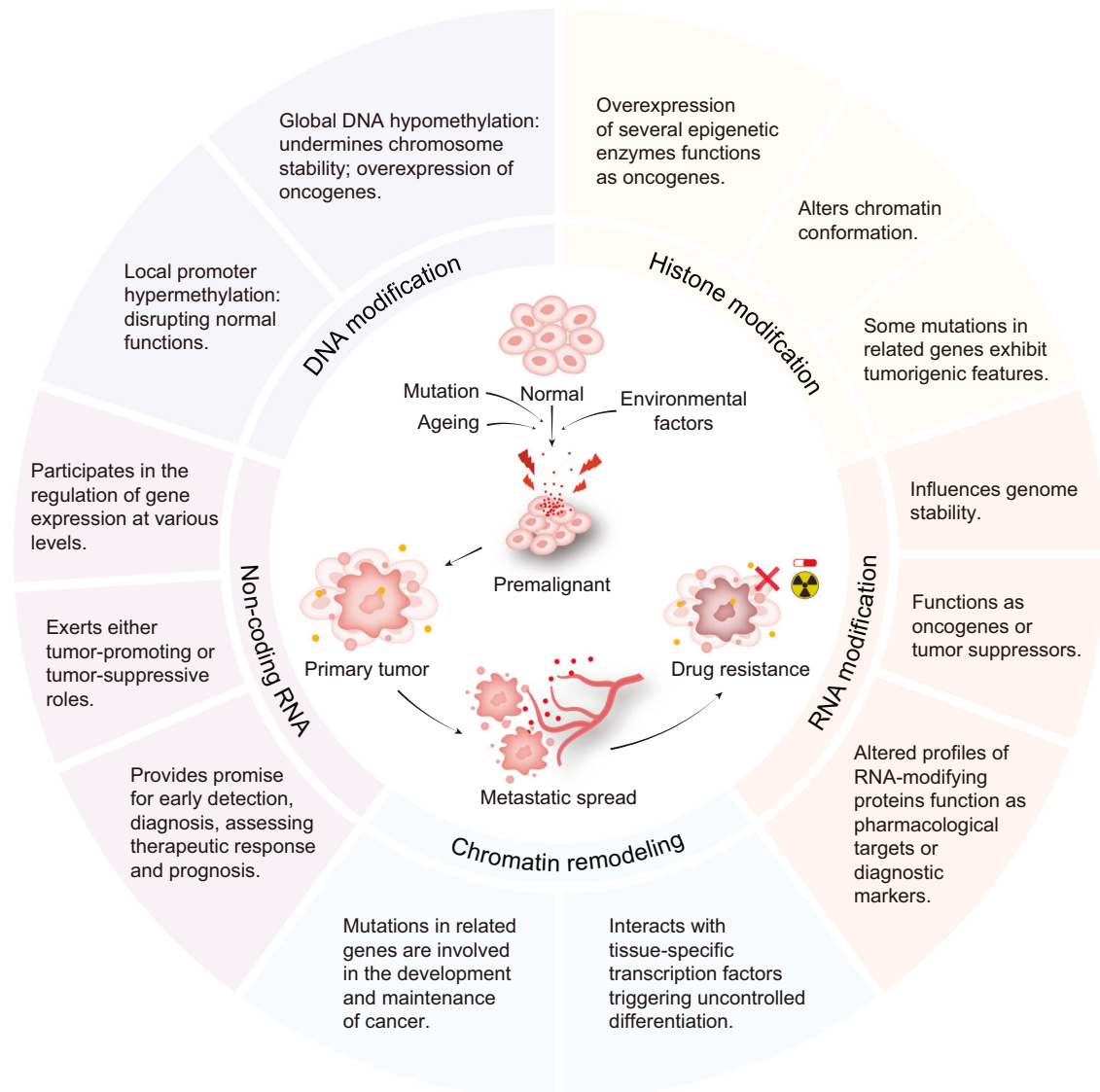


Fig. 3 Epigenetic mechanisms in cancer. Epigenetic alterations in cancer cells affect various cellular responses, such as cell proliferation, invasion, apoptosis, and drug resistance. These modifications, which include DNA modification, histone modification, RNA modification, chromatin remodeling, and non-coding RNAs, significantly affect the pathogenesis and progression of tumors. By targeting these epigenetic mechanisms, novel therapeutic strategies for combating cancer can be developed. The primary roles of epigenetic mechanisms in tumorigenesis and their further development are presented in the illustrator

mechanisms, such as histone modifications and chromatin remodeling, are also strongly linked to aging. A deficiency in sirtuin 7 (SIRT7) and histone methylation patterns like H3K9me2 and H3K27me3, for example, can activate the cyclic guanosine monophosphate-adenosine monophosphate synthase (cGAS)-stimulator of interferon genes pathway, a well-recognized aging-associated signaling pathway, thus exacerbating the aging process.^{48,49} Moreover, the importance of RNA modifications—particularly m6A and m5C—and ncRNA regulation is increasingly studied in aging research.^{50–52}

Epigenetics and cancer

Abnormal epigenetic mechanisms play crucial roles at various stages of tumor development, including initiation, progression, invasion, migration, and the development of chemotherapy resistance (Fig. 3). DNA methylation was the first discovered epigenetic mechanism associated with tumors, initially implicated in the hypermethylation of specific gene promoter regions, which

drives tumor development by silencing gene transcription.⁵³ This silencing leads to the dysfunction of critical genes such as tumor suppressor and DNA repair genes, disrupting normal cell proliferation and differentiation and fostering the malignant phenotype of tumor cells.^{54,55} Moreover, methylation loss at specific sites in tumor cell genomes, particularly in oncogene promoter regions, and extensive demethylation in DNA repeat sequences, undermines chromosome stability, facilitating tumor development.^{56,57} Changes in histone modifications are also prevalent in tumors. The roles of histone methylation and acetylation in tumor progression have been extensively explored, with numerous reviews summarizing therapeutic strategies targeting these histone modifications or their associated epigenetic-modifying enzymes, underscoring their pathological significance and therapeutic potential.^{58,59} Noticeably, bromodomain (BD) and extra-terminal (BET) family member proteins, including BRD2, BRD3, BRD4, and BRDT, serving as interpreters of histone acetylation modification, have recently been found to facilitate tumorigenesis when overexpressed.^{60,61} Upregulated BET

proteins can function as oncogenic transcriptional factors in tissue cells, driving a unique transcriptional program and controlling cell phenotype.^{62,63} Therefore, potent inhibitors targeting BET proteins may be considered potential agents for tumor treatment. Research into metabolic reprogramming and the Warburg effect in tumor cells has recently highlighted histone lactylation's function in pathological processes.^{64,65} Histone lactylation, induced by glycolysis, has been studied extensively in various malignancies such as endometrial cancer, pancreatic ductal adenocarcinoma, and glioblastoma, where it plays roles in tumor progression and the suppression of the immune microenvironment.^{66–68} Additionally, dysregulation in RNA modifications, particularly m6A, is linked to the malignant potential and resistance of tumor cells,^{69,70} affecting multiple pathways that ensure tumor cell survival, including the maintenance of stemness,⁷¹ the establishment of vascular networks,⁷² and the formation of an immunosuppressive tumor microenvironment (TME).⁷³ Thus, targeting aberrant RNA modifications could effectively disrupt the survival mechanisms of tumor cells, offering new avenues for cancer treatment.⁷⁴ Changes in ncRNA families have also been observed in various tumors, first noted in chronic lymphocytic leukemia with chromosome 13q14 deletion, characterized by decreased levels of miR-15 and miR-16.⁷⁵ Among these, various ncRNA molecules with antitumor effects have been identified and are commonly suppressed in various tumor diseases, representing promising targets for therapeutic intervention.⁷⁶ Furthermore, ncRNAs can participate in the post-translational regulation of other epigenetic-modifying enzymes, integrating into broader epigenetic networks.⁷⁷ In addition to functioning as pathogenic triggers in different tumors, ncRNAs present in extracellular vesicles in the TME also hold promise for assessing therapeutic response.^{78–80} According to clinical data from well-organized observational studies, unique plasma exosomal miRNA profiles are associated with predicting the efficacy of antitumor therapies in various tumor diseases, such as advanced non-small cell lung cancer,⁸¹ colorectal cancer,^{82,83} and breast cancer.⁸⁴

Epigenetics and metabolic syndrome and related disorders

Metabolic syndrome encompasses a constellation of pathological conditions characterized by abnormal aggregation of metabolic components, notably abdominal obesity or overweight, dyslipidemia, insulin resistance and/or glucose tolerance abnormalities, and hypertension.⁸⁵ These metabolic dysfunctions significantly elevate the risk of developing diseases such as type 2 diabetes mellitus (T2DM), nonalcoholic fatty liver disease (NAFLD), and cardiovascular diseases.⁸⁶ Epigenetic modifications play a crucial role in nutrient metabolism under physiological conditions and also bridge the genetic and environmental factors contributing to metabolic disorders.⁸⁷ For instance, dietary patterns significantly influence epigenetic markers; studies have shown that a high-fat diet in mice leads to hypermethylation in the promoter regions of genes like Rac family small guanosine triphosphate hydrolase (GTPase) 1, which promotes the progression of diabetic retinopathy.⁸⁸ Dietary-induced epigenetic changes can impact subsequent generations, increasing their risk of glucose intolerance and diabetes.⁸⁹ Moreover, epigenetic alterations linked to diet are implicated in developing gout and NAFLD.^{90,91}

Additionally, the activity of epigenetics-modifying enzymes and their cofactors, such as TET and α -ketoglutarate (α -KG) from the tricarboxylic acid cycle (TCA), can be influenced by abnormal metabolite levels in patients with metabolic diseases, further disrupting epigenetic regulation and exacerbating disease progression.⁹² Epigenetic markers, especially DNA methylation landscapes, also provide diagnostic tools;⁹³ in T2DM, for example, differential methylation in genes such as thioredoxin interacting protein, adenosine triphosphate (ATP)-binding cassette subfamily G member 1, peroxisome proliferator-activated receptor gamma-coactivator 1 alpha, and protein tyrosine phosphatase receptor

type N2, can elucidate pathophysiological mechanisms.⁹⁴ Understanding these epigenetic mechanisms in metabolic diseases is thus pivotal for developing innovative prevention, diagnosis, and treatment strategies.

Epigenetics and immune system disease

Epigenetic modifications are integral to the development and differentiation of immune cells and the regulation of immune functions. These modifications influence the differentiation of functional B and T cell subpopulations and maintain the homeostasis of innate immune cells by controlling specific gene expressions.^{95–97} Epigenetic dysregulation is closely linked to immune system diseases, including allergic reactions and autoimmune diseases, which have been extensively studied.^{98,99} For instance, allergic bronchial asthma involves reduced TET2 expression in regulatory T cells, leading to hypermethylation in the promoter region of forkhead box protein P3 and impaired immune function in controlling inflammatory responses.¹⁰⁰ Additionally, low expression of METTL3 in monocyte-derived macrophages in allergy patients exacerbates airway inflammation through M2 macrophage polarization.⁹⁷ Histone modification also plays a critical role in sustaining the therapeutic effects of glucocorticoids in asthma; oxidative stress in severe asthma cases leads to reduced histone deacetylase (HDAC) levels in alveolar macrophages, contributing to glucocorticoid resistance.¹⁰¹ Consequently, elevating HDAC levels in patients with severe, steroid-insensitive asthma could be a viable strategy to reduce airway hyperresponsiveness and restore steroid sensitivity.¹⁰²

Epigenetic mechanisms play a significant role in the pathogenesis and progression of autoimmune diseases. For instance, hypomethylation mediated by TET2 within the promoter region of absent in melanoma 2, a critical component of the inflammasome, influences T follicular helper cell-dependent humoral immune responses in systemic lupus erythematosus (SLE).¹⁰³ Additionally, altered patterns of miRNA in serum exosomes and immune cells have been identified, promising potential as biomarkers for diagnosis and indicators of disease severity.^{104,105} Histone modifications also play a pivotal role in SLE, where the administration of HDAC inhibitors has been shown to reduce cytokine profiles and improve pathogenesis in SLE and other inflammatory conditions.¹⁰⁶ Moreover, the therapeutic potential of BET proteins in antibody-mediated diseases (e.g., SLE) has recently been evaluated. BET inhibitors alter the pro-inflammatory phenotypes of mononuclear phagocytes and impair the recruitment of dendritic cells in vitro.¹⁰⁷ Beyond SLE, epigenetic mechanisms are implicated in the progression of other autoimmune diseases such as rheumatoid arthritis,¹⁰⁸ autoimmune thyroid diseases,¹⁰⁹ multiple sclerosis,¹¹⁰ T1DM,¹¹¹ and severe aplastic anemia,¹¹² highlighting the potential for epigenetics-modifying drugs in treatment strategies.

Epigenetics and neurodegenerative disease

Epigenetic modifications significantly influence learning, memory, and cognition, which are essential in maintaining synaptic plasticity.^{113,114} Disruptions in epigenetic regulation lead to the abnormal expression of genes involved in protein aggregation, neuroinflammation, and neuronal apoptosis, contributing to the pathogenesis of neurodegenerative diseases such as Alzheimer's disease (AD), Parkinson's disease (PD), and Huntington's disease (HD).¹¹⁵ The deposits of extracellular A β plaques and tau phosphorylation, as well as the loss of plasticity, are basic pathogenesis of AD. In AD, aberrant histone modification patterns, particularly histone acetylation, have been observed in hippocampal neurons of AD mouse models, potentially driving cognitive decline and inadequate removal of A β plaques.¹¹⁶ Lactylation modifications of histones H4K12 and H3K18 affect the metabolic activity of various glial cells, influencing the progression of the AD phenotype.^{117,118} In addition, aberrant DNA-methylation

patterns in the promoter regions of functional genes are linked to the accumulation of toxic peptides and the development of memory deficiency.^{119,120} Recent studies also consider RNA modifications and ncRNAs as potential therapeutic targets and diagnostic biomarkers for AD.^{121,122} PD is characterized by the misfolding and aggregation of α -synuclein, leading to the formation of Lewy bodies. Altered DNA methylation patterns have been observed in brain and blood samples from individuals with PD.^{123,124} TET2 may play a critical pathogenic role in PD, where its inactivation has shown a neuroprotective effect on dopaminergic neurons.¹²⁵ Histone acetylation dysregulation is extensively studied in PD, associated with the accumulation of phosphorylated α -synuclein and mitochondrial respiratory dysfunction.^{126,127} Dysregulation in ncRNAs, particularly long non-coding RNAs (lncRNAs) and miRNAs affects the mRNA levels of pathogenic factors post-transcriptionally and is linked with clinical symptoms such as non-motor symptoms, cognitive deficits, and inflammation, presenting potential targets for PD treatment.¹²⁸ In HD, epigenetic modification alterations are vital markers of its pathogenesis. Studies have shown the positive effects of using DNMT inhibitors, HDAC inhibitors, and extracellular vesicles loaded with miRNAs in preventing mutant huntingtin-induced neurotoxicity, emphasizing the potential roles of epigenetic dysregulations in HD.^{129–131} Recently, the impact of aberrant m6A RNA methylation on the progression of HD has been increasingly recognized. Hyper-methylation of m6A in genes related to HD and synaptic function has been linked to memory deficits. Conversely, inhibition of the fat mass and obesity associated protein (FTO) in the hippocampal regions of HD mouse models has shown promise in reversing cognitive symptoms, suggesting a potential therapeutic target.¹³²

In summary, the dynamic nature of epigenetic modifications plays a crucial role in maintaining physiological functions and life cycle processes. During embryonic development, precise epigenetic regulation is crucial to cell differentiation and ensures proper tissue specialization by activating or suppressing specific genes. Furthermore, epigenetic modifications are closely linked to an individual's adaptation to environmental influences such as nutritional status, stress, and toxin exposure, which can alter epigenetic landscapes and impact health and disease risk. On the other hand, understanding epigenetics offers a new perspective for disease prevention and treatment. The development and progression of many diseases, including cancer, metabolic disorders, immune system diseases, and neurodegenerative disorders, are closely associated with aberrant epigenetic modifications. A deeper understanding of epigenetic modulators could lead to novel therapeutic strategies, laying the groundwork for drug interventions targeting epigenetic processes.

EPIGENETICS-TARGETED DRUGS APPROVED FOR CLINICAL USE

Epigenetic modifications and the enzymes involved can either activate or suppress the expression of specific genes at different levels (Table 1). Therefore, in contrast to traditional therapies, drugs targeting epigenetic-modifying enzymes have been developed with a focus on gene regulation. This unique mechanism provides epigenetic-targeted drugs with an advantage over other traditional treatments, especially for the treatment of tumors. More specifically, epigenetics-targeted drugs specifically target the abnormal epigenetic hallmarks of cancer cells, restoring their normal cellular function or enhancing the immune system's recognition of tumor cells.^{133,134} Compared to traditional radiotherapy and chemotherapy, which directly kill cancer cells or prevent their proliferation, or immunotherapy, which activates or enhances the patient's own immune system, the administration of epigenetic agents achieves maximized damage to tumor cells with usually fewer side effects.^{135–137} The application of these novel

agents helps to reverse the progression of drug resistance caused by altered epigenetic characteristics in traditional antitumor therapies.^{138–140} Hence, these features endow epigenetic agents with importance and possibilities as monotherapy or adjuvants in combination with other therapeutic methods.¹⁴¹ Currently, some of these drugs have been approved for marketing, primarily for cancer treatment, and they exhibit exciting clinical potential. These drugs are categorized into four main types based on their mechanisms: DNMT inhibitors, HDAC inhibitors, isocitrate dehydrogenase (IDH) inhibitors, and enhancer of zeste homolog 2 (EZH2) inhibitors (Table 2).

DNMT inhibitors

Azacitidine and decitabine, known as hypomethylating agents (HMAs), target DNMT1 and were among the first epigenetics-targeted drugs approved for clinical use. The US FDA approved azacitidine in May 2004 and decitabine in June 2006 for treating myelodysplastic syndrome (MDS).^{142,143} The clinical success of these HMAs has led to a focus on optimizing their dosing schedules and administration methods. Early studies involving azacitidine and decitabine assessed their therapeutic potential through continuous and/or frequent intravenous or subcutaneous injections, establishing standard doses and delivery methods in clinical settings.^{144–146} Recent advancements have explored reduced dosages for patients at lower risk. In recent phase II clinical trials, azacitidine or decitabine was applied for three or five consecutive days in a 28-day cycle, exhibiting satisfactory therapeutic efficacy and tolerable safety.^{147,148} Some patients who received decitabine experienced myelosuppression, and future efforts are required to take steps to avoid this.¹⁴⁷

Additionally, the development of oral formulations of azacitidine and decitabine, such as oral azacitidine (CC-486), approved in September 2020 for patients with acute myeloid leukemia (AML) who are not candidates for intensive curative therapy, has improved patient convenience and treatment adherence.¹⁴⁹ In a well-organized phase III randomized trial, the median overall survival and relapse-free survival of patients treated with CC-486 were greatly improved.¹⁴⁹ Importantly, fewer grades 3 or 4 adverse events were observed during CC-486 treatment, allowing the preservation of overall health-related quality of life.¹⁴⁹ The doses of CC-486 used in clinical settings are approximately four times higher than the standard doses administered by intravenous or subcutaneous routes due to their reduced bioavailability.¹⁵⁰ Meanwhile, the poor bioavailability of oral decitabine has led to the development of ASTX727, an oral combination of decitabine with cedazuridine. This cytidine deaminase enzyme inhibitor enhances decitabine exposure after oral administration. This combination has been approved for marketing in treating MDS and AML in some countries.^{151,152} However, in China, the therapeutic potential of CC-486 and ASTX727 for AML and MDS is still under evaluation in clinical trials (NCT05413018, NCT04102020, NCT06091267, NCT02649790). Furthermore, there has been an increased focus on the synergistic effects of HMA and traditional antitumor treatments to enhance therapeutic outcomes and prevent resistance in hematologic malignancies refractory to monotherapy. This topic is further summarized in the subsequent section on drug combination applications.

Inspired by the successful application of HMAs in hematologic malignancies, their therapeutic potential in treating solid tumors is also being explored. However, as of now, HMAs are not approved for treating solid tumors. In 2017, Linnekamp et al.¹⁵³ conducted a systematic review to illustrate the clinical and biological effects of HMAs on solid tumors based on previously completed clinical trials. The efficacy of azacitidine and decitabine in solid tumors was less pronounced than in hematological malignancies, primarily because most study participants had advanced-stage tumors with short life expectancies.¹⁵³ Moreover, many early-stage studies were small-sized cohorts, lacking

Table 1. Key examples of epigenetics-modifying enzymes that are considered targets for drug development and their major biological functions

| Epigenetic modification | Type | Epigenetics-modifying enzyme | Biological processes | Reference(s) |
|-------------------------|--------|------------------------------|---|--------------|
| DNA methylation | Writer | DNMT1 | Maintains DNA methylation after replication | 907 |
| | | DNMT2 | Binds to DNA with very weak methyltransferase activity; involved in RNA methylation | 908 |
| | | DNMT3A | Promotes the genome-wide de novo DNA methylation | 909 |
| | | DNMT3B | Promotes the genome-wide de novo DNA methylation | 910,911 |
| | | DNMT3L | Increases the methyltransferase activity of DNMT3A or DNMT3B | 912 |
| | Eraser | TET1 | Active DNA demethylation and binds to DNA via the CXXC zinc finger domain | 913 |
| | | TET2 | Active DNA demethylation and binds to DNA via the interaction with DNA binding proteins | 914 |
| | | TET3 | Active DNA demethylation and binds to DNA via the CXXC zinc finger domain | 915 |
| | Reader | MeCP1 | Preferentially binds to methylated DNA and represses transcription | 916 |
| | | MeCP2 | Binds to a single methyl-CpG pair, not influenced by sequences flanking the methyl-CpGs | 917 |
| | | MBD1 | Recruits chromatin-modifying enzymes to both methylated and unmethylated CpG islands; largely silence transcription | 918,919 |
| | | MBD2 | A transcriptional repressor or activator depending on the cellular context | 920 |
| | | MBD3 | Interacts with NuRD complex to cause transcriptional repression | 921,922 |
| | | MBD4 | Exerts DNA glycosylase activity and functions in DNA repair | 923 |
| | | UHRF1 | Negatively regulates transcription via the binds to 5hmC and 5mC on DNA, as well as H3K9me3, and H3R2me0; recruits DNMT1 to methylate DNA | 356,358 |
| | | UHRF2 | Allosterically activated by 5hmC and participates in DNA demethylation during neuronal commitment | 924–926 |
| Histone acetylation | Writer | HAT1 (KAT1) | Acetylates H4K12/K5 predominantly; has less activity for H2A | 927–929 |
| | | GCN5 (KAT2A) | Acetylates H3 and H4 and its primary sites are H3K14 | 930 |
| | | PCAF (KAT2B) | Acetylates H3 and H4 predominantly and its primary sites are H3K14; has less activity for H2A and H4B | 387 |
| | | CBP/P300 (KAT3A/KAT3B) | Acetylates H2A, H2B, H3, H4 and its primary sites are H3K14/K18/K27 | 931,932 |
| | Eraser | HDAC1 | Removes acetylated modifications from H1, H2A, H2B type 1/2 and H3 | 933 |
| | | HDAC2 | Removes acetylated modifications from H1, H2A, H2B type 1/2 and H3 | 933 |
| | | HDAC3 | Removes acetylated modifications from H2BK12/K15/K16 | 933 |
| | | HDAC4 | A very weak deacetylase activity on histone | 934 |
| | | HDAC9 | A very weak deacetylase activity on histone | 935 |
| | | SIRT1 | Removes acetylated modifications from H1K2, H3K9, and H4K16 | 936,937 |
| | | SIRT2 | Removes acetylated modifications from histones during G2/M transition and mitosis | 938,939 |
| | | SIRT6 | Removes acetylated modifications from H3K9 and H3K56 | 940,941 |
| | | SIRT7 | Removes acetylated modifications from H3K18 | 942 |
| | Reader | BRD2 | Recognizes H4K12ac preferentially | 943 |
| | | BRD4 | Recognizes H3K27ac preferentially | 944,945 |
| | | ENL | Recognizes H3K9/K18/27ac preferentially | 946 |
| | | AF9 | Recognizes H3K9ac preferentially and H3K27/K18ac to a lesser extent | 946 |
| | | YEATS2 | Recognizes H3K9ac preferentially; functions as a selective histone crotonylation reader | 947,948 |
| | | GAS41 | Recognizes H3K18/K27ac preferentially; binds to H3K14 | 949,950 |
| Histone methylation | Writer | EZH2 | Catalyzes mono-, di-, and tri-methylation of H3K27 and H3K9, as well as non-histones substrates | 951 |
| | | DOT1L | Catalyzes mono-, di-, and tri-methylation of H3K79 | 952 |
| | | SETDB1 | Catalyzes trimethylation of H3K9 | 953 |
| | | GLP/G9a | Catalyzes mono- or di-methylation of H3K9 and non-histone substrates | 954,955 |
| | | SMYD2 | Catalyzes both trimethylation of H3K36 and non-histone substrates | 956,957 |
| | | NSD | Catalyzes the dimethylation of H3K36 | 958 |
| | | PRMT1 | The member of type I PRMTs; the dominant enzyme catalyzing asymmetric dimethylarginine production in proteins and mainly functions as a transcriptional activator | 959 |

Table 1. continued

| Epigenetic modification | Type | Epigenetics-modifying enzyme | Biological processes | Reference(s) |
|-------------------------|--------|------------------------------|--|--------------|
| RNA methylation | Eraser | PRMT5 | The member of type II PRMTs; functions as a transcriptional suppressor or coactivator depending on the cellular context | 960,961 |
| | | LSD1(KDM1A) | Removes methylation modifications at H3K4 and H3K9; acts as a coactivator or a corepressor depending on the cellular context | 962 |
| | | KDM2 | Removes methylation modifications at H3K4; H3K9, and H3K36; stimulates and inhibits gene transcription | 963 |
| | | KDM7 | Removes mono- and di-methylated modifications on H3K9 and H3K27 | 964 |
| | | KDM3 | Removes mono- and di-methylated modifications from lysine H3K9 | 965 |
| | | KDM4 | Removes methylated modifications from H3K9 and H3K36 | 965,966 |
| | | KDM5 | Removes di- and tri-methylated modifications from H3K4 | 967 |
| | | KDM6A | An X-linked protein removing methylated modifications from H3K27 | 968 |
| | | KDM6B | Removes trimethylated modifications from H3K27 | 969 |
| | Reader | MBT | Recognizes lysine residues on H3 and H4, and helps form monomethylated, dimethylated, or trimethylated modifications | 970 |
| | | Chromodomain | Recognizes dimethylated lysine residues of H3K9 and H3K27 | 971,972 |
| | | Tudor | Recognizes methylated lysine and arginine residues on histones H3 and H4 | 973 |
| | | PWWP | Recognizes H3K36me2/3; binds to dsDNA in a non-specific manner | 974,975 |
| | | PHD | Recognizes H3K4me2/3/0, H3K14ac or H3K27me0 to a lesser extent | 976 |
| | | WDR | Recognizes lysine and arginine methylation of H3 | 977 |
| | writer | METTL3 | Catalyzes m6A methylation | 978 |
| | | METTL14 | Binds to METTL3 and enhances the catalytic activity of METTL3 | 979 |
| | Eraser | FTO | RNA m6A demethylation; regulates RNA splicing | 980,981 |
| | | ALKBH5 | RNA m6A demethylation; regulates RNA metabolism and export | 982 |
| Chromatin remodeling | Reader | ALKBH3 | Removes the methyl group at the m6A from tRNA; functions as an m1A demethylase | 983 |
| | | YTHDF1 | Responsible for mRNA translation | 984 |
| | | YTHDF2 | Responsible for mRNA degradation | 984,985 |
| | | IGF2BP | Regulates mRNA translation | 714 |
| | | SMARCA2 | DNA-stimulated ATPase in the SWI/SNF complex | 986 |
| | Mover | SMARCA4 | DNA-stimulated ATPase in the SWI/SNF complex; binds to acetylated peptides on H3 and H4 | 986,987 |
| | | Polybromo-1 | Recognizes H3K14ac preferentially | 988 |
| | Reader | BRD7 | Recognizes acetylated modifications on histones and non-histones substrates | 989,990 |
| | | BRD9 | Recognizes butyryllysine, and crotonyllysine histone peptide modifications | 991 |
| | | | | |

AF9 acute lymphocytic leukemia 1 (ALL1)-fused gene from chromosome 9 protein, *ALKBH* ALKB homolog, *ARID* AT-rich interactive domain, *ATPase* adenosine triphosphate hydrolase, *BAF* BRG1-associated factor, *BRD* bromodomain-containing protein, *CBP/P300* cyclic adenosine monophosphate-responsive element-binding protein (CREB)-binding protein/histone acetyltransferase P300, DNMT DNA methyltransferase, *DOT1L* disruptor of telomeric silencing-1-like, *dsDNA* double-stranded DNA, *ENL* eleven-nineteen leukemia, *EZH2* enhancer of zeste homolog 2, *FTO* fat mass and obesity associated protein, *GAS41* glioma amplified sequence 41, *GCN5* general control non-depressible 5, *GLP* G9a-like protein, *HAT* histone acetyltransferase, *HDAC* histone deacetylase, *H4K12ac* histone 3 lysine 12 acetylation, *H3K9me3* histone 3 lysine 9 trimethylation, *IGF2BP* insulin-like growth factor 2 mRNA-binding protein, *KAT* lysine acetyltransferase, *LSD1(KDM1A)* lysine-specific demethylase 1, *m6A* N6-methyladenosine, *MBD* methyl-CpG binding domain protein, *MBT* malignant brain tumor, *MeCP* methyl-CpG-binding protein, *METTL* methyltransferase-like, *NSD* nuclear receptor binding SET domain protein, *NuRD* nucleosome remodeling and deacetylase, *PBAF* polybromo, brahma-related gene 1-associated factor, *PCAF* P300/CBP associated factor, *PHD* plant homeodomain, *PRMT* protein arginine methyltransferase, *PWWP* proline-tryptophan-tryptophan-proline, *SETDB1* SET domain bifurcated histone lysine methyltransferase 1, *SIRT* sirtuin, *SMARCA2* SWI/SNF-related, matrix-associated, actin-dependent regulator of chromatin A2, *SMYD2* SET and MYND domain containing 2, *SWI/SNF* Switch/Sucrose nonfermentable chromatin-modifying complex, *TET* ten-eleven translocation, *UHRF1* ubiquitin-like with PHD and RING finger domains 1, *WDR* WD40 repeat, *YEATS2* YAF9, eleven-nineteen-leukemia protein (ENL), acute lymphocytic leukemia 1-fused gene from chromosome 9 protein (AF9), TAF14, and SAS5 (YEATS) domain-containing 2, *YTHDC1* YTH domain-containing protein 1, *YTHDF1* YTH domain family protein 1, *5hmC* 5-hydroxymethylcytosine, *5mC* 5-methylcytosine

sufficient evidence to generalize therapeutic effects across different tumor types. With significant advances in optimizing HMA formulations and dosages, as well as the increasing number of combination therapies showing promising effects on solid tumors in vitro and in vivo, clinical trials of HMAs, particularly the oral formulations CC-486 and ASTX727, among patients with solid tumors, are being extensively conducted, and their results are eagerly anticipated.^{154–156}

HDAC inhibitors

Over the past two decades, substantial progress has been made in developing HDAC inhibitors, with six approved for clinical use. These include vorinostat (SAHA), romidepsin (FK228), belinostat (PXD101), and panobinostat (LBH589, although the FDA canceled it in 2022). These drugs have been approved by various regulatory bodies, such as the US FDA, the European Medicines Agency, and the Pharmaceuticals and Medical Devices Agency (PMDA). They

Table 2. Broad indications and common treatment-related adverse events of marketed epigenetic-targeting drugs

| Drug(s) | Target(s) | Route(s) of administration | FDA approval | EMA/NMPA approvals | Broad indications | Common grades 3 or worse treatment-related adverse effects reported in clinical trials | Reference(s) |
|-----------------------------|----------------|------------------------------------|-----------------------------|--------------------|--|---|---------------|
| Azacitidine | DNMT1 | Intravenous/ subcutaneous | Yes | — | Juvenile myelomonocytic leukemia AML; CMML; MDS | Thrombocytopenia, neutropenia, anemia, sepsis, infection, and pneumonia | 992–994 |
| | | Subcutaneous | Not approved yet | EMA | | | |
| | | Intravenous | Not approved yet | NMPA | AML; MDS; Philadelphia chromosome positive CML | | |
| Oral azacitidine | DNMT1 | Oral | Yes | — | AML | Febrile neutropenia, thrombocytopenia, leukopenia, pneumonia, respiratory failure, bacteraemia, and sepsis | 149,995 |
| Decitabine | DNMT1 | Intravenous | Yes | NMPA | MDS | Febrile neutropenia, thrombocytopenia, anemia, pneumonia, and infection | 996–998 |
| | | Intravenous | Yes | — | CMML; Refractory anemia (with/without) excess blasts | | |
| | | Intravenous | Not approved yet | EMA | AML | | |
| Decitabine/ Cedazuridine | DNMT1, CDA | Oral | Yes | — | MDS | Thrombocytopenia, febrile neutropenia, pneumonia, respiratory failure, bacteraemia, and sepsis | 831,997,999 |
| | | Oral | Not approved yet | EMA | AML | | |
| Vorinostat | HDACs | Oral | Yes | — | CTCL | Cellulitis, pulmonary embolism, sepsis, anorexia, increased creatinine phosphokinase, rash, and thrombocytopenia | 1000,1001 |
| Romidepsin | HDACs | Intravenous | Yes | — | CTCL; PTCL | Lymphopenia, neutropenia, leukopenia, thrombocytopenia, infections, and tumor lysis syndrome | 1002–1004 |
| Belinostat | HDACs | Intravenous | Yes | — | PTCL | Anemia, thrombocytopenia, dyspnea, neutropenia, infections, tumor lysis syndrome, and ventricular fibrillation | 180,1005,1006 |
| Panobinostat | HDACs | Oral | Canceled by the FDA in 2022 | EMA | MDS | QTc prolongation, hemorrhage, thrombocytopenia, lymphopenia, and asthenia | 1007,1008 |
| Chidamide | Class I HDAC | Oral | Not approved yet | NMPA | Breast cancer; DLBCL; PTCL | Neutropenia, thrombocytopenia, anemia, leukopenia, diarrhea, and mucositis | 202,1009,1010 |
| Givinostat | HDAC1, HDAC3 | Oral | Yes | — | DMD | Diarrhea | 207 |
| Enasidenib | IDH2 | Oral | Yes | — | AML | Febrile neutropenia, IDH differentiation syndrome, and indirect hyperbilirubinemia | 228,234 |
| Ivosidenib | IDH1 | Oral | Yes | NMPA | AML | QT interval prolongation, IDH differentiation syndrome, anemia, and ascites | 1011,1012 |
| | | Oral | Yes | — | Cholangiocarcinoma; MDS | | |
| | | Oral | Not approved yet | EMA | IDH1-mutated AML; IDH1-mutated cholangiocarcinoma | | |
| Ivosidenib/ Azacitidine | IDH1/ DNMT1 | Oral; intravenous/ subcutaneous | Yes | — | IDH1-mutated AML | Febrile neutropenia, neutropenia, bleeding events, infection, IDH differentiation syndrome, and QT interval prolongation | 240,241 |
| Olutasidenib | IDH1 | Oral | Yes | — | IDH1-mutated AML | Thrombocytopenia, febrile neutropenia, anemia, alanine aminotransferase increased, and aspartate aminotransferase increased | 248,249 |

| Table 2. continued | | | | | |
|---|-----------|----------------------------|------------------|-------------------------------------|-------------------|
| Drug(s) | Target(s) | Route(s) of administration | FDA approval | EMA/NMPA approvals | Broad indications |
| Tazemetostat | EZH2 | Oral | Yes | — | FL; Sarcoma |
| Valemetostat tosilate | EZH2/EZH1 | Oral | Not approved yet | *Only approved by the PMDA in Japan | ATL |
| Common grades 3 or worse treatment-related adverse effects reported in clinical trials | | | | | |
| Hyperglycemia, hyponatremia, anemia, thrombocytopenia, neutropenia, lymphopenia, and weight loss | | | | | |
| Thrombocytopenia, anemia, lymphopenia, leukopenia, and neutropenia | | | | | |
| AML acute myeloid leukemia, ATL adult T-cell leukemia/lymphoma, CDA cytidine deaminase, CML chronic myelogenous leukemia, CMML chronic myelomonocytic leukemia, CTCL cutaneous T-cell lymphoma, DLBCL diffuse large B-cell lymphoma, DMD Duchenne muscular dystrophy, DNMT1 DNA methyltransferase 1, EMA European Medicines Agency, EZH2 enhancer of zeste homolog 2, FDA Food and Drug Administration, FL follicular lymphoma, HDAC histone deacetylase, IDH isocitrate dehydrogenase, MDS myelodysplastic syndrome, NMPA National Medical Products Administration, PMDA Pharmaceuticals and Medical Devices Agency, PTCL peripheral T-cell lymphoma | | | | | |

are used to treat conditions such as multiple myeloma (MM), cutaneous T-cell lymphoma (CTCL), and peripheral T-cell lymphoma (PTCL).^{157,158} Additionally, chidamide (tucidinostat) was approved by PMDA in Japan and National Medical Products Administration (NMPA) in China for the treatment of PTCL and advanced breast cancer,^{159,160} and more recently, givinostat (ITF2357) was approved by the FDA in March 2024 as the first nonsteroidal treatment for Duchenne muscular dystrophy (DMD) for patients aged six years and older.¹⁶¹

Vorinostat was the first pan-inhibitor of HDACs approved by the FDA in October 2006 for CTCL.¹⁶² Soon after, in July 2011, it was also approved for clinical therapy by PMDA. In addition to CTCL, the application of vorinostat to AML, MM, malignant pleural mesothelioma, newly diagnosed high-grade glioma (NCT01236560), and advanced non-small cell lung cancer (NCT00473889) therapy has entered phase III clinical trials.^{163–166} Disappointingly, though vorinostat exerts effective therapeutic effects in diverse hematological malignancies, limited efficacy has been observed in solid tumors.¹⁶⁶ Another thing to note when using vorinostat as a clinical medication is the potential adverse events that may occur. While generally mild, adverse events related to vorinostat can include thrombosis, QT interval prolongation, and potentially fatal ventricular tachycardia or torsional tachycardia.^{167–169} These findings have driven the development of other HDAC inhibitors, with the expectation of elevated safety and efficacy in vivo.

Romidepsin, another pan-HDAC inhibitor, was approved by the FDA in November 2009 for CTCL and later for PTCL. It has shown a higher affinity to class I HDAC proteins.¹⁷⁰ Subsequently, it was approved for treating patients with PTCL by the FDA and PMDA. Phase III randomized controlled trials are performed to evaluate the therapeutic value of the first-line treatment of PTCL, referring to the combination of cyclophosphamide, doxorubicin, vincristine, prednisone (CHOP), and romidepsin plus CHOP in patients with PTCL, while the addition of romidepsin failed to increase efficacy as expected.^{171,172} However, after a six-year follow-up, the application of romidepsin shows beneficial effects in prolonging median progression-free survival.¹⁷¹ In addition, the combination of romidepsin with other therapeutics, such as oral 5-azacytidine, tenalisib (an inhibitor of phosphoinositide-3-kinase and salt-inducible-kinase-3), and lenalidomide (a new generation of immune modulator) shows promising therapeutic potential in various types of T-cell lymphoma in the initial stages of clinical practice, supporting further exploration.^{173–176} Meanwhile, investigations on the therapeutic effects of romidepsin against other diseases are ongoing, particularly in antiretroviral treatment in human immunodeficiency virus-1 (HIV-1) infection.^{177,178}

Belinostat was FDA-approved in July 2014 for relapsed or refractory (R/R) PTCL, showing pan-inhibitory effects on HDAC proteins.¹⁷⁹ Common adverse effects of belinostat include nausea, vomiting, diarrhea, dysgeusia, fatigue, and severe hematologic treatment-related adverse events.^{180,181} Further, dosing considerations are needed for patients with hepatic impairment due to liver metabolism.¹⁸² Belinostat is being explored for other myeloid malignancies and solid tumors, including glioblastoma and small-cell lung cancer.^{183–186}

Panobinostat, an oral broad-spectrum HDAC inhibitor approved in January 2015 for R/R MM in combination with dexamethasone and bortezomib, showed a slight overall survival benefit in phase II and III trials.¹⁸⁷ However, many participants experienced adverse events like thrombocytopenia, lymphopenia, asthenia, and fatigue, which raises concerns about its tolerability.^{187–189} In a recent randomized phase II clinical trial, it was proposed that administering bortezomib via subcutaneous application rather than intravenous injection could improve the safety and tolerability of the triplet regimen, including panobinostat.¹⁹⁰ Beyond its primary indications, panobinostat is being explored for its efficacy in various other tumor diseases such as lymphoma, primary

myelofibrosis, glioma, clear cell renal cell carcinoma, and prostate cancer, both as monotherapy and in combination with other tumor therapeutics, showing promising efficacy across multiple malignancies.^{191–195} However, the safety of panobinostat continues to be a primary concern and requires further evaluation in advanced clinical studies.

Chidamide, the first orally administered selective HDAC inhibitor targeting HDAC1, HDAC2, HDAC3, and HDAC10,¹⁶⁰ is currently under investigation for a variety of solid and hematological malignancies, autoimmune diseases, and neurodegenerative diseases.^{196–201} It offers advantages over pan-inhibitors in the treatment of tumor diseases and in minimizing severe adverse effects.¹⁶⁰ Recent therapeutic strategies using chidamide in combination with a second antitumor intervention have shown promising prospects. For instance, a combination of chidamide and exemestane has proven effective as a neoadjuvant treatment for patients with stage II–III breast cancer that is hormone receptor-positive and human epidermal growth factor receptor 2-negative.^{199,202} A recent phase III clinical trial reported an increased occurrence of grades 3–4 hematological adverse events in the tucidostat plus exemestane group, while the median progression-free survival of these patients was notably improved.²⁰² Furthermore, synergistic effects have been observed when chidamide is used in conjunction with immunotherapy, endocrine therapy, or chemoradiation, offering novel adjuvant approaches for tumor therapy.^{203–206}

Givinostat, developed by Italfarmaco SpA, is a potent inhibitor of HDAC1 and HDAC3 recently approved for clinical use. In a pivotal, multicenter, randomized phase III clinical trial involving 179 patients with DMD aged at least six years, givinostat effectively delayed disease progression.²⁰⁷ The most common adverse events reported were diarrhea and vomiting.²⁰⁷ Additionally, givinostat shows promise as a treatment for polycythemia vera, particularly in patients unresponsive to hydroxycarbamide monotherapy.²⁰⁸ In phase I/II clinical trials, givinostat demonstrated promising efficacy and tolerability in patients with polycythemia vera.^{209,210} Subsequently, long-term follow-up over four years has further substantiated the therapeutic benefits and safety profile of givinostat.²¹¹ Throughout the follow-up period, the overall response rate consistently exceeded 80% among patients with PV, while only 10% of these patients experienced grade 3 treatment-related adverse events, suggesting its potential for prolonged clinical use.²¹¹

IDH inhibitors

IDH is a key enzyme in the TCA cycle that normally catalyzes the conversion of isocitrate to α -KG and carbon dioxide. In cells with mutated IDH, this enzyme instead produces 2-hydroxyglutarate (2-HG), a metabolite that inhibits α -KG-dependent epigenetic enzymes and contributes to the aberrant epigenetic landscape seen in various diseases, particularly tumors.^{212,213} Currently, three IDH inhibitors are approved for clinical use: enasidenib (AG-221), ivosidenib (AG-120), and olutasidenib (FT-2102), targeting different forms of the enzyme mutation.^{214–217} Additional IDH1/2 inhibitors that are allowed to be investigated in clinical practice include the dual inhibitor of mutant IDH1/2 vorasidenib (AG-881),²¹⁸ the irreversible mutant IDH1 inhibitor LY3410738 (NCT06181045, NCT06181084), and the pan-mutant-IDH1 inhibitor BAY1436032.^{219,220}

Enasidenib is an allosteric inhibitor targeting mutated IDH2, approved by the FDA in August 2017 for the treatment of R/R AML with IDH2 mutations.²²¹ Based on preliminary animal experiments and preclinical evidence, enasidenib effectively reduces the 2-HG levels derived from IDH2 mutations, reversing excessive histone and DNA methylation landscapes.^{222–224} Subsequently, enasidenib has entered clinical trials and demonstrated good efficacy in treating AML and MDS patients, which is further considered a promising option for elderly AML patients over 60 years old,

especially those who are not suitable for intensified chemotherapy.^{225–230} Combination therapy with enasidenib and azacitidine has shown acceptable tolerability and potential to improve outcomes for patients with IDH-mutated AML.^{229,231,232} However, potential severe adverse effects include hyperbilirubinemia, thrombocytopenia, pneumonia, and IDH differentiation syndrome, the latter of which can be life-threatening and requires careful management.^{233–235} Noticeably, IDH differentiation syndrome is one of the potentially lethal entities that require prompt recognition and more appropriate management.²³⁶ Enasidenib is also being explored for other conditions caused by IDH2 mutations, such as D-2-hydroxyglutaric aciduria type II,²³⁷ chondrosarcoma,²³⁸ angioimmunoblastic T-cell lymphoma (NCT02273739), and malignant sinonasal or skull base tumors (NCT06176989).

Ivosidenib, targeting mutated IDH1, was first approved by the FDA in July 2018 for R/R AML.²³⁹ In April 2022, with the data from a completed phase III clinical trial being made public, the therapeutic potentials and good safety of the combination of azacitidine and ivosidenib among patients with AML received broader attention.²⁴⁰ Subsequently, the FDA approved this regimen for elderly patients with newly diagnosed IDH1-mutated AML in May 2022.²⁴¹ Besides AML, ivosidenib is approved for MDS and cholangiocarcinoma, with ongoing phase III studies in unresectable or metastatic cholangiocarcinoma with IDH1 mutations.^{216,242} The most significant adverse events include ascites and other severe conditions, necessitating vigilant monitoring.²⁴² Ivosidenib has also shown promising results in phase I clinical trials for IDH-mutated advanced glioma, with a daily dose of 500 mg proving effective in reducing 2-HG levels and controlling the disease.^{243–246}

Olutasidenib, an oral IDH1 inhibitor, was approved by the FDA in December 2022 for treating R/R AML with specific IDH1 mutations. It has also shown promise as a therapeutic option for patients with IDH1-mutated AML who are insensitive to venetoclax, offering a new avenue for treatment where previous therapies may have failed.²¹⁷ Clinical trials have demonstrated that olutasidenib, combined with azacitidine, provides comparable efficacy and tolerability in AML and MDS patients harboring mutant IDH1.²⁴⁷ Treatment-emergent side effects of grade 3–4, such as febrile neutropenia, anemia, thrombocytopenia, and neutropenia, occur at a low frequency with olutasidenib monotherapy or in combination therapies, suggesting a manageable safety profile.^{247,248} Beyond hematological malignancies, the therapeutic potential of olutasidenib is also being explored in other diseases, such as IDH-mutated R/R gliomas. In a multicenter, open-label, phase Ib/II clinical trial involving 26 patients, olutasidenib achieved a disease control rate of 48%. Notable grade 3–4 adverse events included increases in alanine aminotransferase and aspartate aminotransferase, indicating the need for careful liver function monitoring during treatment.²⁴⁹

EZH2 inhibitors

Currently, two EZH2 inhibitors, tazemetostat (EPZ-6438) and valemestostat tosilate (DS-3201, DS-3201B), targeting EZH1/2 or EZH2 have been approved and are being utilized in various therapeutic strategies.

Tazemetostat, the first oral EZH2 inhibitor, was approved by the FDA in January 2020 for patients over 16 years of age with advanced epithelioid sarcoma that is ineligible for complete resection.²⁵⁰ It is also the first targeted drug for epithelioid sarcoma treatment.²⁵¹ In a phase II clinical trial (NCT02601950), tazemetostat demonstrated good tolerability and clinical activity, with a low incidence of severe treatment-related adverse events such as anemia and weight loss.²⁵² Tazemetostat has also been studied as a monotherapy in R/R follicular lymphoma (FL), showing promising, durable responses and an acceptable safety profile.^{253,254} Common severe adverse events included

thrombocytopenia, neutropenia, and anemia.²⁵⁴ In Japan, a phase I/II trial evaluated tazemetostat 800 mg twice daily in R/R EZH2 mutation-positive FL, showing encouraging response rates and tolerability, which helped to accelerate its approval by the FDA and PMDA for this indication.^{255,256} Furthermore, tazemetostat is being investigated as a single agent for malignant mesothelioma, with ongoing efforts to refine biomarkers for its activity in malignant pleural mesothelioma.²⁵⁷ Research is also underway to assess the efficacy and tolerability of tazemetostat in combination with other therapeutic agents, including programmed cell death 1 (PD-1)/programmed cell death 1 ligand 1 (PD-L1) inhibitors, chemotherapy, and targeted therapeutics across different tumor types.^{258–260} Notably, phase III clinical trials are exploring tazemetostat in combination with lenalidomide and rituximab (NCT04224493) or doxorubicin (NCT04204941) focusing on R/R FL and epithelioid sarcoma, which are highly anticipated for their potential to redefine treatment paradigms.

Valemetostat tosilate is an innovative dual inhibitor of EZH1/2 that received approval from the PMDA in Japan for the treatment of R/R adult T-cell leukemia/lymphoma (ATL) in September 2022.²⁶¹ It is administered orally and should be taken on an empty stomach to avoid adverse food effects.²⁶² Dosage adjustments are necessary when valemetostat is used concurrently with strong inhibitors of cytochrome P450 3A and P-glycoprotein, which can affect its metabolism and excretion.²⁶³ In a multicenter phase 2 trial involving patients with R/R aggressive ATL, valemetostat demonstrated promising efficacy, even in heavily pretreated patients. The common treatment-associated adverse effects reported were manageable, including thrombocytopenia, anemia, alopecia, dysgeusia, neutropenia, lymphopenia, leukopenia, decreased appetite, and pyrexia.²⁶⁴ However, resistance to valemetostat has been observed in some patients with ATL, potentially due to acquired mutations in the polycomb repressive complex 2 (PRC2) within tumor cells, highlighting a significant challenge in long-term treatment scenarios.⁵⁹ Currently, valemetostat and its analogs are also being investigated in various preclinical studies and animal models for conditions such as tumor protein p63 gene-rearranged lymphoma, sinonasal teratocarcinoma, ibuprofen-resistant mantle cell lymphoma, and human T-cell leukemia virus type 1-associated myelopathy. These studies further expand the potential therapeutic applications of valemetostat and warrant continued exploration of valemetostat-based treatment strategies.^{265–268}

OTHER EPIGENETICS-TARGETED DRUGS UNDER RESEARCH AS CLINICAL CANDIDATES

Although there have been many advances in the research of marketed drugs, they still belong to the tip of the iceberg relative to the entire field of epigenetics-targeting drug development. Many small molecule inhibitors and agonists targeting epigenetic-modifying enzymes are being identified and progressively advancing into the early stages of clinical trials. This section categorizes and summarizes these drugs based on their mechanisms, highlighting agents that show potential therapeutic value in clinical settings (Fig. 4).

Epigenetics-targeted drugs and DNA methylation

This critical regulatory mechanism of gene expression involves adding methyl groups to DNA, primarily at cytosine bases in CpG dinucleotides, which generally leads to gene silencing. The process is dynamically regulated by DNMT and TET enzymes. The aberrant activity of these enzymes is closely linked with the pathogenesis of a wide range of diseases, not only cancers but also metabolic, inflammatory, and neurological disorders, underscoring the therapeutic relevance of targeting these pathways.^{269–272} Therefore, DNA methylation provides a promising

platform for developing epigenetics-targeted drugs in clinical practice (Table 3).

Targeting the writer of DNA methylation: DNMT. Current therapeutic strategies primarily involve DNMT inhibitors, which suppress the expression or enzymatic activities of DNMTs, thereby counteracting improper DNA methylation patterns. These inhibitors are crucial in correcting the abnormal addition of methyl groups to DNA, a common feature in many pathologies.

DNMT inhibitors: Most DNMT inhibitors under investigation are employed in treating hematological or solid tumors, with a smaller portion used for inflammatory or proliferative benign diseases.^{153,273} DNMT inhibitors fall into two principal categories based on their mechanisms of action: nucleoside DNA methylation inhibitors and non-nucleoside DNA methylation inhibitors.

Among the nucleoside analogs, the FDA-approved drugs azacitidine and decitabine are noteworthy. These compounds integrate into the DNA structure and are recognized by DNMTs during DNA replication, thereby obstructing normal DNA methylation processes.²⁷⁴ Another significant compound, guadecitabine (SGI110, an antimetabolite of decitabine), represents a second-generation DNA methylation inhibitor. It is an antinucleotide molecule that resists degradation by cytidine deaminase.²⁷⁵ Research primarily focuses on its application against various malignant tumors. However, although guadecitabine has demonstrated good tolerance and favorable outcomes in many clinical trials, there is still clinical evidence indicating that its application may cause serious treatment-related adverse effects, such as pneumonia, sepsis, aspiration pneumonia, metabolic disorders, neutropenia, leukopenia, and pruritis.^{276,277} Furthermore, a recent phase II clinical trial conducted among patients with succinate dehydrogenase-deficient tumors was terminated due to low objective response rates.²⁷⁸ Additionally, when combined with traditional antitumor agents such as chemotherapeutic drugs and immune checkpoint blockade agents, guadecitabine demonstrates a potent synergistic effect, enhancing long-term clinical benefits.^{279–282} Promisingly, its use in treating patients with AML has advanced to phase III clinical trials, indicating high response rates and comparable safety, positioning it as a promising future alternative.^{279,283} Other nucleoside DNA methylation inhibitors include CP-4200, with cellular uptake less dependent on the nucleoside transporters involved in azacytidine uptake;²⁸⁴ F-aza-T-dCyd (NSC801845), optimized structurally from T-dCyd, F-T-dCyd, and Aza-T-dCyd;²⁸⁵ DHDAC, which is less cytotoxic and more stable;²⁸⁶ NPEOC-DAC, a decitabine derivative modified at the N4 position of the azacitidine ring, displaying significantly reduced potency at low doses in inhibiting DNA methylation;²⁸⁷ and zebularine, known for its high selectivity and better biocompatibility towards pathological cells,²⁸⁸ demonstrating significant therapeutic effects not only on tumors but also on non-tumor diseases like renal fibrosis,²⁸⁹ T2DM,²⁹⁰ and NAFLD.²⁹¹ Additionally, clofarabine, an FDA-approved purine nucleoside analog for treating pediatric AML, primarily inhibits DNA biosynthesis and the ribonucleotide reductase enzyme and has shown potential in early-stage carcinogenesis through DNMT1 inhibition.²⁹²

Beyond these, various non-nucleoside DNMT inhibitors have been identified. These include specific artificially synthesized inhibitors and naturally occurring agents with a demethylation function.^{293,294} Enhancements to the physical properties of these natural compounds, such as solubility and stability, benefit the development of more effective DNMT-targeted inhibitors.^{295,296} Non-nucleoside DNMT inhibitors are categorized based on their diverse mechanisms of action into competitors of S-adenosylmethionine (SAM), competitive or non-competitive inhibitors of DNMT, regulators of DNMT expression, and binders of DNA substrates.⁵³ MG98 and hydralazine are the only two drugs currently in clinical trials. MG98, an antisense oligodeoxynucleotide,

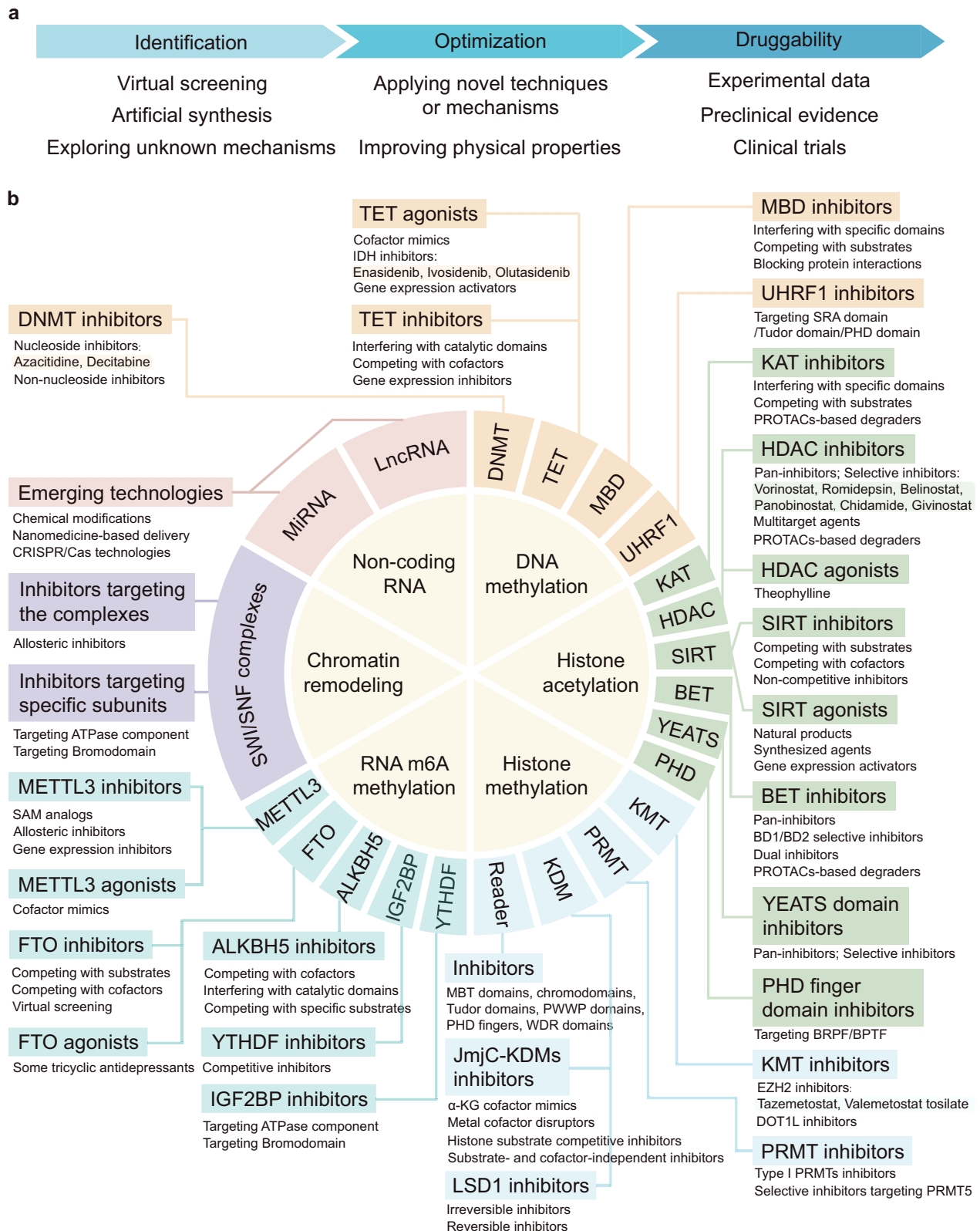


Fig. 4 The development direction and major categories of epigenetics-targeted drugs. **a** Epigenetics-targeted drugs are developed through the virtual screening of compound libraries, drug design based on molecular structure, and the exploration of potential mechanisms of known agents. Subsequently, applying PROTAC, CRISPR/Cas, and other technologies or mechanisms to optimize the physical properties, and inhibitory or agonistic effects of compounds. Finally, the druggability of possible agents should be improved in experimental, preclinical and clinical studies. **b** The classification of epigenetics-targeted drugs and their corresponding marketed representative agents are depicted in this section. Among them, epigenetics-targeted drugs that have already been approved and applied in clinical treatment are highlighted in corresponding colors

Table 3. Summary of DNA methylation-targeted drugs for different diseases in clinical trials

| Type | Drug | Target(s) | Condition(s) | Status/outcome(s) | Phase(s) | Other intervention(s)/drug(s) | Study ID/reference(s) |
|----------------|---------------|-----------|---|--|----------|--|--------------------------------|
| DNMT inhibitor | Guadecitabine | DNMT1 | Platinum refractory germ cell cancer | Completed (exhibits tolerable safety and satisfied activity) | Phase I | In combination with Cisplatin | NCT02429466 ¹⁰¹³ |
| | Guadecitabine | DNMT1 | Liver cancer, pancreatic cancer, bile duct cancer, gallbladder cancer | Active, not recruiting | Phase I | In combination with Durvalumab | NCT03257761 |
| | Guadecitabine | DNMT1 | Lung cancer | Active, not recruiting | Phase I | In combination with Pembrolizumab and Mocetinostat | NCT03220477 |
| | Guadecitabine | DNMT1 | AML | Completed (exhibits an overall unfavorable benefit-risk profile at the investigated dose levels) | Phase I | In combination with Atezolizumab | NCT02892318 ²⁷⁶ |
| | Guadecitabine | DNMT1 | Colorectal cancer | Completed (no significant clinical activity of the Guadecitabine with Cy/GVAX is observed) | Phase I | CY/GVAX (active comparator/ followed by Guadecitabine) | NCT01966289 ²⁷⁷ |
| | Guadecitabine | DNMT1 | Castration-resistant prostatic cancer, NSCLC | Recruiting (helps to reverse resistance to immune checkpoint inhibitors according to early clinical data) | Phase I | ASTX727 (active comparator); in combination with Pembrolizumab | NCT02998567 ¹⁰¹⁴ |
| | Guadecitabine | DNMT1 | AML | Completed (subcutaneous administration of large doses may be beneficial for improving treatment efficacy while increases the risk of adverse events) | Phase I | — | NCT02293993 |
| | Guadecitabine | DNMT1 | Melanoma | Completed (helps to achieve long-term clinical benefits) | Phase I | In combination with Ipilimumab | NCT02608437 ^{281,282} |
| | Guadecitabine | DNMT1 | SCLC | Completed (unpublished) | Phase I | In combination with platinum-based first-line chemotherapy, Durvalumab, and Tremelimumab | NCT03085849 |
| | Guadecitabine | DNMT1 | AML, MDS | Terminated (not due to patient safety) | Phase II | — | NCT03603964 |
| | Guadecitabine | DNMT1 | MDS | Active, not recruiting | Phase II | — | NCT02131597 |
| | Guadecitabine | DNMT1 | AML, MDS | Unknown | Phase II | ASCT | NCT03454984 |
| | Guadecitabine | DNMT1 | Paraganglioma, GIST, RCC, pheochromocytoma | Terminated (exhibits manageable toxicity with low objective response rates) | Phase II | — | NCT03165721 ²⁷⁸ |
| | Guadecitabine | DNMT1 | SCLC | Completed (exhibits good efficacy but with the possibility of adverse events) | Phase II | In combination with Cisplatin | NCT03913455 ¹⁰¹⁵ |
| | Guadecitabine | DNMT1 | Urothelial carcinoma | Active, not recruiting (possibly prolongs patient survival) | Phase II | In combination with Atezolizumab | NCT03179943 ¹⁰¹⁶ |
| | Guadecitabine | DNMT1 | Philadelphia-negative myeloproliferative neoplasms | Completed (helps improve quality of life and exhibits acceptable adverse events) | Phase II | — | NCT03075826 |
| | Guadecitabine | DNMT1 | HCC | Completed (high incidence of treatment-related adverse events) | Phase II | — | NCT01752933 |
| | Guadecitabine | DNMT1 | AML | Completed (exhibits comparable clinical response rates and safety) | Phase II | With or without Cladribine or Idarubicin | NCT02096055 |
| | Guadecitabine | DNMT1 | Fallopian tube carcinoma, peritoneal carcinoma | Completed (exhibits clinical benefit and possibly activates antitumor immunity) | Phase II | In combination with Pembrolizumab | NCT02901899 ¹⁰¹⁷ |

Table 3. continued

| Type | Drug | Target(s) | Condition(s) | Status/outcome(s) | Phase(s) | Other intervention(s)/drug(s) | Study ID/reference(s) |
|------|---------------|-------------|--|---|------------|--|----------------------------------|
| | Guadecitabine | DNMT1 | MDS | Completed (exhibits potential therapeutic effects on high-risk patients who failed azacitidine) | Phase II | — | NCT02197676 ¹⁰¹⁸ |
| | Guadecitabine | DNMT1 | AML, CMML, MDS | Active, not recruiting | Phase II | In combination with donor lymphocytes | NCT02684162 |
| | Guadecitabine | DNMT1 | Melanoma, NSCLC | Not yet recruiting | Phase II | With or without Ipilimumab plus Nivolumab | NCT04250246 |
| | Guadecitabine | DNMT1 | Central chondrosarcoma | Active, not recruiting | Phase II | In combination with Belinostat or ASTX727 | NCT04340843 ¹⁰¹⁹ |
| | Guadecitabine | DNMT1 | Ovarian cancer | Completed (helps to increase progression-free survival within six months) | Phase II | In combination with Carboplatin | NCT01696032 ¹⁰²⁰ |
| | Guadecitabine | DNMT1 | Colorectal cancer | Withdrawn (due to the insufficient funding) | Phase I/II | In combination with Nivolumab | NCT03576963 |
| | Guadecitabine | DNMT1 | RCC | Active, not recruiting (exhibits satisfied safety and tolerability) | Phase I/II | In combination with Durvalumab | NCT03308396 ¹⁰²¹ |
| | Guadecitabine | DNMT1 | AML, MDS, CMML | Active, not recruiting (exhibits manageable adverse events and typical cytopenia-related safety concerns) | Phase I/II | In combination with Atezolizumab | NCT02935361 ¹⁰²² |
| | Guadecitabine | DNMT1 | AML, MDS, CMML | Completed (exhibits well clinically active and acceptable tolerability) | Phase I/II | — | NCT01261312 ^{1023–1026} |
| | Guadecitabine | DNMT1 | Colorectal cancer | Completed (exhibits comparable efficacy and safety profiles) | Phase I/II | In combination with Irinotecan; Regorafenib or TAS-102 (active comparator) | NCT01896856 ^{280,1027} |
| | Guadecitabine | DNMT1 | Platinum-resistant fallopian tube carcinoma, platinum-resistant ovarian carcinoma, platinum-resistant primary peritoneal carcinoma | Active, not recruiting | Phase I/II | Atezolizumab (active comparator)/followed by Guadecitabine; with or without CDX-1401 vaccine | NCT03206047 |
| | Guadecitabine | DNMT1 | MDS, CMML | Completed (exhibits comparable therapeutic effects and safety profiles) | Phase III | Low-dose Cytarabine/standard IC/BSC (active comparator) | NCT02907359 |
| | Guadecitabine | DNMT1 | AML | Completed (exhibits higher clinical response rates and comparable safety) | Phase III | High-dose Cytarabine/low-dose Cytarabine/BSC(active comparator) | NCT02920008 ²⁷⁹ |
| | Guadecitabine | DNMT1 | AML | Completed (no significant clinical activity of the Guadecitabine and active comparators is observed) | Phase III | Low-dose Cytarabine/high-dose Cytarabine (active comparator) | NCT02348489 ²⁸³ |
| | MG98 | DNMT1 | Solid tumors | Completed (exhibits early evidence of clinical activity with good tolerability) | Phase I | — | NCT00003890 ²⁹⁷ |
| | MG98 | DNMT1 | Metastatic renal carcinoma | Terminated (exhibits no antitumor activities) | Phase II | — | ²⁹⁸ |
| | Hydralazine | DNMT1/3a/3b | Lung cancer | Completed (unpublished) | Phase I | In combination with Valproic acid | NCT00996060 |
| | Hydralazine | DNMT1/3a/3b | Refractory solid tumors | Completed (exhibits the potential to overcome chemotherapy resistance) | Phase II | In combination with Magnesium valproate | NCT00404508 ²⁰⁴ |
| | Hydralazine | DNMT1/3a/3b | Cervical cancer | Completed (unpublished) | Phase II | In combination with Magnesium valproate and Cisplatin chemoradiation | NCT00404326 |

Table 3. continued

| Type | Drug | Target(s) | Condition(s) | Status/outcome(s) | Phase(s) | Other intervention(s)/drug(s) | Study ID/reference(s) |
|---|---------------------|-------------|----------------------------------|---|-------------|--|-------------------------------|
| TET agonist | Hydralazine | DNMT1/3a/3b | BC | Terminated (treatment is well-tolerated) | Phase II | In combination with Magnesium valproate | NCT00395655 ³⁰¹ |
| | Hydralazine | DNMT1/3a/3b | HCC | Completed (exhibits good efficacy and manageable toxicities) | Phase II | In combination with Valproic acid | TPVGH97-07-07 ¹⁰²⁸ |
| | Hydralazine | DNMT1/3a/3b | MDS | Unknown | Phase II | In combination with Valproic acid; BSC (active comparator) | NCT01356875 |
| | Hydralazine | DNMT1/3a/3b | BC | Withdrawn (IRB request) | Phase I/II | — | NCT00575978 |
| | Hydralazine | DNMT1/3a/3b | Rectal cancer | Withdrawn (No enrollment) | Phase I/II | — | NCT00575640 |
| | Hydralazine | DNMT1/3a/3b | Ovarian cancer | Unknown | Phase III | In combination with Magnesium valproate; placebo-controlled | NCT00533299 |
| | Hydralazine | DNMT1/3a/3b | Cervical cancer | Completed (exhibits advantages in progression-free survival) | Phase III | In combination with Magnesium valproate; placebo-controlled | NCT00532818 |
| | Hydralazine | DNMT1/3a/3b | Cervical cancer | Unknown | Phase III | In combination with Magnesium valproate, Carboplatin, and Paclitaxel; placebo-controlled | NCT02446652 |
| | Vitamin C/Ascorbate | TET1/2/3 | MDS, AML | Completed (enhances the biological effects of DNMT inhibitors) | Pilot trial | In combination with Azacitidine | NCT02877277 ³³⁵ |
| | Vitamin C/Ascorbate | TET1/2/3 | MDS, AML | Completed (identifies an appropriate dose of the drug combination for phase II studies) | Phase I | In combination with Decitabine and Arsenic trioxide | NCT00671697 ³³⁶ |
| AML acute myeloid leukemia, ASCT allogeneic stem cell transplant, BC breast cancer, BSC best supportive care, CMML chronic myelomonocytic leukemia, CY Cyclophosphamide/Cytoxan, DNMT DNA methyltransferase, GVAX colon cancer tumor vaccine, GIST gastrointestinal stromal tumor, HCC hepatocellular carcinoma, IC intensive chemotherapy, IRB institutional review board, MDS myelodysplastic syndrome, NSCLC non-small cell lung cancer, RCC renal cell carcinoma, SCLC small cell lung cancer, TET ten-eleven translocation | Vitamin C/Ascorbate | TET1/2/3 | TET2-mutant MDS, TET2-mutant AML | Completed (unpublished) | Phase II | In combination with Azacitidine | NCT03397173 |
| | Vitamin C/Ascorbate | TET1/2/3 | TET2-mutant MDS | Completed (exhibits good safety profiles and tolerability) | Phase I/II | — | NCT03433781 |

reduces DNMT1 mRNA levels by targeting its 3' untranslated region. However, several clinical trials are far from satisfactory.²⁹⁷ A two-stage phase II clinical trial evaluated the antitumor efficacy of MG98 in seventeen patients with metastatic renal carcinoma.²⁹⁸ However, it failed to detect a decrease in DNMT1 activity caused by MG98, urging caution against potential side effects like transaminase elevation and fatigue from excessive dosages during intravenous administration.^{298,299} Hydralazine, a low molecular weight molecule, interacts with DNMT through a network of hydrogen bonds with arginine and glutamic acid residues.³⁰⁰ Its combination with traditional chemotherapeutic agents has been found to mitigate the progression of both hematological malignancies and solid tumors.^{301–304} In a completed phase II clinical trial, 15 patients with solid tumors qualified for the assessment of the therapeutic response to hydralazine and magnesium valproate. The majority of patients (80%), benefited from treatment and exhibited satisfactory clinical efficacy and tolerability.³⁰⁴ These findings underpin the hypothesis that epigenetic aberrations induced by chemotherapeutic agents are a primary cause of chemoresistance, providing a theoretical basis for the combined use of epigenetics-targeted drugs and chemotherapy in tumor therapy.

Targeting the eraser of DNA methylation: TET. Due to the significant heterogeneity in the roles that TET enzymes play across various diseases, the effectiveness of targeting TET enzymes as a therapeutic strategy depends on the specific disease or even different stages within a disease.³⁰⁵ While there are currently no epigenetic-targeted drugs that modulate TET available on the market, experimental studies and clinical trials suggest that reshaping methylation landscapes through TET inhibitors and agonists may be a viable approach to treating diseases.

TET inhibitors: To date, numerous small molecules have been identified that inhibit TET enzymes. Research into these TET inhibitors primarily focuses on elucidating their underlying molecular mechanisms. We categorize these inhibitors into three groups based on their distinct mechanisms of action, which will be discussed in detail below.

Auranofin, C35, and eltrombopag specifically target and bind to the catalytic domains of TET proteins, directly inhibiting their enzymatic activities. C35 exhibits potent inhibitory effects on all members of the TET family.^{306,307} In contrast, the effects of auranofin and eltrombopag are specific to TET1 and TET2, respectively.^{308,309} Notably, eltrombopag, a nonpeptidyl thrombopoietin receptor agonist approved by the US FDA for use in patients with aplastic anemia as an iron chelator.³¹⁰ Recently, Guan et al.³⁰⁹ reported the negative effects of eltrombopag on TET2. Intriguingly, this mechanism is independent of its iron chelation properties, presenting it as a potential TET2-targeted epigenetic agent and providing new insights for epigenetics-oriented therapy.³⁰⁹ Further, well-designed studies are essential to evaluate the clinical application potential of these molecular-level discoveries.

Itaconic acid, fumarate, and succinate are in vivo synthesized metabolites that indirectly impair TET catalytic activity by competitively binding to TET2 alongside α -KG, a crucial cofactor.^{311–313} These metabolites are promising precursors for developing TET-targeted epigenetic drugs, as they are well tolerated in vivo.^{314,315} However, considerable work is necessary before clinical application, such as designing appropriate carriers that can deliver these agents directly to pathological cells, given their potential impact on the vital activities of normal cells. Additionally, synthetic compounds like dimethyloxallylglycine and TETi76, which mimic the properties of α -KG, serve as competitive inhibitors of TET.^{316,317} These agents represent a novel approach to developing TET inhibitors.

Bobcat339,^{318,319} NSC-311068,³²⁰ NSC-370284,³²⁰ and UC-514321,³²⁰ inhibit DNA methylation by reducing intracellular TET levels. Bobcat339 induces the degradation of TET3 directly,³¹⁸ and its inhibitory effects on TET1 and TET2 are observed only in the presence of coordinating copper(II).³²¹ NSC-370284 and UC-514321 bind directly to the DNA-binding domain of signal transducer and activator of transcription 3 (STAT3) or STAT5, transcriptional activators of TET1, leading to suppressed expression of TET1 in vivo.³²⁰ This mechanism has been confirmed in mouse models of AML and medulloblastoma, showing synergistic effects with standard chemotherapy.^{320,322} The therapeutic potential of these compounds in additional diseases is an exciting area for future research.

TET agonists: As previously discussed, TET inhibitors are highly valued for treating diseases. Conversely, research into TET agonists is also anticipated to yield promising breakthroughs and pave the way for clinical applications. Most TET agonists currently under investigation are drugs that upregulate cofactors of TET, such as vitamin C and enzymes involved in α -KG metabolism; other small molecules, including 3-nitroflavanones,²⁹⁵ retinoic acid,³²³ ioperamide hydrochloride,³²⁴ and mitoxantrone,³²⁵ are reported to directly upregulate TET expression.

Vitamin C, or ascorbate, uniquely interacts with the C-terminal catalytic domain of TET, positioning it as a novel epigenetic-modifying agent.^{326,327} As an antioxidant, it also helps maintain the divalent state of iron ions, indirectly supporting TET activity.³²³ Characterized by TET repair and increased 5hmC levels, vitamin C administration can exert therapeutic roles in various tumors and non-tumor diseases.^{328–330} Furthermore, it has been used as an adjuvant, synergizing with other immunotherapeutic or chemotherapeutic agents.^{331–334} The synergistic treatment of vitamin C with azacitidine or decitabine in clinical trials has shown positive outcomes for patients with myeloid tumors.^{335,336} However, the optimal doses, frequency, and duration of vitamin C administration remain debated, with long-term treatment and follow-up required for further investigation.³³⁷ Therefore, the full exploration of the therapeutic role of vitamin C as an epigenetic-modifying drug is crucial for its future clinical applications.

Inhibitors of IDH and α -KG dehydrogenase elevate α -KG levels. In tumor cells, IDH1/2 mutations lead to the production of the oncometabolite 2-HG, which competes with α -KG for binding sites on TET, potentially leading to reversible inhibition of TET proteins and dysregulation of DNA methylation levels.^{338,339} The administration of enasidinib and siRNA against IDH2 has been shown to restore the low methylated state of the genome, consistent with the reactivation of TET enzymatic activities.^{340–342} However, observations suggest that other α -KG-dependent enzymes, such as histone demethylases and prolyl hydroxylases, might play a more dominant role in the progression of IDH-mutant diseases.³⁴³ These findings highlight the potential for developing TET agonists based on IDH inhibitors to reshape the epigenetic landscape, warranting further investigation. Additionally, inhibiting α -KG dehydrogenase enhances α -KG levels and TET activities, restoring DNA methylation and ameliorating the progression of T2DM and breast cancer.^{344,345} Similarly, IOX1, an inhibitor of α -KG oxygenases and a potent inhibitor of lysine demethylase 3A (KDM3A) and KDM4A, has been found to reduce TET enzymatic activities in helper T cells, emerging as a potential epigenetic drug for various autoimmune diseases.³⁴⁶ In conclusion, α -KG represents a promising target for TET-targeted drug development that should be further explored in clinical trials.

Targeting the reader of DNA methylation: MBD and UHRF1. With the discovery of proteins that read methylated DNA sites, burgeoning research has aimed to identify small molecules

targeting these enzymes, sparking considerable enthusiasm for developing novel therapeutic targets.

MBD inhibitors: The MBD protein family, critical readers of DNA methylation, consists of six members: methyl-CpG-binding protein 1 (MeCP1), MeCP2, MBD1, MBD2, MBD3, and MBD4.³⁴⁷ The aberrant activities and expression of these proteins observed in various diseases have recently positioned them as potential targets for epigenetic drugs. Current research predominantly focuses on MBD2 inhibitors, which are rapidly progressing.

The development of MBD2 inhibitors hinges on two prerequisite factors. Firstly, the knockdown of MBD2 or the application of targeted siRNA has demonstrated positive effects in tumor treatment, underscoring the therapeutic potential of targeting MBD2.³⁴⁸ Secondly, successfully elucidating MBD2's molecular structure and associated mechanisms lays the scientific groundwork for identifying and designing inhibitory molecules. MBD2 inhibitors can be categorized into three groups based on their mechanisms of action. The first group disrupts the binding of the N-terminal MBD to methylated DNA.³⁴⁹ Through docking analysis, molecules such as CID3100583 and 8,8-ethylenebis(2-ethyl-2-phenyl-2-oxo-1,3-dioxane-5-carboxamide) have been identified to target the interaction between MBD2 and DNA.³⁵⁰ The second category aims to block the interaction between the C-terminal coiled-coil domain and the GATA zinc finger domain containing 2A, which has shown potent inhibitory effects on MBD2-dependent DNA methylation.³⁵¹ However, no drugs based on this mechanism are currently in use, highlighting a gap in research that demands further exploration. The third group prevents the interaction with HDAC and the formation of the nucleosome remodeling and deacetylase complex via an intrinsically disordered region.^{352–354} Utilizing this concept, Na et al.³⁵⁵ developed a novel technique that efficiently discriminates potential compounds interacting with intrinsically disordered proteins through expanded virtual screening. This approach led to identifying two MBD2 inhibitors, ABA and APC. These findings provide a sound basis for a therapeutic strategy targeting MBD2 and advocate for more comprehensive *in vivo* studies to assess their efficacy and safety.

UHRF1 inhibitors: Ubiquitin-like with plant homeodomain (PHD) and RING finger domains 1 (UHRF1) plays a pivotal role in recruiting DNMT1 during replication, primarily through the recognition of hemimethylated DNA and the subsequent flipping of hemimethylated CpG sites via the SET and RING associated domain (SRA).^{356–358} UHRF1 also interacts with HDAC1 and facilitates the di- and tri-methylation of H3, contributing to the ubiquitination of histones and the formation of heterochromatin.³⁵⁹ The upregulation of UHRF1 has been observed in various pathologies, particularly in tumors, making it a promising target for therapeutic intervention.^{360,361}

The initial identification of small inhibitors targeting the SRA domain of UHRF1 was based on structure-based screening and computational analyses. Compounds such as NSC232003,³⁶² UM63,³⁶³ UF146,³⁶⁴ chicoric acid,³⁶⁵ have been shown to block the interaction between UHRF1 and 5mC sites, effectively preventing the proliferation of diverse cancer cell lines. Advanced screening techniques, such as nonequilibrium capillary electrophoresis of the equilibrium mixture, have facilitated the identification of proanthocyanidins and baicalein as promising inhibitors.³⁶⁶ Furthermore, Ciaco et al.³⁶⁷ reported the development of novel UHRF1 inhibitors, AMSA2 and MPB7, based on the structure of UM63. These inhibitors suppress SRA-mediated base-flipping activities without DNA intercalation and demonstrate minimal effects on non-cancer cells, offering a basis for further optimization. In addition to the SRA domain, the tandem Tudor domain and PHD domain of UHRF1, which are involved in recognizing methylated lysine and arginine residues on H3, have also become targets for inhibitor design.^{368,369} While some inhibitors targeting

these domains have been reported, their effects have only been validated *in vitro*, and more evidence is needed before proceeding to clinical trials.^{370–372} Current research suggests that inhibiting UHRF1 alone may not be sufficient to restore gene silencing affected by hypermethylation. However, combining UHRF1 inhibitors with other epigenetic inhibitors, such as HDAC inhibitors, can lead to synergistic effects and improved therapeutic outcomes.^{373,374} Consequently, multi-target inhibitors have been developed and are emerging as clinical candidates for tumor therapy.^{375–378} Additionally, the use of small molecules that act as UHRF1 degraders, such as diosgenin and MK2206, has been explored for prostate cancer treatment, representing a novel therapeutic approach.^{379,380} Natural substances like hinokitiol have shown therapeutic effects in mouse models in a UHRF1 depletion-dependent manner although the underlying mechanisms remain to be fully elucidated and represent a direction for future research.³⁸¹

Epigenetics-targeted drugs and histone acetylation

The dynamic equilibrium and normal function of histone acetylation and deacetylation are regulated by the cooperative actions of the lysine acetyltransferase (KAT) and HDAC families, along with various reader proteins. Acetylation of lysine residues at the N-terminus of histones induces negative charges that trigger gene transcription, while decreased acetylation down-regulates gene expression. Conversely, an imbalance in histone acetylation/deacetylation disrupts normal gene expression patterns, leading to the onset and progression of diseases. The development of related epigenetic drugs is ongoing, offering new therapeutic options for treating these conditions (Table 4).

Targeting the writer of histone acetylation: KAT. Histone acetylation serves various functions within cells. However, aberrant acetylation catalyzed by KAT can trigger the pathogenesis of various human diseases, including neurodegenerative diseases, metabolic diseases, and tumors.^{382–385} Developing epigenetic drugs that regulate KAT activity is a promising avenue for treating these diseases.

KAT inhibitors: Numerous inhibitors targeting KAT have been developed, primarily focusing on the P300/CBP, GNAT/PCAF, and MYST classes.^{386–388} These enzymes contain two accessible domains: the acetyl-lysine binding BD and the catalytic domain, utilizing acetyl CoA as a cofactor to transfer acetyl groups. Thus, designed inhibitors can target the enzymatic activity and the binding sites for acetyl CoA. Moreover, research into KAT degraders is advancing with the advent of proteolysis-targeting chimeras (PROTACs). These degraders are ternary complexes comprising ligands for targeted proteins and E3 ubiquitin ligase, along with a connecting linker, allowing targeted degradation via a ubiquitination-dependent method.³⁸⁹

Five KAT inhibitors have entered clinical practice, including CCS1477, FT-7051, NEO2734, PRI-724, and PF-07248144. CCS1477 targets the P300/CBP (KAT3A/KAT3B) via interaction with the BD fragment, exhibiting potent antitumor effects in cancer cell lines and animal models.^{390,391} This has led to its application in monotherapy and in combination with chemotherapeutic drugs in phase I and II clinical trials (NCT04068597, NCT03568656), with the potential to improve therapeutic strategies for both advanced solid tumors and hematological malignancies. FT-7051, another P300/CBP inhibitor targeting the BD domain, has been shown to reduce H3K27Ac at specific promoter sites and is currently under study in a phase I clinical trial for patients with hormone receptor-positive prostate cancer.³⁹² NEO2734, a dual P300/CBP and BET inhibitor, demonstrates therapeutic potential comparable to the combination of a BET inhibitor and a P300/CBP inhibitor in treating certain cancers.^{393,394} It is currently being evaluated in a phase I clinical trial focusing on castration-resistant prostate

Table 4. Summary of histone acetylation-targeted drugs for different diseases in clinical trials

| Type | Drug | Target(s) | Condition(s) | Status/outcome(s) | Phase(s) | Other intervention(s)/drug(s) | Study ID/reference(s) |
|----------------|--------------|-----------------------------|---|---|------------|---|-----------------------------|
| KAT inhibitor | CCS1477 | P300/CBP (KAT3A/KAT3B) | NHL, MM, AML, MDS, PTCL | Recruiting | Phase I/II | With or without Pomalidomide plus Dexamethasone, or Azacitidine plus Venetoclax | NCT04068597 |
| | CCS1477 | P300/CBP (KAT3A/KAT3B) | CRPC, BC, NSCLC | Recruiting | Phase I/II | With or without Abiraterone acetate or Enzalutamide or Darolutamide or Olaparib or Atezolizumab | NCT03568656 |
| | FT-7051 | P300/CBP (KAT3A/KAT3B) | CRPC | Terminated (due to sponsors' decision) | Phase I | — | NCT04575766 |
| | NEO2734 | P300/CBP (KAT3A/KAT3B), BET | CRPC, NUT carcinoma | Recruiting | Phase I | — | NCT05488548 |
| | PRI-724 | CBP/β-catenin | HCV-induced cirrhosis | Completed (causes liver injury in the high-dose cohort) | Phase I | — | NCT02195440 ³⁹⁷ |
| | PRI-724 | CBP/β-catenin | PDAC | Completed (unpublished) | Phase I | — | NCT01764477 |
| | PRI-724 | CBP/β-catenin | Solid tumors | Terminated (due to low enrollment) | Phase I | — | NCT01302405 |
| | PRI-724 | CBP/β-catenin | HIV/HCV co-induced cirrhosis | Completed (unpublished) | Phase I | — | NCT04688034 |
| | PRI-724 | CBP/β-catenin | PBC | Completed (unpublished) | Phase I | — | NCT04047160 |
| | PRI-724 | CBP/β-catenin | HIV/HCV co-induced cirrhosis | Recruiting | Phase II | — | NCT06144086 |
| HDAC inhibitor | PRI-724 | CBP/β-catenin | AML, CML | Completed (unpublished) | Phase I/II | — | NCT01606579 |
| | PRI-724 | CBP/β-catenin | HCV-induced cirrhosis, HBV-induced cirrhosis | Completed (exhibits insufficient evidence of improvement in hepatic function) | Phase I/II | — | NCT03620474 ³⁹⁶ |
| | PF-07248144 | KAT6 | HR-positive, HER2-negative BC, CRPC, NSCLC | Recruiting | Phase I | With or without Fulvestrant, or Letrozole plus Palbociclib, or Fulvestrant plus PF-07220060 | NCT04606446 |
| | Ivaltinostat | Pan-HDAC | Malignant tumors | Active, not recruiting | Phase I | Placebo-controlled | NCT05716919 |
| | Ivaltinostat | Pan-HDAC | Healthy volunteers | Completed (the oral formulation of Ivaltinostat is well tolerated) | Phase I | Placebo-controlled | NCT05345912 ¹⁰²⁹ |
| | Ivaltinostat | Pan-HDAC | PDAC | Unknown (exhibits good efficacy and an acceptable safe profile according to disclosed data) | Phase I/II | In combination with Gemcitabine and Erlotinib | NCT02737228 ⁴⁰¹ |
| | Ivaltinostat | Pan-HDAC | PDAC | Recruiting | Phase I/II | Capecitabine (active comparator/in combination with Ivaltinostat) | NCT05249101 |
| | Abexinostat | Pan-HDAC | High-grade glioma | Recruiting | Phase I | In combination with Temozolomide | NCT05698524 |
| | Abexinostat | Pan-HDAC | DLBCL, MCL | Active, not recruiting | Phase I | In combination with Ibrutinib | NCT03939182 |
| | Abexinostat | Pan-HDAC | Melanoma, squamous cell carcinoma of head and neck, urothelial carcinoma, NSCLC | Completed (unpublished) | Phase I | In combination with Pembrolizumab | NCT03590054 |
| Abexinostat | Abexinostat | Pan-HDAC | NHL, HL, MM | Completed (unpublished) | Phase I | — | NCT01149668 |
| | Abexinostat | Pan-HDAC | Solid tumors | Active, not recruiting (exhibits good tolerability and antitumor effects according to disclosed data) | Phase I | In combination with Pazopanib | NCT01543763 ⁴⁰⁴ |
| | Abexinostat | Pan-HDAC | NHL, HL, MM | Completed (unpublished) | Phase I | — | NCT00562224 |
| | Abexinostat | Pan-HDAC | Malignant tumors | Completed (unpublished) | Phase I | — | NCT00473577 |

Table 4. continued

| Type | Drug | Target(s) | Condition(s) | Status/outcome(s) | Phase(s) | Other intervention(s)/drug(s) | Study ID/reference(s) |
|------|-------------|--------------------|---|--|--------------|---|--|
| | Abexinostat | Pan-HDAC | MDS, AML, ALL | Terminated (due to limited clinical benefit) | Phase I | — | ISRCTN 99680465 ¹⁰³⁰ |
| | Abexinostat | Pan-HDAC | FL | Active, not recruiting | Phase II | — | NCT03600441 |
| | Abexinostat | Pan-HDAC | DLBCL | Recruiting | Phase II | — | NCT03936153 |
| | Abexinostat | Pan-HDAC | FL | Recruiting | Phase II | — | NCT03934567 |
| | Abexinostat | Pan-HDAC | NHL | Active, not recruiting | Phase I/II | — | NCT04024696 |
| | Abexinostat | Pan-HDAC | NHL, HL | Completed (exhibits tolerable safety and significant clinical activity in FL) | Phase I/II | — | NCT00724984 ⁴⁰⁶ |
| | Abexinostat | Pan-HDAC | Sarcoma | Completed (exhibits manageable toxicities and tumor responses) | Phase I/II | In combination with Doxorubicin and GCSF | NCT01027910 ¹⁰³¹ |
| | Abexinostat | Pan-HDAC | B cell lymphoma, CML | Completed (exhibits manageable toxicity and partial responses) | Phase I/II | — | EudraCT 2009-013691-47 ^{405,1032} |
| | Abexinostat | Pan-HDAC | RCC | Recruiting | Phase III | Pazopanib (active comparator/in combination with Abexinostat) | NCT03592472 |
| | AR-42 | Pan-HDAC | Vestibular schwannoma, meningioma, acoustic neuroma, neurofibromatosis type 2 | Terminated (due to a lack in drug supply) | Phase I | — | NCT02282917 |
| | AR-42 | Pan-HDAC | RCC, soft tissue sarcoma | Terminated (due to a lack in drug supply) | Phase I | In combination with Pazopanib | NCT02795819 |
| | AR-42 | Pan-HDAC | AML | Completed (exits possibilities of serious treatment-associated adverse events) | Phase I | In combination with Decitabine | NCT01798901 ¹⁰³³ |
| | AR-42 | Pan-HDAC | Hematologic malignancies | Completed (exhibits tolerable safety) | Phase I | — | NCT01129193 ^{402,1034} |
| | AR-42 | Pan-HDAC | Plasma cell myeloma | Completed (unpublished) | Phase I | In combination with Dexamethasone and Pomalidomide | NCT02569320 |
| | AR-42 | Pan-HDAC | Neurofibromatosis type 2 | Recruiting | Phase II/III | Placebo-controlled | NCT05130866 |
| | Pracinostat | HDAC class I/II/IV | Healthy volunteers | Completed (unpublished) | Phase I | — | NCT03495934 |
| | Pracinostat | HDAC class I/II/IV | Healthy volunteers | Completed (unpublished) | Phase I | Fasted or fed conditions | NCT02058784 |
| | Pracinostat | HDAC class I/II/IV | Healthy volunteers | Completed (unpublished) | Phase I | In combination with Ciprofloxacin or Itraconazole | NCT02118909 |
| | Pracinostat | HDAC class I/II/IV | Solid tumors, leukemia | Completed (unpublished) | Phase I | — | NCT01184274 |
| | Pracinostat | HDAC class I/II/IV | Solid tumors, hematologic malignancies | Completed (exhibits safety and modest single-agent activity in hematologic malignancies) | Phase I | With or without Azacitidine | NCT00741234 ¹⁰³⁵ |
| | Pracinostat | HDAC class I/II/IV | AML | Completed (unpublished) | Phase I | Gemtuzumab Ozogamicin (active comparator/in combination with Pracinostat) | NCT03848754 |
| | Pracinostat | HDAC class I/II/IV | Solid tumors | Completed (unpublished) | Phase I | — | NCT00504296 |
| | Pracinostat | HDAC class I/II/IV | Solid tumors | Completed (exhibits good tolerability and inhibitory effects) | Phase I | — | SCS-PN0022 ¹⁰³⁶ |
| | Pracinostat | HDAC class I/II/IV | Solid tumors | Completed (exhibits good tolerability) | Phase I | — | ¹⁰³⁷ |
| | Pracinostat | HDAC class I/II/IV | Solid tumors | Completed (exhibits good tolerability and inhibitory effects) | Phase II | — | NCT01912274 ¹⁰³⁸ |
| | Pracinostat | HDAC class I/II/IV | MDS | Terminated (due to sponsors' decision) | Phase II | In combination with Azacitidine | NCT03151304 |
| | Pracinostat | HDAC class I/II/IV | Myelofibrosis | Completed (worsening anemia and other adverse events do not support the continued development) | Phase II | In combination with Ruxolitinib and Questionnaire | NCT02267278 ¹⁰³⁹ |

Table 4. continued

| Type | Drug | Target(s) | Condition(s) | Status/outcome(s) | Phase(s) | Other intervention(s)/drug(s) | Study ID/reference(s) |
|------|--------------|---------------------|---|--|------------|--|---------------------------------|
| | Pracinostat | HDAC class I/II/IV | MDS | Completed (reduced doses exhibit improved tolerability and efficacy) | Phase II | In combination with Azacitidine and Decitabine | NCT01993641 ¹⁰⁴⁰ |
| | Pracinostat | HDAC class I/II/IV | CRPC | Completed (exhibits insufficient activity as a single agent) | Phase II | — | NCT01075308 ¹⁰⁴¹ |
| | Pracinostat | HDAC class I/II/IV | MDS | Completed (fails to improve outcomes at the available dosing regimen) | Phase II | Azacitidine (active comparator/in combination with Pracinostat) | NCT01873703 ¹⁰⁴² |
| | Pracinostat | HDAC class I/II/IV | Myeloproliferative disorders | Completed (exhibits reasonable tolerability and modest activity in myelofibrosis) | Phase II | — | NCT01200498 ¹⁰⁴³ |
| | Pracinostat | HDAC class I/II/IV | Sarcoma | Completed (premature stop due to the prolonged unavailability) | Phase II | — | NCT01112384 ¹⁰⁴⁴ |
| | Pracinostat | HDAC class I/II/IV | AML | Terminated (due to a lack of efficacy) | Phase III | Azacitidine (active comparator/in combination with Pracinostat) | NCT03151408 ¹⁰⁴⁵ |
| | Resminostat | HDAC class I/IIb/IV | CTCL, MF | Completed (unpublished) | Phase I | — | NCT04955340 |
| | Resminostat | HDAC class I/IIb/IV | Biliary tract cancer, pancreatic cancer | Completed (exhibits acceptable tolerability) | Phase I | In combination with chemotherapy | JapicCTI-152864 ¹⁰⁴⁶ |
| | Resminostat | HDAC class I/IIb/IV | Solid tumors | Completed (exhibits on-target pharmacodynamic activity at dose levels ≥ 400 mg and signs of antitumor efficacy) | Phase I | — | ¹⁰⁴⁷ |
| | Resminostat | HDAC class I/IIb/IV | CTCL, MF | Active, not recruiting | Phase II | Placebo-controlled | NCT02953301 |
| | Resminostat | HDAC class I/IIb/IV | HL | Completed (exhibits acceptable safety and efficacy) | Phase II | — | NCT01037478 ¹⁰⁴⁸ |
| | Resminostat | HDAC class I/IIb/IV | HCC | Completed (exhibits early signs of efficacy and good tolerability) | Phase II | With or without Sorafenib | NCT00943449 ¹⁰⁴⁹ |
| | Resminostat | HDAC class I/IIb/IV | Biliary tract cancer | Completed (exhibits no significant improve in clinical activity) | Phase II | In combination with chemotherapy | JapicCTI-183883 ¹⁰⁵⁰ |
| | Resminostat | HDAC class I/IIb/IV | HCC | Completed (no significant efficacy advantage over sorafenib monotherapy) | Phase I/II | Sorafenib (active comparator/in combination with Resminostat) | NCT02400788 ¹⁰⁵¹ |
| | Resminostat | HDAC class I/IIb/IV | NSCLC | Completed (fails to improve progression-free survival and increases toxicity) | Phase I/II | In combination with Docetaxel | JapicCTI-132123 ¹⁰⁵² |
| | Resminostat | HDAC class I/IIb/IV | Colorectal carcinoma | Completed (unpublished) | Phase I/II | Chemotherapy (active comparator/in combination with Resminostat) | NCT01277406 |
| | Tacedinaline | HDAC class I/II/III | Solid tumors | Completed (exhibits antitumor activity) | Phase I | In combination with Carboplatin and Paclitaxel | ¹⁰⁵³ |
| | Tacedinaline | HDAC class I/II/III | Solid tumors | Completed (thrombocytopenia is the main principal dose-limiting toxicity) | Phase I | In combination with Capecitabine | ¹⁰⁵⁴ |
| | Tacedinaline | HDAC class I/II/III | Solid tumors | Completed (exhibits preliminary efficacy and potential adverse events) | Phase I | In combination with Gemcitabine hydrochloride | ¹⁰⁵⁵ |
| | Tacedinaline | HDAC class I/II/III | Solid tumors | Completed (exhibits preliminary efficacy and potential adverse events) | Phase I | — | ¹⁰⁵⁶ |
| | Tacedinaline | HDAC class I/II/III | MM | Completed (unpublished) | Phase II | — | NCT00005624 |

Table 4. continued

| Type | Drug | Target(s) | Condition(s) | Status/outcome(s) | Phase(s) | Other intervention(s)/drug(s) | Study ID/reference(s) |
|------|---------------------|---------------------|--|---|------------|--|-----------------------------|
| | Tacedinaline | HDAC class I/II/III | Pancreatic cancer | Completed (exhibits no evidence for improving efficacy) | Phase II | In combination with Gemcitabine hydrochloride; placebo-controlled | NCT00004861 ¹⁰⁵⁷ |
| | Tacedinaline | HDAC class I/II/III | NSCLC | Completed (unpublished) | Phase III | In combination with Gemcitabine hydrochloride; placebo-controlled | NCT00005093 |
| | FRM-0334 | HDAC class I/I | Frontotemporal dementia with granulin mutation | Unknown (exhibits tolerable safety while insufficient efficacy according to disclosed data) | Phase II | Placebo-controlled | NCT02149160 ¹⁰⁵⁸ |
| | Trichostatin A | HDAC class I/II | Hematologic malignancies | Unknown | Phase I | — | NCT03838926 |
| | Quisinostat | HDAC class I/II | Leukemia, MDS | Terminated (due to sponsors' decision) | Phase I | — | NCT00676728 |
| | Quisinostat | HDAC class I/II | Solid tumors, lymphomas | Completed (intermittent schedules exhibit better tolerated than continuous schedules) | Phase I | — | NCT00677105 ¹⁰⁵⁹ |
| | Quisinostat | HDAC class I/II | NSCLC, ovarian cancer | Completed (unpublished) | Phase I | In combination with Cisplatin plus Gemcitabine, or Paclitaxel plus Carboplatin | NCT02728492 |
| | Quisinostat | HDAC class I/II | MM | Completed (exhibits efficacy and tolerable safety) | Phase I | In combination with Dexamethasone and Bortezomib | NCT01464112 ¹⁰⁶⁰ |
| | Quisinostat | HDAC class I/II | CTCL | Completed (exhibits an acceptable safety profile) | Phase II | — | NCT01486277 ¹⁰⁶¹ |
| | Quisinostat | HDAC class I/II | Ovarian cancer | Completed (unpublished) | Phase II | In combination with Paclitaxel and Carboplatin | NCT02948075 |
| | CXD101 | HDAC class I | Malignant tumors | Completed (exhibits acceptable tolerability with efficacy in HL, T cell lymphoma, and FL) | Phase I | — | NCT01977638 ¹⁰⁶² |
| | CXD101 | HDAC class I | HCC | Recruiting | Phase II | In combination with Teptanolimab, Lenvatinib and Sorafenib (active comparator) | NCT05873244 |
| | CXD101 | HDAC class I | Colorectal carcinoma | Unknown (exhibits good tolerability and efficacy according to disclosed data) | Phase I/II | In combination with Nivolumab | NCT03993626 ¹⁰⁶³ |
| | CXD101 | HDAC class I | DLBCL | Withdrawn (due to insufficient funds) | Phase I/II | In combination with Pembrolizumab | NCT03873025 |
| | Magnesium valproate | HDAC class I | Solid tumors | Completed (exhibits the potential to overcome chemotherapy resistance) | Phase II | In combination with Hydralazine | NCT00404508 ³⁰⁴ |
| | Magnesium valproate | HDAC class I | Cervical cancer | Completed (unpublished) | Phase II | In combination with Hydralazine | NCT00404326 |
| | Magnesium valproate | HDAC class I | Colorectal carcinoma | Recruiting | Phase II | With or without Panitumumab and Cetuximab | NCT05694936 |
| | Magnesium valproate | HDAC class I | BC | Terminated (treatment is well-tolerated) | Phase II | In combination with Hydralazine | NCT00395655 ³⁰¹ |
| | Magnesium valproate | HDAC class I | Ovarian cancer | Unknown | Phase III | In combination with Hydralazine, placebo-controlled | NCT00533299 |
| | Magnesium valproate | HDAC class I | Cervical cancer | Completed (exhibits advantages in progression-free survival) | Phase III | In combination with Hydralazine; placebo-controlled | NCT00532818 |
| | Magnesium valproate | HDAC class I | Cervical cancer | Unknown | Phase III | In combination with Hydralazine, Carboplatin, and Paclitaxel; placebo-controlled | NCT02446652 |
| | OBP-801 | HDAC class I | Solid tumors | Unknown (large-scale trials should be held according to disclosed data) | Phase I | — | NCT02414516 ¹⁰⁶⁴ |
| | Nanatinostat | HDAC class I | Malignant tumors (excluding gastrointestinal tumors) | Recruiting | Phase I | With or without Valganciclovir | NCT06302140 |
| | Nanatinostat | HDAC class I | EBV-associated lymphoma, PTCL, PTLD | Recruiting | Phase II | In combination with Valganciclovir | NCT05011058 |

Table 4. continued

| Type | Drug | Target(s) | Condition(s) | Status/outcome(s) | Phase(s) | Other intervention(s)/drug(s) | Study ID/reference(s) |
|------|--------------|--------------|--|--|---------------|---|----------------------------------|
| | Nanatinostat | HDAC class I | EBV-associated lymphoma | Completed (exhibits encouraging efficacy) | Phase I/II | In combination with Valganciclovir | NCT03397706 ¹⁰⁶⁵ |
| | Nanatinostat | HDAC class I | EBV-associated solid tumors | Recruiting | Phase I/II | In combination with Valganciclovir, with or without Pembrolizumab | NCT05166577 |
| | Entinostat | HDAC class I | TNBC | Terminated (due to funding withdrawn) | Early phase I | — | NCT03361800 |
| | Entinostat | HDAC class I | Healthy volunteers | Completed (unpublished) | Phase I | — | NCT02922946 |
| | Entinostat | HDAC class I | Healthy volunteers; renal impairment | Completed (unpublished) | Phase I | — | NCT03192111 |
| | Entinostat | HDAC class I | Solid tumors | Completed (unpublished) | Phase I | Placebo-controlled | NCT02897778 |
| | Entinostat | HDAC class I | CRPC | Completed (exhibits an acceptable safety profile) | Phase I | In combination with Enzalutamide | NCT03829930 ¹⁰⁶⁶ |
| | Entinostat | HDAC class I | HR-positive HER2-negative BC | Completed (exhibits reasonable safety, tolerability, and encouraging efficacy) | Phase I | In combination with Exemestane | NCT02833155 ¹⁰⁶⁷ |
| | Entinostat | HDAC class I | MM, MDS, myeloproliferative diseases | Completed (exhibits effective inhibition on HDAC in vivo) | Phase I | — | NCT00015925 ¹⁰⁶⁸ |
| | Entinostat | HDAC class I | Solid tumors, lymphomas | Completed (exhibits good tolerability at the studied doses) | Phase I | — | NCT00020579 ¹⁰⁶⁹ |
| | Entinostat | HDAC class I | Healthy volunteers; renal impairment | Completed (unpublished) | Phase I | In combination with Midazolam | NCT03187015 |
| | Entinostat | HDAC class I | Solid tumors | Completed (unpublished) | Phase I | In combination with Pembrolizumab | NCT02909452 |
| | Entinostat | HDAC class I | MDS | Active, not recruiting (exhibits limited clinical efficacy and substantial toxicity according to disclosed data) | Phase I | In combination with Pembrolizumab | NCT02936752 ¹⁰⁷⁰ |
| | Entinostat | HDAC class I | BC | Completed (unpublished) | Phase I | In combination with Capecitabine | NCT03473639 |
| | Entinostat | HDAC class I | Ovarian cancer, peritoneal cancer, fallopian tube cancer | Terminated (due to changes in participant landscape and other treatment availability) | Phase I | In combination with Olaparib | NCT03924245 |
| | Entinostat | HDAC class I | HR-positive BC | Completed (unpublished) | Phase I | In combination with Exemestane | NCT02820961 |
| | Entinostat | HDAC class I | HR-positive BC, NSCLC | Completed (results published along with phase II studies) | Phase I | In combination with Erlotinib and Exemestane | NCT01594398 |
| | Entinostat | HDAC class I | Lymphoma | Completed (exhibits tolerable safety) | Phase I | In combination with Isotretinoin | NCT00098891 ¹⁰⁷¹ |
| | Entinostat | HDAC class I | AML, MDS, CMML | Completed (increases toxicity in treating myeloid neoplasms) | Phase I | In combination with Azacitidine | NCT00101179 ¹⁰⁷²⁻¹⁰⁷⁴ |
| | Entinostat | HDAC class I | Healthy volunteers | Completed (unpublished) | Phase I | Dietary supplements (Omeprazole and Famotidine) | NCT02922933 |
| | Entinostat | HDAC class I | HR-positive BC | Completed (exhibits no additional safety concerns) | Phase I | In combination with KHK2375 | NCT02623751 ¹⁰⁷⁵ |
| | Entinostat | HDAC class I | Colorectal carcinoma | Completed (the combination is poorly tolerated without evident activity) | Phase I | In combination with Hydroxychloroquine and Regorafenib | NCT03215264 ⁴¹³ |
| | Entinostat | HDAC class I | SCLC | Completed (further exploration should not be applied) | Phase I | In combination with Atezolizumab, Carboplatin, and Etoposide | NCT04631029 ⁴¹⁴ |
| | Entinostat | HDAC class I | CNS tumors, lymphoma | Completed (exhibits good tolerability) | Phase I | — | NCT02780804 ¹⁰⁷⁶ |

Table 4. continued

| Type | Drug | Target(s) | Condition(s) | Status/outcome(s) | Phase(s) | Other intervention(s)/drug(s) | Study ID/reference(s) |
|------|------------|--------------|---|--|----------|--|----------------------------------|
| | Entinostat | HDAC class 1 | Endometrial endometrioid adenocarcinoma | Completed (no immediate effect on the regulation of progesterone receptor) | Phase I | With or without Medroxyprogesterone acetate | NCT03018249 ⁴¹⁵ |
| | Entinostat | HDAC class 1 | HER2-positive BC, TNBC | Terminated (due to slow accrual and company reasons) | Phase I | M7824 and Ado-trastuzumab emtansine (active comparator)/in combination with Entinostat | NCT04296942 |
| | Entinostat | HDAC class 1 | HER2-positive BC | Completed (exhibits acceptable tolerability and antitumor activity) | Phase I | In combination with Lapatinib ditosylate | NCT01434303 ¹⁰⁷⁷ |
| | Entinostat | HDAC class 1 | ALL, ABL | Completed (exhibits less activities in relapsed/refractory patients) | Phase I | In combination with Clofarabine | NCT01132573 ⁴¹⁶ |
| | Entinostat | HDAC class 1 | Solid tumors | Terminated (exhibits good tolerability according to disclosed data) | Phase I | In combination with Sorafenib | NCT01159301 ¹⁰⁷⁸ |
| | Entinostat | HDAC class 1 | NSCLC | Terminated | Phase I | In combination with Azacitidine | NCT01886573 |
| | Entinostat | HDAC class 1 | RCC | Active, not recruiting (exhibits acceptable safety and efficacy according to disclosed data) | Phase I | In combination with Aldesleukin | NCT01038778 |
| | Entinostat | HDAC class 1 | BC | Terminated | Phase I | — | NCT00754312 |
| | Entinostat | HDAC class 1 | HR-positive BC, TNBC | Active, not recruiting (exhibits good efficacy according to disclosed data) | Phase I | In combination with Ipilimumab and Nivolumab | NCT02453620 ¹⁰⁷⁹ |
| | Entinostat | HDAC class 1 | BC | Completed (exhibits acceptable safety) | Phase II | Exemestane (active comparator)/in combination with Entinostat; placebo-controlled | NCT03291886 ¹⁰⁸⁰ |
| | Entinostat | HDAC class 1 | Uveal melanoma | Completed (exhibits durable responses in a subset of patients) | Phase II | In combination with Pembrolizumab | NCT02697630 ^{1081,1082} |
| | Entinostat | HDAC class 1 | TNBC | Active, not recruiting (exhibits good tolerability but fails to meet primary endpoint according to disclosed data) | Phase II | In combination with Azacitidine | NCT01349959 |
| | Entinostat | HDAC class 1 | HL | Terminated (due to corporate decision) | Phase II | — | NCT00866333 ¹⁰⁸³ |
| | Entinostat | HDAC class 1 | MDS, AML | Completed (increases toxicity in treating myeloid neoplasms) | Phase II | Azacitidine (active comparator)/in combination with Entinostat | NCT00313586 ^{1072,1084} |
| | Entinostat | HDAC class 1 | HR-positive BC | Completed (exhibits good tolerability and clinical activity) | Phase II | Exemestane (active comparator); placebo-controlled | NCT00676663 ¹⁰⁸⁵ |
| | Entinostat | HDAC class 1 | Neuroendocrine tumors | Terminated (due to a lack of funding and drug supply) | Phase II | — | NCT03211988 |
| | Entinostat | HDAC class 1 | Cholangiocarcinoma, PDAC | Completed (exhibits promising efficacy) | Phase II | In combination with Nivolumab | NCT03250273 |
| | Entinostat | HDAC class 1 | Melanoma | Completed (unpublished) | Phase II | — | NCT00185302 |
| | Entinostat | HDAC class 1 | Lymphomas | Active, not recruiting | Phase II | In combination with Pembrolizumab | NCT03179930 |
| | Entinostat | HDAC class 1 | RCC | Active, not recruiting | Phase II | Interleukin-2 (active comparator)/in combination with Entinostat | NCT03501381 |
| | Entinostat | HDAC class 1 | Melanoma | Completed (exhibits preliminary antitumor effects) | Phase II | In combination with Pembrolizumab | NCT03765229 |
| | Entinostat | HDAC class 1 | Bladder cancer | Active, not recruiting | Phase II | Pembrolizumab (active comparator)/in combination with Entinostat | NCT03978624 |
| | Entinostat | HDAC class 1 | RCC | Active, not recruiting | Phase II | In combination with Nivolumab and Ipilimumab | NCT03552380 |
| | Entinostat | HDAC class 1 | AML, ALL | Completed (exhibits preliminary antitumor effects) | Phase II | In combination with Sargramostim | NCT00462605 |

Table 4. continued

| Type | Drug | Target(s) | Condition(s) | Status/outcome(s) | Phase(s) | Other intervention(s)/drug(s) | Study ID/reference(s) |
|------|--------------|--------------|--|--|------------|---|----------------------------------|
| | Entinostat | HDAC class 1 | NSCLC | Terminated (due to business reasons) | Phase II | In combination with Erlotinib | NCT00750698 |
| | Entinostat | HDAC class 1 | AML | Active, not recruiting | Phase II | In combination with Azacitidine | NCT01305499 |
| | Entinostat | HDAC class 1 | NSCLC | Terminated (due to slow accrual) | Phase II | In combination with Azacitidine | NCT01207726 |
| | Entinostat | HDAC class 1 | Colon cancer, rectal cancer | Completed (exhibits preliminary antitumor effects) | Phase II | In combination with Azacitidine | NCT01105377 |
| | Entinostat | HDAC class 1 | HR-positive BC | Completed (risks of treatment-associated adverse events are high) | Phase II | In combination with Aromatase inhibitor | NCT00828854 |
| | Entinostat | HDAC class 1 | NSCLC | Completed (combination is a promising tool in future exploration) | Phase II | In combination with Azacitidine and Nivolumab; Nivolumab with or without CC-486 300 (active comparator) | NCT01928576 |
| | Entinostat | HDAC class 1 | TNBC | Terminated (due to slow accrual) | Phase II | In combination with Anastrozole | NCT01234532 |
| | Entinostat | HDAC class 1 | NSCLC | Terminated | Phase II | In combination with Azacitidine and chemotherapy; chemotherapy (active comparator) | NCT01935947 |
| | Entinostat | HDAC class 1 | NSCLC | Completed (exhibits clinically meaningful benefit) | Phase I/II | In combination with Pembrolizumab | NCT02437136 ¹⁰⁸⁶ |
| | Entinostat | HDAC class 1 | CNS tumors | Recruiting | Phase I/II | In combination with Nivolumab; placebo-controlled | NCT03838042 ¹⁰⁸⁷ |
| | Entinostat | HDAC class 1 | RCC | Active, not recruiting (exhibits promising clinical activities according to disclosed data) | Phase I/II | In combination with Aldesleukin | NCT01038778 ¹⁰⁸⁸ |
| | Entinostat | HDAC class 1 | NSCLC | Completed (exhibits improvement in progression-free rates and overall survival) | Phase I/II | With or without Azacitidine | NCT00387465 ¹⁰⁸⁹ |
| | Entinostat | HDAC class 1 | NSCLC | Completed (the combination fails to improve the outcomes) | Phase I/II | Erlotinib (active comparator/in combination with Entinostat); placebo-controlled | NCT00602030 ¹⁰⁹⁰ |
| | Entinostat | HDAC class 1 | Ovarian cancer, peritoneal cancer, fallopian tube cancer | Completed (exhibits comparable efficacy and tolerability) | Phase I/II | Avelumab (active comparator/in combination with Entinostat); placebo-controlled | NCT02915523 |
| | Entinostat | HDAC class 1 | BC | Completed (exhibits clinical activity) | Phase I/II | Atezolizumab (active comparator/in combination with Entinostat); placebo-controlled | NCT02708680 |
| | Entinostat | HDAC class 1 | RCC | Suspended (due to major review underway) | Phase I/II | In combination with Atezolizumab and Bevacizumab | NCT03024437 |
| | Entinostat | HDAC class 1 | HPV-associated malignancies, small bowel cancer,colon cancer | Recruiting | Phase I/II | In combination with Bintrafusp Alfa/NHS-IL12, or NHS-IL12 | NCT04708470 |
| | Entinostat | HDAC class 1 | Solid tumors | Recruiting | Phase I/II | In combination with ZEN-3694 | NCT05053971 |
| | Entinostat | HDAC class 1 | Esophageal cancer | Suspended (due to revisions to design) | Phase I/II | In combination with Nivolumab, Montanide(R) ISA-51 VG Adjuvant, and H1299 Cell Lysates | NCT05898828 |
| | Entinostat | HDAC class 1 | ALL | Terminated (due to low accrual) | Phase I/II | In combination with Imatinib mesylate | NCT01383447 |
| | Entinostat | HDAC class 1 | BC | Active, not recruiting | Phase I/II | Umbrella study | NCT03280563 |
| | Entinostat | HDAC class 1 | HR-positive HER2-negative BC | Active, not recruiting (the combination fails to improve survival according to disclosed data) | Phase III | Exemestane/Goserelin/Goserelin acetate (active comparator/in combination with Entinostat); placebo-controlled | NCT02115282 ^{1091,1092} |
| | Entinostat | HDAC class 1 | HR-positive BC | Unknown (exhibits encouraging outcomes according to disclosed data) | Phase III | Exemestane (active comparator/in combination with Entinostat); placebo-controlled | NCT03538171 ⁴¹² |
| | Mocetinostat | HDAC class 1 | CRPC, BC, NSCLC | Terminated (due to terminated collaboration) | Phase I | In combination with Docetaxel | NCT00511576 |

Table 4. continued

| Type | Drug | Target(s) | Condition(s) | Status/outcome(s) | Phase(s) | Other intervention(s)/drug(s) | Study ID/reference(s) |
|------|--------------|--------------|--|---|------------|--|-----------------------------|
| | Mocetinostat | HDAC class 1 | MDS, lymphomas | Completed (unpublished) | Phase I | Given twice weekly | NCT00324194 |
| | Mocetinostat | HDAC class 1 | MDS, lymphomas | Completed (dose-limiting toxicities of fatigue, nausea, vomiting, and diarrhea observed at higher doses) | Phase I | Given three-times weekly | NCT00324129 ¹⁰⁹³ |
| | Mocetinostat | HDAC class 1 | NHL | Completed (unpublished) | Phase I | Given twice weekly | NCT00323934 |
| | Mocetinostat | HDAC class 1 | Squamous cell carcinoma of head and neck, squamous cell carcinoma of oral cavity | Withdrawn (due to a change in internal prioritization) | Phase I | In combination with Durvalumab | NCT02993991 |
| | Mocetinostat | HDAC class 1 | Rhabdomyosarcoma | Recruiting | Phase I | In combination with Vinorelbine | NCT04299113 |
| | Mocetinostat | HDAC class 1 | Lung cancer | Active, not recruiting | Phase I | In combination with Pembrolizumab and Guadecitabine | NCT03220477 |
| | Mocetinostat | HDAC class 1 | Melanoma | Terminated (exhibits favorable response rates but with high levels of toxicity according to disclosed data) | Phase I | In combination with Ipilimumab and Nivolumab | NCT03565406 ¹⁰⁹⁴ |
| | Mocetinostat | HDAC class 1 | Urothelial carcinoma | Completed (exhibits modest clinical activity) | Phase II | — | NCT02236195 ¹⁰⁹⁵ |
| | Mocetinostat | HDAC class 1 | HL | Terminated (exhibits single-agent clinical activity with manageable toxicity according to disclosed data) | Phase II | — | NCT00358982 ¹⁰⁹⁶ |
| | Mocetinostat | HDAC class 1 | Lymphoma | Completed (exhibits limited single-agent activity in DLBCL and FL but long-term clinical benefit) | Phase II | — | NCT00359086 ¹⁰⁹⁷ |
| | Mocetinostat | HDAC class 1 | AML, MDS | Terminated (due to terminated collaboration) | Phase II | Azacitidine (active comparator/in combination with Mocetinostat) | NCT00666497 |
| | Mocetinostat | HDAC class 1 | NHL, HL | Terminated (due to terminated collaboration) | Phase II | In combination with Azacitidine | NCT00543582 |
| | Mocetinostat | HDAC class 1 | CLL | Completed (exhibits limited activity) | Phase II | — | NCT00431873 ¹⁰⁹⁸ |
| | Mocetinostat | HDAC class 1 | AML, MDS | Terminated (due to the re-evaluation of clinical development program) | Phase II | — | NCT00374296 |
| | Mocetinostat | HDAC class 1 | Leiomyosarcoma | Completed (exhibits insufficient activity) | Phase II | In combination with Gemcitabine | NCT02303262 |
| | Mocetinostat | HDAC class 1 | NSCLC | Terminated (due to sponsors' decision) | Phase II | In combination with Nivolumab; Nivolumab with Sitravatinib or Glesatinib (active comparator) | NCT02954991 |
| | Mocetinostat | HDAC class 1 | Malignant tumors | Completed (exhibits significant toxicities in advanced pancreatic cancer) | Phase I/II | In combination with Gemcitabine | NCT00372437 ¹⁰⁹⁹ |
| | Mocetinostat | HDAC class 1 | DLBCL, lymphomas | Terminated (due to slow accrual) | Phase I/II | — | NCT02282358 ¹¹⁰⁰ |
| | Mocetinostat | HDAC class 1 | NSCLC, solid tumors | Terminated (due to sponsors' decision) | Phase I/II | In combination with Durvalumab | NCT02805660 ¹¹⁰¹ |
| | Mocetinostat | HDAC class 1 | MDS, AML | Completed (unpublished) | Phase I/II | — | NCT00324220 |
| | Mocetinostat | HDAC class 1 | MDS | Completed (unpublished) | Phase I/II | In combination with Azacitidine | NCT02018926 |
| | Mocetinostat | HDAC class 1 | HL | Completed (exhibits preliminary clinical activity) | Phase I/II | In combination with Brentuximab vedotin | NCT02429375 |
| | Domatinostat | LSD1/HDAC | Hematologic malignancies | Completed (exhibits safety and early signs of antitumor activity) | Phase I | — | NCT01344707 ¹¹⁰² |

Table 4. continued

| Type | Drug | Target(s) | Condition(s) | Status/outcome(s) | Phase(s) | Other intervention(s)/drug(s) | Study ID/reference(s) |
|------|-----------------------|------------------------|--|---|------------|--|---------------------------------|
| | Domatinostat | LSD1/ HDAC | Gastrointestinal cancer | Unknown (exhibits an acceptable safety profile according to disclosed data) | Phase II | In combination with Avelumab | NCT03812796 ¹¹⁰³ |
| | Domatinostat | LSD1/ HDAC | Merkel cell carcinoma | Withdrawn (due to sponsors' decision) | Phase II | In combination with Avelumab | NCT04874831 |
| | Domatinostat | LSD1/ HDAC | Merkel cell carcinoma | Completed (unpublished) | Phase II | In combination with Avelumab | NCT04393753 |
| | Domatinostat | LSD1/ HDAC | Melanoma | Active, not recruiting (Domatinostat addition fails to increase treatment efficacy according to disclosed data) | Phase I/II | Nivolumab (active comparator)/in combination with Domatinostat; in combination with Nivolumab and Ipilimumab | NCT04133948 ⁴²⁴ |
| | CUDC101 | EGFR/ HER2/ HDAC | Solid tumors | Terminated | Phase I | — | NCT01702285 |
| | CUDC101 | EGFR/ HER2/ HDAC | Squamous cell carcinoma of head and neck, gastric cancer, BC, HCC, NSCLC | Completed (exhibits acceptable safety) | Phase I | — | NCT01171924 |
| | CUDC101 | EGFR/ HER2/ HDAC | Squamous cell carcinoma of head and neck | Completed (the combination exhibits promising feasibility) | Phase I | In combination with Cisplatin and radiation therapy | NCT01384799 ⁴²⁰ |
| | CUDC101 | EGFR/ HER2/ HDAC | Solid tumors | Completed (exhibits good tolerability and antitumor activity) | Phase I | — | NCT00728793 ⁴²¹ |
| | CUDC-907 | PI3K/ HDAC | Lymphoma | Completed (exhibits tolerable safety profile and durable antitumor activity) | Phase I | With or without Rituximab or Venetodax | NCT01742988 ^{422,1104} |
| | CUDC-907 | PI3K/ HDAC | Diffuse intrinsic pontine glioma, medulloblastoma, high-grade glioma | Active, not recruiting | Phase I | — | NCT03893487 |
| | CUDC-907 | PI3K/ HDAC | TNBC, ovarian cancer, NUT carcinoma | Completed (unpublished) | Phase I | — | NCT02307240 |
| | CUDC-907 | PI3K/ HDAC | CNS tumors, lymphoma | Active, not recruiting | Phase I | — | NCT02909777 |
| | CUDC-907 | PI3K/ HDAC | DLBCL | Completed (exhibits preliminary antitumor effects) | Phase II | — | NCT02674750 ⁴²³ |
| | CUDC-907 | PI3K/ HDAC | Cushing disease | Not yet recruiting | Phase II | — | NCT05971758 |
| | CUDC-907 | PI3K/ HDAC | Thyroid cancer | Terminated (due to investigator's reasons) | Phase II | — | NCT03002623 |
| | Sodium phenylbutyrate | PRKCA/ HDAC | MCAD deficiency | Recruiting | Phase II | — | NCT06069375 |
| | Tinostamustine | DNA/ HDAC | Melanoma | Unknown | Phase I | — | NCT03903458 |
| | Tinostamustine | DNA/ HDAC | Glioblastoma multiforme | Active, not recruiting | Phase I | — | NCT05432375 |
| | Tinostamustine | DNA/ HDAC | MM, HL, CTCL | Active, not recruiting | Phase I | — | NCT02576496 |
| | Tinostamustine | DNA/ HDAC | MGMT-promoter unmethylated glioblastoma | Completed (unpublished) | Phase I | With or without radiation therapy | NCT03452930 |
| | Tinostamustine | DNA/ HDAC | DLBCL | Withdrawn (given the safety data on drug) | Phase I | In combination with Pembrolizumab and Rituximab | NCT04279938 |
| | Tinostamustine | DNA/ HDAC | MM | Terminated (due to sponsors' decision based on adverse events) | Phase I/II | ASCT | NCT03687125 |

Table 4. continued

| Type | Drug | Target(s) | Condition(s) | Status/outcome(s) | Phase(s) | Other intervention(s)/drug(s) | Study ID/reference(s) |
|---------------|----------------|--------------------------|---|--|----------------|---|-----------------------------|
| HDAC agonist | Tinostamustine | DNA/HDAC | SCLC, soft tissue sarcoma, TNBC, ovarian cancer, endometrial cancer | Completed (unpublished) | Phase I/II | — | NCT03345485 |
| | GSK3117391 | Esterase-sensitive motif | RA | Terminated (due to development portfolio) | Phase II | Placebo-controlled | NCT02965599 |
| | Tefinostat | Esterase-sensitive motif | Hematologic malignancies | Completed (exhibits early signs of efficacy and absence of significant toxicity) | Phase I | — | NCT00820508 ¹¹⁰⁵ |
| | Tefinostat | Esterase-sensitive motif | HCC | Unknown | Phase I/II | — | NCT02759601 |
| | Theophylline | Pan-HDAC | COPD | Completed (exhibits an increase in total HDAC activity and potential clinical benefit) | Phase II | With or without Fluticasone propionate | NCT00241631 ¹¹⁰⁶ |
| | Theophylline | Pan-HDAC | COPD | Completed (fails to enhance the anti-inflammatory properties of ICS) | Phase III | In combination with ICS; placebo-controlled | NCT01599871 ⁴²⁶ |
| | Theophylline | Pan-HDAC | Bronchiectasis | Completed (unpublished) | Phase IV | With or without Formoterol-budesonide | NCT01769898 |
| | Theophylline | Pan-HDAC | COPD | Completed (exhibits an increase in total HDAC activity and potential clinical benefit) | Not applicable | With or without standard therapy | NCT00671151 ¹¹⁰⁷ |
| | Resveratrol | SIRT1 | T2DM | Completed (H3K56ac is located in the key position between SIRT1 and T2DM) | Phase III | Placebo-controlled | NCT02244879 ¹¹⁰⁸ |
| | ZEN-3694 | Pan-BET | Colorectal carcinoma | Recruiting | Phase I | In combination with Capecitabine | NCT05803382 |
| BET inhibitor | ZEN-3694 | Pan-BET | Endometrial carcinoma | Recruiting | Phase I | In combination with Tuvusertib | NCT05950464 |
| | ZEN-3694 | Pan-BET | Platinum-resistant ovarian carcinoma | Recruiting | Phase I | In combination with Nivolumab or Nivolumab plus Ipilimumab | NCT04840589 |
| | ZEN-3694 | Pan-BET | Colorectal carcinoma | Recruiting | Phase I | In combination with Cetuximab and Encorafenib | NCT06102902 |
| | ZEN-3694 | Pan-BET | BC, NUT carcinoma | Recruiting | Phase I | In combination with Abemaciclib | NCT05372640 |
| | ZEN-3694 | Pan-BET | Ovarian cancer, solid tumors | Recruiting | Phase I | In combination with Niraparib | NCT06161493 |
| | ZEN-3694 | Pan-BET | Ovarian cancer, solid tumors | Recruiting | Phase I | In combination with Binimetinib | NCT05111561 |
| | ZEN-3694 | Pan-BET | CRPC | Completed (unpublished) | Phase I | — | NCT02705469 |
| | ZEN-3694 | Pan-BET | CRPC | Recruiting | Phase II | In combination with Enzalutamide and Pembrolizumab | NCT04471974 |
| | ZEN-3694 | Pan-BET | Solid tumors | Recruiting | Phase II | In combination with Talazoparib | NCT05327010 |
| | ZEN-3694 | Pan-BET | CRPC | Recruiting | Phase II | Enzalutamide (active comparator/in combination with ZEN-3694) | NCT04986423 |
| | ZEN-3694 | Pan-BET | Squamous cell lung cancer | Recruiting | Phase II | — | NCT05607108 |
| | ZEN-3694 | Pan-BET | Ovarian cancer, peritoneal cancer, fallopian tube cancer | Recruiting | Phase II | In combination with Talazoparib | NCT05071937 |
| | ZEN-3694 | Pan-BET | TNBC | Terminated (based on results from an interim futility analysis and not due to safety concerns) | Phase II | In combination with Talazoparib | NCT03901469 ⁴⁸⁵ |
| | ZEN-3694 | Pan-BET | NUT carcinoma | Recruiting | Phase I/II | In combination with Cisplatin and Etoposide | NCT05019716 |
| | ZEN-3694 | Pan-BET | Solid tumors, lymphomas | Recruiting | Phase I/II | In combination with Entinostat | NCT05053971 |
| | ZEN-3694 | Pan-BET | CRPC | Completed (exhibits acceptable safety and efficacy) | Phase I/II | In combination with Enzalutamide | NCT02711956 ⁴⁸⁶ |
| | ZEN-3694 | Pan-BET | CRPC | Enrolling by invitation | Phase I/II | In combination with Enzalutamide | NCT04145375 |
| | Trotabresib | Pan-BET | HER2-positive BC with CNS metastasis and leptomeningeal disease | Withdrawn (due to sponsors' decision) | Phase I | In combination with Vinorelbine and radiation therapy | NCT06137651 |
| | | | | | | | |
| | | | | | | | |

Table 4. continued

| Type | Drug | Target(s) | Condition(s) | Status/outcome(s) | Phase(s) | Other intervention(s)/drug(s) | Study ID/reference(s) |
|-------------|----------|---|--|-------------------|---|----------------------------------|----------------------------------|
| Trotabresib | Pan-BET | Astrocytoma, glioblastoma | Terminated (due to a change in business objectives) | Phase I | — | — | NCT04047303 ¹¹⁰⁹ |
| | Pan-BET | Solid tumors, NHL | Active, not recruiting (exhibits good tolerability and single-agent activity in advanced solid tumors according to disclosed data) | Phase I | — | — | NCT03220347 ^{1110,1111} |
| Trotabresib | Pan-BET | Pediatric cancer | Active, not recruiting | Phase I | BMS-986158 (active comparator) | NCT03936465 | |
| Trotabresib | Pan-BET | Solid tumors | Withdrawn (due to a change in business objectives) | Phase I | — | NCT05678283 | |
| Trotabresib | Pan-BET | Glioblastoma | Active, not recruiting | Phase I | In combination with Temozolomide and radiation therapy; radiation therapy (active comparator) | NCT04324840 | |
| Alobresib | Pan-BET | Solid tumors, lymphomas, HR-positive BC | Completed (reports pharmacokinetics and pharmacodynamics) | Phase I | With or without Exemestane or Fulvestrant | NCT02392611 | |
| Alobresib | Pan-BET | CRPC | Terminated (exhibits acceptable tolerability) | Phase I/II | With or without Enzalutamide | NCT02607228 | |
| GSK3358699 | Pan-BET | Healthy volunteers | Terminated (due to strategic reasons) | Phase I | Placebo-controlled | NCT03426995 ¹¹¹² | |
| TEN-010 | Pan-BET | AML, MDS | Completed (exhibits insufficient activity as a single agent) | Phase I | — | NCT02308761 ¹¹¹³ | |
| TEN-010 | Pan-BET | Solid tumors | Completed (exhibits evidence of target engagement and preliminary single-agent activity) | Phase I | — | NCT01987362 ¹¹¹⁴ | |
| TEN-010 | Pan-BET | Ovarian cancer, TNBC | Terminated (due to development portfolio) | Phase I | In combination with Atezolizumab | NCT03292172 | |
| TEN-010 | Pan-BET | MM | Completed (exhibits infrequent and short duration of clinical response rates) | Phase I | With or without Daratumumab | NCT03068351 ¹¹¹⁵ | |
| TEN-010 | Pan-BET | DLBCL | Completed (unpublished) | Phase I | In combination with Venetoclax and Rituximab | NCT03255096 ¹¹¹⁶ | |
| ODM-207 | Pan-BET | Solid tumors | Completed (exhibits safety at doses up to 2 mg/kg but has a narrow therapeutic window) | Phase I/II | — | NCT03035591 ¹¹¹⁷ | |
| ABBV-744 | Pan-BET | Myelofibrosis | Terminated (due to strategic reasons) | Phase I | — | NCT03360006 | |
| ABBV-744 | Pan-BET | AML | Active, not recruiting | Phase I | With or without Ruxolitinib or Navitoclax | NCT04454658 | |
| Birabresib | BRD2/3/4 | Solid tumors | Completed (exhibits a favorable safety profile with clinical activity in NUT carcinoma) | Phase I | — | NCT02259114 ¹¹¹⁸ | |
| Birabresib | BRD2/3/4 | AML, ALL, DLBCL, MM | Completed (exhibits evidence of clinical activity though does not meet objective response criteria in non-leukemia cohort) | Phase I | — | NCT01713582 ^{1119,1120} | |
| Birabresib | BRD2/3/4 | AML, DLBCL | Terminated (due to limited efficacy) | Phase I | — | NCT02698189 | |
| Birabresib | BRD2/3/4 | NUT carcinoma, TNBC, NSCLC, CRPC | Terminated (due to limited efficacy) | Phase I | — | NCT02698176 | |
| Birabresib | BRD2/3/4 | GBM | Terminated (due to limited efficacy) | Phase II | — | NCT02296476 | |
| Birabresib | BRD2/3/4 | AML | Withdrawn | Phase I/II | Azacitidine (active comparator/in combination with Birabresib) | NCT02303782 | |
| Molibresib | BRD2/3/4 | Solid tumors, lymphomas | Withdrawn (due to disapproved protocol) | Phase I | In combination with Entinostat | NCT03925428 | |
| Molibresib | BRD2/3/4 | NUT carcinoma, solid tumors | Completed (exhibits acceptable safety) | Phase I | — | NCT01587703 ^{1121,1122} | |
| Molibresib | BRD2/3/4 | HR-positive HER2-negative BC | Terminated (due to meeting protocol-defined BC futility) | Phase I | In combination with Fulvestrant; placebo-controlled | NCT02964507 ¹¹²³ | |
| Molibresib | BRD2/3/4 | CRPC | Terminated (due to meeting protocol-defined futility) | Phase I | In combination with Abiraterone plus Prednisone or Enzalutamide | NCT03150056 | |
| Molibresib | BRD2/3/4 | Healthy volunteers | Completed (CYP3A enzymes play a major role in the elimination of Molibresib) | Phase I | In combination with Itraconazole or Rifampicin | NCT02706535 ¹¹²⁴ | |

Table 4. continued

| Type | Drug | Target(s) | Condition(s) | Status/outcome(s) | Phase(s) | Other intervention(s)/drug(s) | Study ID/reference(s) |
|------|-------------|------------------|--|--|---------------|---|----------------------------------|
| | Molibresib | BRD2/3/4 | Hematologic malignancies | Completed (exhibits antitumor activity but is limited by gastrointestinal and thrombocytopenia toxicities) | Phase II | — | NCT01943851 ¹¹²⁵ |
| | Molibresib | BRD2/3/4 | SCLC, solid tumors | Withdrawn (due to reevaluation) | Phase II | In combination with Trametinib | NCT03266159 |
| | Molibresib | BRD2/3/4 | NUT carcinoma | Withdrawn (due to disapproved protocol) | Phase I/II | In combination with Cisplatin, Etoposide, and Etoposide phosphate | NCT04116359 |
| | Mivebresib | BRD2/4/T | Myelofibrosis | Terminated (due to strategic reasons) | Phase I | With or without Ruxolitinib or Navitoclax | NCT04480086 |
| | Mivebresib | BRD2/4/T | BC, NSCLC, AML, MM PC, SCLC, NHL | Completed (exhibits good tolerability and potential efficacy in advanced solid tumors) | Phase I | With or without Venetoclax | NCT02391480 ^{1126,1127} |
| | BAY1238097 | BRD4-BD1; BRD2/3 | Malignant tumors | Terminated | Phase I | — | NCT02369029 |
| | PLX-2853 | BRD4 | AML, MDS | Completed (unpublished) | Phase I | — | NCT03787498 |
| | PLX-2853 | BRD4 | Malignant tumors | Completed (unpublished) | Phase I | — | NCT03297424 |
| | PLX-2853 | BRD4 | CRPC | Terminated (due to business realignment) | Phase I/II | In combination with Abiraterone acetate plus Prednisone, or Olaparib | NCT04556617 |
| | PLX-2853 | BRD4 | Uveal melanoma | Withdrawn (drug company has withdrawn support) | Phase I/II | In combination with Trametinib | NCT05677373 |
| | PLX-2853 | BRD4 | Platinum-resistant ovarian carcinoma | Terminated (due to business realignment) | Phase I/II | With or without Carboplatin | NCT04493619 |
| | INCB054329 | BRD4 | Solid tumors, hematologic malignancies | Terminated (due to interindividual pharmacokinetic variability) | Phase I/II | — | NCT02431260 ¹¹²⁸ |
| | SYHA1801 | BRD4 | Solid tumors | Unknown | Phase I | — | NCT04309968 |
| | CC-95775 | BRD4 | AML, MDS, NHL | Completed (unpublished) | Phase I | With or without Azacitidine | NCT02543879 |
| | CC-95775 | BRD4 | Solid tumors, NHL | Completed (unpublished) | Phase I | — | NCT04089527 |
| | PLX51107 | BRD4 | Solid tumors, hematologic malignancies | Terminated (due to business reasons) | Phase I | — | NCT02683395 |
| | PLX51107 | BRD4 | AML, MDS | Completed (unpublished) | Phase I | In combination with Azacitidine | NCT04022785 |
| | PLX51107 | BRD4 | Acute GVHD | Terminated (due to sponsors' decision) | Phase I/II | — | NCT04910152 |
| | BMS-986158 | BRD4 | Pediatric Cancer | Active, not recruiting | Phase I | BMS-986378 (active comparator) | NCT03936465 |
| | BMS-986158 | BRD4 | Myelofibrosis | Active, not recruiting | Phase I/II | In combination with Ruxolitinib or Fedratinib | NCT04817007 |
| | BMS-986158 | BRD4 | Solid tumors, hematologic malignancies | Completed (exhibits insufficient evidence of improvement) | Phase I/II | With or without Nivolumab | NCT02419417 |
| | BMS-986158 | BRD4 | MM | Recruiting | Phase I/II | In combination with Tazemetostat plus Dexamethasone or BMS-986158 plus Dexamethasone or Trametinib plus Dexamethasone, or Dexamethasone | NCT05372354 |
| | AZD5153 | BRD4 | Solid tumors, lymphomas | Completed (exhibits tolerable safety as monotherapy and in combination) | Phase I | With or without Olaparib | NCT03205176 ¹¹²⁹ |
| | AZD5153 | BRD4 | NHL, DLBCL | Completed (unpublished) | Phase I | In combination with Acalabrutinib, Acalabrutinib, Rituximab, plus Hu5F9-G4 (active comparator) | NCT03527147 |
| | AZD5153 | BRD4 | AML | Recruiting | Phase I/II | Umbrella study | NCT03013998 |
| | B1894999 | BRD4-BD1/BD2 | Malignant tumors, NUT carcinoma | Completed (exhibits preliminary antitumor effects and reports the maximum tolerated dose at different cohorts) | Phase I | — | NCT02516553 ¹¹³⁰ |
| | Apabetalone | BRD4-BD2 | PAH | Completed (exhibits good tolerability and clinical benefits) | Early phase I | — | NCT03655704 ¹¹³¹ |
| | Apabetalone | BRD4-BD2 | PAH | Not yet recruiting | Phase II | Placebo-controlled | NCT04915300 |
| | Apabetalone | BRD4-BD2 | Atherosclerosis, CAD | Completed (exhibits good tolerability) | Phase II | Placebo-controlled | NCT01058018 ¹¹³² |

Table 4. continued

| Type | Drug | Target(s) | Condition(s) | Status/outcome(s) | Phase(s) | Other intervention(s)/drug(s) | Study ID/reference(s) |
|------|-------------|---------------|--|---|--------------|--|--------------------------------------|
| | Apabetalone | BRD4-BD2 | T2DM | Completed (exhibits potential to against the disease development) | Phase II | Placebo-controlled | NCT01728467 ¹¹³³ |
| | Apabetalone | BRD4-BD2 | Dyslipidemia, CAD | Completed (exhibits good tolerability) | Phase II | Placebo-controlled | NCT01423188 ¹¹³⁴ |
| | Apabetalone | BRD4-BD2 | CAD | Completed (exhibits no significant improvement) | Phase II | Placebo-controlled | NCT01067820 ^{1134–1137} |
| | Apabetalone | BRD4-BD2 | Dyslipidemia, CAD | Terminated | Phase II | In combination with Rosuvastatin or Atorvastatin | NCT01863225 |
| | Apabetalone | BRD4-BD2 | Chronic kidney failure | Not yet recruiting | Phase I/II | Placebo-controlled | NCT03160430 |
| | Apabetalone | BRD4-BD2 | Fabry disease | Withdrawn (due to changed development priorities) | Phase I/II | — | NCT03228940 |
| | Apabetalone | BRD4-BD2 | Healthy volunteers, dyslipidemia, atherosclerosis, CAD | Completed (unpublished) | Phase I/II | Placebo-controlled | NCT00768274 |
| | Apabetalone | BRD4-BD2 | T2DM, CAD | Completed (fails to reduce the risk of major adverse cardiovascular events) | Phase III | In combination with Rosuvastatin or Atorvastatin; | NCT02586155 ^{477,1137–1142} |
| | Apabetalone | BRD4-BD2 | COVID-19 infection | Terminated (fails to recruit subjects) | Phase II/III | Standard of care (active comparator/in combination with Apabetalone) | NCT04894266 |
| | NUV-868 | BRD4-BD2 | Solid tumors | Recruiting | Phase I/II | With or without Olaparib or Enzalutamide | NCT05252390 |
| | Pelabresib | BRD4-BD1 | Malignant tumors | Completed (unpublished) | Phase I | — | NCT05391022 |
| | Pelabresib | BRD4-BD1 | Lymphoma | Completed (exhibits good tolerability and inhibitory effects) | Phase I | — | NCT01949883 ¹¹⁴³ |
| | Pelabresib | BRD4-BD1 | MM | Completed (unpublished) | Phase I | — | NCT02157636 |
| | Pelabresib | BRD4-BD1 | Peripheral nerve tumor | Withdrawn (due to a lack of enrollment) | Phase II | — | NCT02986919 |
| | Pelabresib | BRD4-BD1 | Myelofibrosis, AML, MDS, myeloproliferative disorders | Active, not recruiting (exhibits potential disease-modifying activity in myelofibrosis according to disclosed data) | Phase I/II | With or without Ruxolitinib | NCT02158858 ^{484,1144} |
| | Pelabresib | BRD4-BD1 | Malignant tumors | Not yet recruiting | Phase III | Placebo-controlled | NCT06401356 |
| | Pelabresib | BRD4-BD1 | Myelofibrosis | Active, not recruiting | Phase III | In combination with Ruxolitinib, placebo-controlled | NCT04603495 |
| | TQB3617 | BET | Malignant tumors | Unknown | Phase I | — | NCT05110807 |
| | TQB3617 | BET | Myelofibrosis | Recruiting | Phase I/II | In combination with TQ05105 | NCT06122831 |
| | TQB3617 | BET | Esophageal squamous cell carcinoma | Not yet recruiting | Phase I/II | In combination with TQB2618, Paclitaxel, and Cisplatin, or Paclitaxel plus Cisplatin, or TQB2618 plus Penpulimab | NCT05834543 |
| | EP31670 | P300/CBP/ BET | CRPC, NUT carcinoma | Recruiting | Phase I | — | NCT05488548 ¹¹⁴⁵ |

ABL acute biphenotypic leukemia, *ALL* acute lymphoblastic leukemia, *AML* acute myeloid leukemia, *ASCT* autologous stem cell transplant, *BC* breast cancer, *BD* bromodomain, *BET* bromodomain and extraterminal domain, *BRD* bromodomain containing protein, *CAD* coronary artery disease, *CBP* cyclic adenosine monophosphate-responsive element-binding protein (CREB)-binding protein, *CLL* chronic lymphocytic leukemia, *CML* chronic myeloid leukemia, *CMMML* chronic myelomonocytic leukemia, *CNS* central nervous system, *COPD* chronic obstructive pulmonary disease, *COVID-19* corona virus disease 2019, *CRPC* Castration-resistant prostate cancer, *CTCL* cutaneous T cell lymphoma, *CYP3A* cytochrome P4503A, *DLBCL* diffuse large B cell lymphoma, *EBV* Epstein-Barr virus, *EGFR* epidermal growth factor receptor, *FL* follicular lymphoma, *GBM* glioblastoma multiforme, *GCSF* granulocyte colony-stimulating factor, *GVHD* graft versus host disease, *HCC* hepatocellular carcinoma, *HDAC* histone deacetylase, *HCV* hepatitis C virus, *HER2* human epidermal growth factor receptor 2, *HIV* human immunodeficiency virus, *H3K56ac* histone 3 acetylation at the 56 lysine residue, *HL* Hodgkin lymphoma, *HPV* human papilloma virus, *HR* hormone receptor, *ICS* inhaled corticosteroid, *KAT* lysine acetyltransferase, *LSI* lysine specific demethylase 1, *MCL* mantle cell lymphoma, *MCAD* medium-chain acyl-CoA dehydrogenase, *MDS* myelodysplastic syndrome, *MF* mycosis fungoides, *MGMT* O⁶-methylguanine-DNA methyltransferase, *MM* multiple myeloma, *NHL* non-Hodgkin lymphoma, *N5CLC* non-small cell lung cancer, *NUT* nuclear protein in testis carcinoma, *PAH* pulmonary arterial hypertension, *PBC* primary biliary cholangitis, *PDAC* pancreatic ductal adenocarcinoma, *PI3K* phosphoinositide 3-kinase, *PRKCA* protein kinase C alpha, *PTCL* peripheral T cell lymphoma, *PTLD* post-transplant lymphoproliferative disorder, *RA* rheumatoid arthritis, *RCC* renal cell carcinoma, *SIRT1* Sirtuin1, *T2DM* type 2 diabetes, *TNBC* triple-negative breast cancer

cancer and other advanced solid tumors to assess its maximum tolerated oral dose (NCT05488548). PRI-724 effectively disrupts the interaction between β -catenin and CBP, ameliorating various diseases by inhibiting the Wnt/ β -catenin signaling pathway.³⁹⁵ Its safety, tolerability, and antifibrotic effects have been further evaluated in two completed clinical trials among patients with hepatitis C virus (HCV)- and HBV-induced cirrhosis.^{396,397} However, PRI-724 fails to exhibit sufficient evidence of improvement in hepatic function according to existing data.³⁹⁶ Lastly, PF-07248144, a selective inhibitor of KAT6 (a member of the MOZ/MORF family), is currently under clinical investigation for the treatment of advanced breast cancer (NCT04606446). In summary, KATs represent compelling targets for therapeutic strategies, and developing novel and high-quality inhibitors with improved safety and efficacy is reaching an exciting phase.

Targeting the eraser of histone acetylation: HDAC. Zn^{2+} -dependent classical HDACs and nicotinamide adenine dinucleotide (NAD)⁺-dependent HDACs (sirtuins) are crucial for dynamic deacetylation modifications on histones and non-histone proteins, playing significant roles in ontogeny and tumorigenesis. Despite the HDAC family's broad substrate range *in vitro*, their specific subcellular localization restricts their biological functions and target proteins. Using inhibitors and agonists of HDACs and sirtuins to correct abnormal acetylation patterns is a promising therapeutic strategy.^{398,399} Notably, the therapeutic effectiveness of these interventions, in an epigenetic-dependent manner, hinges on the participation of target enzymes in histone deacetylation.

HDAC inhibitors: HDAC inhibitors are designed based on the spatial structure of their targets, characterized by highly conserved and homologous catalytic domains, including a catalytic channel, a zinc cation, and secondary pockets. Most HDAC inhibitors consist of a surface binding region, binding to the catalytic channel, and a zinc-binding group along with the linker, chelating the zinc ion.⁴⁰⁰ Four main categories of HDAC inhibitors are extensively studied: pan-inhibitors, selective inhibitors, multitarget agents, and PROTACs-based HDAC degraders.³⁹⁸ We will now discuss the recent applications of these HDAC inhibitors in clinical trials.

Four FDA-approved HDAC inhibitors—vorinostat, romidepsin, belinostat, and panobinostat—demonstrate a pan-inhibitory effect on almost all HDAC members and have made significant progress in treating some hematological malignancies. This success has fueled enthusiasm for developing additional pan-inhibitors to expand the clinical indications of these drugs. Currently, several pan-inhibitors, including ivaltinostat (CG200745), AR-42, abexinostat (PCI-24781), bishthianostat (CF-367), and sodium valproate, are under clinical trials for various tumors. The phase II study on ivaltinostat for advanced pancreatic ductal adenocarcinoma reports enhanced sensitivity of tumor cells to gemcitabine and erlotinib, presenting it as a potential treatment option.⁴⁰¹ Another phase II study aims to determine the maximum tolerated dose and dose-limiting toxicity of ivaltinostat in combination with gemcitabine and erlotinib in patients with advanced pancreatic cancer, although clinical data have not been publicly disclosed, suggesting potential challenges (NCT02737228). In phase I trials, single-agent AR-42 has shown promise in treating type 2-associated meningiomas and schwannomas, with patients exhibiting good tolerance and therapeutic potential.^{402,403} However, a phase I trial focusing on advanced sarcoma and kidney cancer was terminated early due to observed dose-limiting toxicities in six patients (NCT02795819). Abexinostat, an oral small pan-inhibitor, whether used as monotherapy or in combination with chemotherapeutic agents, has shown promising therapeutic potential and acceptable safety profiles in solid tumors and hematological malignancies.^{404–406} Notably, a

phase III study on abexinostat for locally advanced or metastatic renal cell carcinoma is ongoing in various regions, highlighting its potential as a clinical candidate (NCT03592472). Bishthianostat, a novel bithiazole-derived pan-HDAC inhibitor, was studied in phase 1a clinical trial.⁴⁰⁷ Although preliminary data suggested modest efficacy and tolerability as a single agent in patients with R/R MM, this study has been terminated for undisclosed reasons (NCT03618602).

The non-selective inhibition characteristic of pan-HDAC inhibitors often leads to a broad spectrum of adverse effects and off-target toxicities, which restrict their widespread clinical application.⁴⁰⁸ Given the diverse roles of different HDAC classes, there is increasing interest in developing selective HDAC inhibitors, viewed as promising alternatives with better tolerance.^{409,410} However, due to a lack of evidence supporting the involvement of HDAC5/6/7/8/10 in histone deacetylation, selective inhibitors targeting these enzymes are not typically included in summaries of epigenetic-targeted drugs.⁴¹¹ Chidamide and givinostat, both FDA-approved selective inhibitors, have shown superior therapeutic efficacy and safety profiles. Givinostat, in particular, has promisingly expanded the clinical indications of HDAC inhibitors to include non-tumor diseases. Beyond these marketed drugs, several selective inhibitors have entered clinical practice. Notably, four such inhibitors are undergoing phase III clinical trials: pracinostat (NCT03151408), entinostat,⁴¹² magnesium valproate (NCT00533299), and tacedinaline (NCT00005093). Among these, only the phase III trial of entinostat combined with exemestane in treating hormone receptor-positive advanced breast cancer has shown satisfactory efficacy and manageable toxicities.⁴¹² However, among patients with other types of tumors such as colorectal carcinoma, lung cancer, endometrial endometrioid adenocarcinoma, and hematologic malignancies, entinostat fails to improve survival despite exhibiting good clinical efficacy.^{413–416} Importantly, according to an early-terminated, phase I clinical trial that evaluated the combination of entinostat, hydroxychloroquine, and regorafenib, the drug regimen among patients with metastatic colorectal carcinoma was poorly tolerated, with higher risks of weight loss, fatigue, and anorexia.⁴¹³ Despite these advancements, selective inhibitors still face significant developmental challenges as they emerge as the next generation of HDAC inhibitors.

Recently, multitarget agents-based HDAC inhibitors have gained attention and have been posited to perform versatile roles in disease treatment.^{417–419} Various such agents, including those dual-targeting HDACs and kinases, receptors, DNA, transcriptional factors, and apoptosis-related proteins, are under preclinical investigation. Examples include curcumin (previously mentioned as a DNMT/METTL3 inhibitor), CUDC-101, tinostamustine (EDO-S101), fimepinostat (CUDC-907), domatinostat (4SC-202), and dacinostat (NVP-LAQ824, LAQ824), all of which are involved in various clinical trials.³⁹⁸ It has been widely reported that these multitarget agents enhance safety and reduce drug resistance in various diseases, both as monotherapy and in combination with radiotherapy or chemotherapy.^{420–423} However, emerging research offers a contrasting perspective. In a recent phase Ib clinical trial focusing on domatinostat (a dual inhibitor of LSD1/HDAC) in patients with advanced melanoma, the drug failed to enhance the efficacy of treatments targeting anti-PD-1 and cytotoxic T lymphocyte-associated antigen-4 (CTLA-4) while unexpected severe skin toxicity was observed.⁴²⁴ Furthermore, as current knowledge about these multitarget agents is still primarily derived from early-stage clinical trials, extensive investigations are necessary to validate their therapeutic value in a broader population.

HDAC agonists: While HDAC inhibitors have been extensively studied, research on HDAC agonists has been less prevalent. However, the therapeutic value of these agents in specific diseases has been demonstrated. Theophylline, used initially as an inhibitor

of phosphodiesterase and adenosine receptors in treating asthma and chronic obstructive pulmonary disease, has recently shown activated effects on HDAC in low doses. These effects synergistically enhance the anti-inflammatory properties of cortisol in asthma and chronic obstructive pulmonary disease treatments.⁴²⁵ Nonetheless, a phase III clinical study revealed that additional administration of low-dose theophylline, along with inhaled long-acting β_2 -agonists and corticosteroids, failed to enhance HDAC activity in vivo. This left no significant difference from the anti-inflammatory properties of standard therapy.⁴²⁶

SIRT inhibitors and agonists: The sirtuin family's role in developing various diseases, including inflammation, cardiovascular diseases, metabolic disorders, neurodegenerative diseases, and cancer, underscores the importance of exploring molecules that modulate their activity. Notably, SIRT2 is involved in the deacetylation of histone H4 during the G2/M transition and mitosis but is predominantly found in the cytosol, where it participates in non-histone deacetylation.^{427,428} SIRT3-5 are mainly located in mitochondria and possess a mitochondrial targeting sequence,⁴²⁹ while SIRT7 is primarily found in the nucleus, though fewer studies have addressed molecules that regulate its activity.⁴³⁰ Consequently, the inhibitors and agonists of SIRT1 and SIRT6 are highlighted as promising epigenetics-targeted drugs with significant potential.

From a mechanistic perspective, five classes of inhibitors of SIRT1 have been identified: First, competitive inhibitors that vie for acylated substrates at the binding sites, exemplified by natural products such as sirtinol, splitomicin, and cambinol analogs;^{431,432} Second, competitive inhibitors that challenge NAD^+ for binding sites, including selisistat (EX-527) and Sosbo;^{433–435} Third, adenosine analogs such as Ro 31-8220;^{436,437} Further, binary inhibitors that compete with substrates or cofactors at separate binding sites, represented by ELT-31, a non-selective SIRT1-3 inhibitor;⁴³⁸ And the last, non-competitive inhibitors, including nicotinamide and its analogs, tenovins, thioacetyl-lysine peptides, and other small peptides.^{439–441} EX527, one of the few sirtuin inhibitors in clinical use, has demonstrated antitumor effects in vitro and potential as an adjunct in tumor therapy.⁴⁴² Additionally, it has proven safe and well-tolerated within the therapeutic concentration range for treating neurodegenerative diseases.⁴⁴³ However, minimal therapeutic effects were observed in a phase I clinical trial focusing on early-stage HD, with further large-scale trials needed to explore its clinical potential.⁴⁴⁴ Ongoing research also investigates EX-527's potential roles in improving other metabolic diseases, including endotoxemia,⁴⁴⁵ diabetic nephropathy,⁴⁴⁶ and infertility (NCT04184323) remains ongoing. Utilizing computational tools to predict potential allosteric sites has led to the identification of some allosteric SIRT6 inhibitors, including JYQ-42,⁴⁴⁷ compound 11e,⁴⁴⁸ and a pyrrole-pyridinimidazole derivative.^{449,450} Given that histone deacetylation catalyzed by SIRT6 promotes both tumor and non-tumor diseases, designing and in-depth study of these allosteric SIRT6 inhibitors represent a promising research field for human disease treatment.^{451,452}

The agonists of the sirtuin family have been widely studied since the discovery of the first SIRT1 agonist, resveratrol, in 2003.⁴⁵³ The initially discovered sirtuin agonists mainly upregulate target enzyme activity through allosteric effects and are classified into two primary categories based on their origins. The first category comprises natural products extracted from plants, including resveratrol and other polyphenolic molecules.^{454–459} The second category consists of synthesized agonists that exhibit greater selectivity, focusing particularly on SIRT1, such as SRT1460, SRT1720, SRT2104, SRT2183, and SRT3025, as well as those targeting SIRT6, including UBCS039 and MDL-800.⁴⁶⁰ Additionally, compounds that are suggested to upregulate SIRT1 expression have been identified. These are predominantly involved in the activation of the mitogen-activated protein kinase pathway,

including DDIT3,⁴⁶¹ phloretin,⁴⁶² puerarin,⁴⁶³ and atractylenolide III.⁴⁶⁴ Other reported agonists include astragaloside intravenous,⁴⁶⁵ hesperidin,⁴⁶⁶ caffeic acid phenethyl ester,⁴⁶⁷ agomelatine,⁴⁶⁸ ligustilide,⁴⁶⁹ tanshinone IIA,⁴⁷⁰ and farnesol.⁴⁷¹ These studies emphasize the role of SIRT1 activation in the deacetylation of various non-histones. However, the potential of these molecules as epigenetics-targeted drugs requires further exploration. Moreover, NAD^+ -enhancing molecules, which promote NAD^+ generation or rescue their levels, represent a novel class of sirtuin agonists. These molecules may activate all sirtuin members with a single compound, attracting considerable attention.⁴⁷² Given the diverse roles of NAD^+ in multiple signaling pathways, additional discussion is needed to determine whether drugs that increase NAD^+ levels have therapeutic effects in a sirtuin-dependent manner.

Targeting the reader of histone acetylation: BET, YEATS, and PHD. BET, YAF9, eleven-nineteen-leukemia protein (ENL), acute lymphocytic leukemia 1-fused gene from chromosome 9 protein (AF9), TAF14, and SAS5 (YEATS) domain, and PHD finger proteins are critical "readers" of acetylated residues and play essential roles as epigenetics-modifying enzymes in the transcription of downstream target genes. Drugs that target aberrant levels or activities of acetyl-recognition domain-containing proteins represent an emerging class of therapies for various diseases.

BET inhibitors: In recent years, a substantial number of BET inhibitors have been identified, encompassing pan-inhibitors, BD1/BD2 selective inhibitors, dual inhibitors of kinases and BET, and PROTACs-based inhibitors.^{60,473,474} From a therapeutic standpoint, BET inhibitors are primarily developed for treating tumors, with some also showing potential in non-tumor diseases, such as VYN-201 and VYN-202, among other BD2 selective inhibitors.⁴⁷⁵

More than twenty BET inhibitors have progressed to clinical trials, with several undergoing advanced phase evaluations. Notably, apabetalone (RVX-208) stands out as the sole BD2-selective inhibitor in phase III trials for addressing cardiovascular diseases and metabolic disorders such as T2DM.^{476–478} Apabetalone demonstrates significant anti-inflammatory properties, providing a robust scientific basis for its ongoing clinical evaluation.^{479–481} Another BET inhibitor, pelabresib (CPI-0610), has also reached phase III trials and shows promise as a treatment for myelofibrosis.⁴⁸² In earlier phase II studies, pelabresib combined with ruxolitinib surpassed the efficacy of Janus kinase inhibitor monotherapy in treating symptomatic myelofibrosis while maintaining a manageable safety profile.^{483,484} ZEN-3694, a leading pan-BET inhibitor, has advanced to phase II trials, demonstrating efficacy when used with cyclin-dependent kinases (CDKs) inhibitors and conventional chemotherapy in cancer treatment.⁴⁸⁵ Preliminary phase Ib/Ia trials indicate that ZEN-3694, in combination with enzalutamide, is beneficial for patients with metastatic castration-resistant prostate cancer.⁴⁸⁶ An increasing number of trials focusing on ZEN-3694 are currently underway, which will provide further data to evaluate its therapeutic promise. Furthermore, recent reports highlight dinaciclib, a well-known CDK inhibitor, now recognized for its novel activity in BET suppression.⁴⁸⁷ The dual inhibitory capability of dinaciclib presents a potential strategy to counteract BET resistance in AML treatment.⁴⁸⁷ These developments underscore the potential of these drugs to achieve market approval for broad clinical use. However, some BET inhibitors as single-agent therapies have shown mixed outcomes in clinical trials for distinct cancer settings, despite their excellent results in preclinical models.⁴⁸⁸ For example, several phases 1 and 2 clinical trials investigating the therapeutic effect of birabresib on solid or hematological malignancies were terminated prematurely because of limited efficacy (NCT02698176, NCT02698189, NCT02698176, NCT02296476). Therefore, combining BET inhibitors with other

traditional drugs may open new possibilities for the development of antitumor strategies.

Numerous novel BET inhibitors have been identified recently, enhancing the landscape of therapeutic options. These include OPN-51107, a pan-BET inhibitor that mitigates T cell dysfunction in chronic lymphocytic leukemia;⁴⁸⁹ XL-126, a BD1-selective inhibitor noted for its potent anti-inflammatory effects;⁴⁹⁰ and DW-71177, another BD1-selective inhibitor geared towards AML treatment.⁴⁹¹ Additional developments involve brain-permeable BD1-selective inhibitors for multiple sclerosis treatment,⁴⁹² compounds with dual HDAC/BET inhibitory action for challenging tumors,⁴⁹³ phenoxyaryl pyridone derivatives as BD2-selective inhibitors for AML,⁴⁹⁴ and SRX3177, a potent triple-action CDK4/6-phosphoinositide 3-kinase-BET inhibitor for respiratory diseases linked to β -coronavirus.⁴⁹⁵ These advancements significantly contribute to understanding BET-targeted drug development, designing small molecule inhibitors tailored to the diverse pathological characteristics of human diseases.

YEATS domain inhibitors: Identified in 2014, the YEATS domain—comprising YAF9, ENL, AF9, TAF14, and SAS5—serves as a novel reader for histone acetylation. This domain also recognizes histone crotonylation and benzoylation, which are critical in regulating gene expression.^{496–498} The human genome encodes four YEATS domain-containing proteins: ENL, YEATS domain-containing 2 (YEATS2), AF9, and glioma amplified sequence 41 (GAS41). These proteins are primarily implicated in the pathogenesis of tumors, particularly hematologic malignancies, and represent promising targets for epigenetic therapies.^{499–502} Research has shown that the YEATS domain binds to acylated lysine side chains through a common binding pocket and engages in π - π stacking interactions, providing a structural and theoretical foundation for developing targeted inhibitors.⁵⁰³ A significant milestone was the identification of the first small-molecule chemical probe, SGC-iMLLT, which targets ENL and its paralog AF9. This probe's inhibitory effects were validated in biological assays.⁵⁰⁴ Furthermore, another approach involves blocking the protein-protein interaction (PPI) between YEATS domain proteins and disruptor of telomeric silencing 1-like (DOT1L), effectively suppressing the activity of YEATS domain proteins.^{505,506} Current research is focused on developing selective inhibitors for various YEATS domain proteins, with the deepest insights into ENL inhibitors. In 2022, Liu et al.⁵⁰⁷ highlighted the promising potential of the oral ENL inhibitor TDI-11055 in treating AML in mouse models, advancing the clinical application of ENL inhibitors for AML treatment. Additionally, combination therapies involving ENL inhibitors with KAT or BET inhibitors have been emphasized.^{508,509} In 2020, Jiang et al.⁵¹⁰ introduced the first selective inhibitor targeting the AF9 YEATS domain, presenting a novel cyclopeptide for in-depth exploration of the functional similarities and differences between AF9 and ENL, thereby laying the groundwork for novel YEATS domain inhibitor development. An optimized method for the solid-phase synthesis of these inhibitory cyclopeptides has since been proposed, significantly reducing preparation time and enhancing yield.⁵¹⁰ Moreover, the study of amide- π interactions between histone acyl-lysine and the AF9 YEATS domain has led to the development of chemical compounds that disrupt this noncovalent interaction, notably those incorporating urea or aromatic rings.^{511,512} In 2021, the first selective GAS41 inhibitor was reported; this synthesized molecule binds to dimerized GAS41 YEATS domains and blocks interaction with acetylated histone H3 in cancer cell lines.⁵¹³

PHD finger domain inhibitors: BD and PHD finger-containing protein (BRPF) and BD and PHD finger transcription factor (BPTF) are crucial targets involved in tumor progression and the development of resistance to molecularly targeted therapy drugs, such as kinase inhibitors and poly ADP-ribose polymerase (PARP)

inhibitors.^{514–516} To date, an array of BRPF inhibitors featuring distinctive scaffolds—such as 3-acetyl-indole, 1,3-dimethylquinolin-2-one, 1,3-dimethyl benzimidazole, 1-(indolin-1-yl)ethan-1-one, 1,3-dimethylquinolin-2-one, and 2,3-dioxo-quinoxaline—has been identified. These compounds represent novel avenues for therapeutic innovation.^{517–522} However, as the inhibitory effects of these agents have primarily been confirmed in vitro, extensive efforts are required to advance these drugs to clinical trials. BPTF inhibitor development has not kept pace with those targeting other proteins with BD motifs, primarily remaining within fragment-based drug discovery. Only a handful have been tested in vivo or in vitro to demonstrate their inhibitory actions and therapeutic potential. AU1, the first small molecule selective for BPTF, has shown effectiveness in mouse models of gastric cancer and neuroblastoma.^{516,523,524} Bromosporine has exhibited significant antitumor effects in breast cancer and melanoma, suggesting promising therapeutic strategies for solid tumors.^{525,526} The novel selective inhibitor C620-0696 has shown cytotoxic effects in non-small-cell lung cancer cells overexpressing BPTF.⁵²⁷ The continued exploration of these inhibitors in oncology is highly anticipated.

Epigenetics-targeted drugs and histone methylation

Histone methylation is a highly dynamic regulator crucial for activating or suppressing gene transcription. Histone methyltransferases, demethylases, and reader proteins modify and maintain epigenetic signals that influence chromatin structure and cellular functions. Their dysregulation is linked to a variety of diseases, particularly malignant tumors. Recent advances in biochemistry and understanding of pathogenesis have led to identifying and developing small-molecule inhibitors that target aberrant demethylation patterns (Table 5).

Targeting the writer of histone methylation: KMT and PRMT. Histone methyltransferases (HMTs), including KMTs and protein arginine methyltransferases (PRMTs), are central to regulating histone methylation and are implicated in numerous biological and pathological processes. Inhibitors of HMTs are extensively researched as potential therapeutic agents. Notably, innovative drug discovery strategies for HMT proteins—such as covalent inhibition independent of SAM-competitive or substrate-competitive mechanisms, dual-target inhibition, and targeted degradation strategies—have received considerable attention and have rapidly progressed.^{528–530} These inhibitors, in addition to marketed drugs, are being advanced to clinical practice for further evaluation and oversight.

EZH2 inhibitors: Since the identification of the suppressor of variegation 3-9 homolog 1 (SUV39H1), the inaugural histone KMT8 discovered in 2000, numerous proteins mediating histone methylation have been reported. These include EZH1/2, euchromatic histone-lysine N-methyltransferase 2 (G9a/EHMT2), G9a-like protein (GLP/EHMT1), DOT1L, and various SET domain-containing histone lysine methyltransferase (SETD) and nuclear receptor binding SET domain protein (NSD) families.^{531–533} Over recent decades, considerable efforts have focused on developing efficient and selective inhibitors targeting various histone KMT subfamilies with potential therapeutic applications in disease treatment.^{534–537}

In addition to the two marketed drugs summarized in the previous section, tazemetostat (EPZ-6438) and valemestostat (DS-3201b), numerous novel EZH2 inhibitors are under investigation, with several advances in clinical studies, particularly compounds featuring the 2-pyridone moiety which encompass both bicyclic heteroaromatic and monocyclic aromatic rings.⁵³⁸ CPI-1205 (liracetostat) has undergone evaluation in three clinical trials (NCT03480646, NCT03525795, NCT02395601) to assess its tolerance and therapeutic potential. Although CPI-1205 has shown good tolerability in phase I stages, phase II trials have yet to

Table 5. Summary of histone methylation-targeted drugs for different diseases in clinical trials

| Type | Drug | Target(s) | Condition(s) | Status/outcome(s) | Phase(s) | Other intervention(s)/drug(s) | Study ID/reference(s) |
|---------------|-------------|-----------|---|--|------------|--|--------------------------------|
| KMT inhibitor | CPI-1205 | EZH2 | B cell lymphoma | Completed (unpublished) | Phase I | — | NCT02395601 |
| | CPI-1205 | EZH2 | Melanoma, NSCLC, RCC, urothelial carcinoma | Completed (unpublished) | Phase I | In combination with Ipilimumab | NCT03525795 |
| | CPI-1205 | EZH2 | Castration-resistant prostate cancer | Unknown | Phase I/II | In combination with Enzalutamide or Abiraterone/Prednisone | NCT03480646 |
| | CPI-0209 | EZH2/EZH1 | Ovarian cancer | Recruiting | Phase I | In combination with Carboplatin | NCT05942300 |
| | CPI-0209 | EZH2/EZH1 | MF/Sezary syndrome | Recruiting | Phase I | — | NCT05944562 |
| | CPI-0209 | EZH2/EZH1 | Urothelial carcinoma, ovarian cancer, endometrial carcinoma, DLBCL, PTCL, mesothelioma | Recruiting | Phase I/II | — | NCT04104776 |
| | SHR2554 | EZH2 | Healthy volunteers | Completed (metabolizing enzymes in vivo regulates the plasma concentration of SHR2554) | Phase I | In combination with Itraconazole | NCT04627129 ⁵⁴² |
| | SHR2554 | EZH2 | Healthy volunteers | Unknown | Phase I | — | NCT05049083 |
| | SHR2554 | EZH2 | Healthy volunteers | Completed (unpublished) | Phase I | — | NCT06010680 |
| | SHR2554 | EZH2 | Healthy volunteers | Completed (unpublished) | Phase I | In combination with Fluconazole | NCT05661591 |
| | SHR2554 | EZH2 | Healthy volunteers | Completed (unpublished) | Phase I | In combination with Omeprazole | NCT06093945 |
| | SHR2554 | EZH2 | Healthy volunteers | Completed (drug exposures are essentially the same in fasted and fed states) | Phase I | Following a high-fat diet or fasting status | NCT04335266 |
| | SHR2554 | EZH2 | Healthy volunteers | Completed (metabolizing enzymes in vivo regulates the plasma concentration of SHR2554) | Phase I | In combination with Rifampin | NCT04577885 |
| | SHR2554 | EZH2 | Mature lymphoid neoplasms | Unknown (exhibits satisfied efficacy and acceptable adverse effects according to available data) | Phase I | — | NCT03603951 ^{540,541} |
| | SHR2554 | EZH2 | FL | Not yet recruiting | Phase II | — | NCT06368167 |
| | SHR2554 | EZH2 | HR-positive, HER2-negative, endocrine-resistant advanced BC | Recruiting | Phase II | Umbrella study | NCT04355858 |
| | SHR2554 | EZH2 | PTCL | Recruiting | Phase I/II | Umbrella study | NCT05559008 |
| | SHR2554 | EZH2 | TNBC | Recruiting | Phase I/II | Umbrella study | NCT03805399 |
| | SHR2554 | EZH2 | B cell lymphoma, solid tumors | Recruiting | Phase I/II | SHR1701 (active comparator/ followed by SHR2554) | NCT04407741 |
| | SHR2554 | EZH2 | HL | Recruiting | Phase I/II | SHR1701 (active comparator/ followed by SHR2554) | NCT05896046 |
| | SHR2554 | EZH2 | PTCL | Recruiting | Phase I/II | In combination with CHOP | NCT06173999 |
| | SHR2554 | EZH2 | Castration-resistant prostate cancer | Completed (unpublished) | Phase I/II | With or without SHR3680 | NCT03741712 |
| | SHR2554 | EZH2 | PTCL | Recruiting | Phase III | Chidamide (active comparator) | NCT06122389 |
| | PF-06821497 | EZH2 | Castration-resistant prostate cancer, SCLC, FL | Recruiting | Phase I | — | NCT03460977 |
| | GSK126 | EZH2 | DLBCL, FL, MM, solid tumors | Terminated (the maximal dose and schedule shows insufficient evidence of clinical activity) | Phase I | — | NCT02082977 ⁵⁴⁶ |
| | XNW5004 | EZH2 | Squamous cell carcinoma of head and neck, urothelial carcinoma, prostate cancer, SCLC, NSCLC, cervical cancer | Recruiting | Phase I/II | In combination with Pembrolizumab | NCT06022757 |
| | AXT-1003 | EZH2 | NHL | Recruiting | Phase I | — | NCT05965505 |
| | EPZ-5676 | DOT1L | AML, ALL | Completed (unpublished) | Phase I | — | NCT02141828 |

Table 5. continued

| Type | Drug | Target(s) | Condition(s) | Status/outcome(s) | Phase(s) | Other intervention(s)/drug(s) | Study ID/reference(s) |
|----------------|-------------|-----------|--|--|------------|---|---------------------------------|
| PRMT inhibitor | EPZ-5676 | DOT1L | AML, ALL, MDS, myeloproliferative disorders | Completed (exhibits good safety profiles while unsatisfied efficacy) | Phase I | — | NCT01684150 ¹¹⁴⁶ |
| | EPZ-5676 | DOT1L | AML with an 11q23 translocation or partial tandem duplication | Completed (large-scale trials should be hold) | Phase I/II | In combination with Azacitidine | NCT03701295 |
| | EPZ-5676 | DOT1L | ALL | Terminated (due to the study agent is no longer available) | Phase I/II | In combination with Cytarabine and Daunorubicin | NCT03724084 |
| | EZM0414 | SETD2 | MM, DLBCL | Recruiting | Phase I | — | NCT05121103 |
| | KTX-1001 | NSD2 | MM | Recruiting | Phase I | — | NCT05651932 |
| | GSK3368715 | PRMT1 | DLBCL, PDAC, bladder cancer, NSCLC | Terminated (due to a lack in observed clinical efficacy and the unfavorable risk/benefit analysis) | Phase I | — | NCT03666988 ⁵⁶¹ |
| | CTS-2190 | PRMT1 | PDAC, NSCLC, TNBC | Recruiting | Phase I/II | — | NCT06224387 |
| | GSK3326595 | PRMT5 | TNBC, TCC, GBM, NHL, ACC, HR-positive BC, HPV-positive solid tumors, NSCLC | Completed (unpublished) | Phase I | With or without Pembrolizumab | NCT02783300 |
| | GSK3326595 | PRMT5 | BC | Completed (unpublished) | Phase II | Blank-controlled | NCT04676516 |
| | GSK3326595 | PRMT5 | MDS, AML | Terminated (due to an internal review of clinical data) | Phase I/II | With or without Azacitidine | NCT03614728 |
| | JNJ64619178 | PRMT5 | NHL, MDS, solid tumors | Active, not recruiting (clinical benefit is limited) | Phase I | — | NCT03573310 ⁵⁶⁸ |
| | JNJ64619178 | PRMT5 | Solid tumors | Completed (exhibits manageable dose-dependent toxicity with limited clinical benefit) | Phase I | — | ¹¹⁴⁷ |
| | PF06939999 | PRMT5 | NSCLC, urothelial carcinoma, squamous cell carcinoma of head and neck | Terminated (exhibits tolerable safety profiles and objective clinical responses in a subset of patients) | Phase I | With or without Docetaxel | NCT03854227 ^{569,1148} |
| | TNG908 | PRMT5 | NSCLC, mesothelioma, PDAC, sarcoma, GBM | Recruiting | Phase I/II | — | NCT05275478 |
| | MRTX1719 | PRMT5 | Mesothelioma, PDAC, NSCLC, malignant peripheral nerve sheath tumor | Recruiting | Phase I/II | — | NCT05245500 ⁵⁷¹ |
| | PRT543 | PRMT5 | DLBCL, myelodysplasia, myelofibrosis, ACC, MCL, AML, CMML | Completed (exhibits limited efficacy in ACC) | Phase I | — | NCT03886831 ⁵⁷² |
| | PRT811 | PRMT5 | Solid tumors, CNS lymphoma, gliomas | Completed (unpublished) | Phase I | — | NCT04089449 |
| | SKL27969 | PRMT5 | Solid tumors | Terminated (due to portfolio prioritization) | Phase I/II | — | NCT05388435 |
| | AMG193 | PRMT5 | Biliary tract cancer, PDAC | Recruiting | Phase I | In combination with Gemcitabine/Cisplatin/ Pembrolizumab, or Gemcitabine/Nab-paclitaxel, or modified FOLFIRINOX | NCT06360354 |
| | AMG193 | PRMT5 | NSCLC | Recruiting | Phase I | With or without Carboplatin/Paclitaxel/ Pembrolizumab, or Carboplatin/Pembrolizumab/ Pemetrexed, or Pembrolizumab, or Sotorasib | NCT06333951 |
| | AMG193 | PRMT5 | MTAP-null solid tumors | Recruiting | Phase I/II | With or without Docetaxel | NCT05094336 |
| | AMG193 | PRMT5 | MTAP-null solid tumors | Recruiting | Phase I/II | In combination with IDE397 | NCT05975073 |
| | SH3765 | PRMT5 | Advanced malignant tumors | Not yet recruiting | Phase I | — | NCT05015309 |
| | TNG462 | PRMT5 | MTAP-null solid tumors | Recruiting | Phase I/II | — | NCT05732831 |
| | SCR6920 | PRMT5 | Solid tumors, NHL | Recruiting | Phase I | — | NCT05528055 |
| | SYHX-2001 | PRMT5 | Solid tumors | Recruiting | Phase I | — | NCT05407909 |

Table 5. continued

| Type | Drug | Target(s) | Condition(s) | Status/outcome(s) | Phase(s) | Other intervention(s)/drug(s) | Study ID/reference(s) |
|---------------|-----------------|-----------|---|---|------------|---|---------------------------------------|
| KDM inhibitor | Tranylcypromine | LSD1 | Non-APL AML subtypes, MDS | Completed (unpublished) | Phase I | In combination with ATRA | NCT02273102 |
| | Tranylcypromine | LSD1 | Non-APL AML subtypes | Unknown | Phase I/II | In combination with ATRA | NCT02261779 |
| | Tranylcypromine | LSD1 | Non-APL AML subtypes | Unknown | Phase I/II | In combination with ATRA and Cytarabine | NCT02717884 |
| | ORY-1001 | LSD1 | AML, MDS | Recruiting | Phase I | In combination with Azacitidine and Venetoclax | NCT06357182 |
| | ORY-1001 | LSD1 | AML | Recruiting | Phase I | In combination with Gilteritinib | NCT05546580 |
| | ORY-1001 | LSD1 | SCLC | Not yet recruiting | Phase I/II | Atezolizumab and Durvalumab (active comparator/ followed by ORY-1001) | NCT06287775 |
| | ORY-1001 | LSD1 | AML | Completed (exhibits a good safety profile without significant extra-hematologic toxicity) | Phase I | — | EudraCT 2013-002447-29 |
| | ORY-1001 | LSD1 | AML | Completed (unpublished) | Phase II | In combination with Azacitidine | EudraCT 2018-000482-36 ⁶²⁴ |
| | ORY-2001 | LSD1 | Healthy volunteers | Completed (exhibits good safety and tolerability) | Phase I | Placebo-controlled | EUDRACT 2015-003721-33 ⁶²⁷ |
| | ORY-2001 | LSD1 | MS | Ongoing (exhibits safety and tolerability according to early clinical data) | Phase II | Placebo-controlled | EudraCT 2017-002838-23 |
| | ORY-2001 | LSD1 | AD | Completed (exhibits good efficacy and tolerability) | Phase II | Placebo-controlled | EudraCT 2017-004893-32 |
| | ORY-2001 | LSD1 | ADHD, BPD, ASD | Completed (exhibits good efficacy and tolerability) | Phase II | — | EudraCT 2018-002140-88 |
| | ORY-2001 | LSD1 | AD | Completed (exhibits good efficacy and tolerability) | Phase II | Placebo-controlled | EudraCT 2019-001436-54 |
| | ORY-2001 | LSD1 | ARDS | Completed (exhibits good efficacy and tolerability) | Phase II | In combination with standard care treatment | EudraCT 2020-001618-39 |
| | ORY-2001 | LSD1 | AD | Completed (unpublished) | Phase II | Placebo-controlled | NCT03867253 |
| | ORY-2001 | LSD1 | BPD | Completed (unpublished) | Phase II | Placebo-controlled | NCT04932291 |
| | GSK-2879552 | LSD1 | SCLC | Terminated (due to the unfavorable risk/benefit analysis) | Phase I | — | NCT02034123 |
| | GSK-2879552 | LSD1 | AML | Terminated (due to the unfavorable risk/benefit analysis) | Phase I | In combination with ATRA | NCT02177812 |
| | GSK-2879552 | LSD1 | MDS | Terminated (due to the unfavorable risk/benefit analysis) | Phase I/II | With or without Azacitidine | NCT02929498 |
| | IMG-7289 | LSD1 | AML | Recruiting | Phase I | In combination with Venetoclax | NCT05597306 |
| | IMG-7289 | LSD1 | AML, MDS | Completed (exhibits a good safe profile) | Phase I/II | With or without ATRA | NCT02842827 |
| | IMG-7289 | LSD1 | SCLC | Active, not recruiting | Phase I/II | In combination with Atezolizumab | NCT05191797 |
| | INCB059872 | LSD1 | Ewing sarcoma | Terminated (due to business decision) | Phase I | — | NCT03514407 |
| | INCB059872 | LSD1 | AML, MDS, SCLC, myelofibrosis, Ewing sarcoma, poorly differentiated neuroendocrine tumors | Terminated (due to business decision) | Phase I/II | With or without ARTA, Azacitidine, and Nivolumab | NCT02712905 |
| | INCB059872 | LSD1 | NSCLC, colorectal cancer | Terminated (due to sponsors' decision) | Phase I/II | In combination with Pembrolizumab and Epacadostat | NCT02959437 |
| | SP-2577 | LSD1 | Solid tumors | Completed (unpublished) | Phase I | — | NCT03895684 |
| | SP-2577 | LSD1 | Ewing sarcoma, myxoid liposarcoma, desmoplastic small round cell tumor | Active, not recruiting | Phase I | With or without Cyclophosphamide and Topotecan | NCT03600649 |
| | SP-2577 | LSD1 | Ovarian cancer, endometrial cancer | Withdrawn (due to salaries discontinued support) | Phase I | In combination with Pembrolizumab | NCT04611139 |
| | SP-2577 | LSD1 | CMML, MDS | Recruiting | Phase I/II | In combination with Azacytidine | NCT04734990 |

Table 5. continued

| Type | Drug | Target(s) | Condition(s) | Status/outcome(s) | Phase(s) | Other intervention(s)/drug(s) | Study ID/reference(s) |
|----------------------|----------|-------------|--|--|------------|---|----------------------------------|
| | SP-2577 | LSD1 | Ewing sarcoma, myxoid liposarcoma, desmoplastic small round cell tumor | Enrolling by invitation | Phase I/II | — | NCT05266196 |
| | CC-90011 | LSD1 | AML | Terminated (due to business decision) | Phase I | Azacitidine and Venetoclax (active comparator/ followed by CC-90011) | NCT04748848 |
| | CC-90011 | LSD1 | Castration-resistant prostate cancer | Completed (unpublished) | Phase I | In combination with Abiraterone and Prednisone | NCT04628988 |
| | CC-90011 | LSD1 | NHL, solid tumors | Terminated (due to business decision) | Phase I | In combination with Rifampicin | NCT02875223 ^{1149,1150} |
| | CC-90011 | LSD1 | SCLC | Active, not recruiting | Phase I | In combination with Cisplatin and Etoposide | NCT03850067 |
| | CC-90011 | LSD1 | SCLC, NSCLC | Completed (unpublished) | Phase II | In combination with Nivolumab | NCT04350463 |
| | 4SC-202 | LSD1 | AML, ALL, MDS, CLL, MM | Completed (exhibits a good safety profile and antitumor activities) | Phase I | — | NCT01344707 ¹¹⁰² |
| | 4SC-202 | LSD1 | Oesophagogastric adenocarcinoma, colorectal cancer | Unknown (oesophagogastric adenocarcinoma cohort meets the criteria to expand to stage 2 according to disclosed data) | Phase II | In combination with Avelumab | NCT03812796 ¹¹⁰³ |
| | 4SC-202 | LSD1 | Melanoma | Completed (unpublished) | Phase I/II | In combination with Pembrolizumab | NCT03278665 |
| | 4SC-202 | LSD1 | Melanoma | Active, not recruiting (4SC-202 addition does not increase treatment efficacy according to early clinical data) | Phase I/II | Nivolumab (active comparator/in combination with 4SC-202); in combination with Nivolumab/ipilimumab | NCT04133948 ⁴²⁴ |
| | JB1-802 | LSD1, HDAC6 | SCLC and other neuroendocrine-derived cancers | Recruiting | Phase I/II | — | NCT05268666 |
| | TAK-418 | LSD1 | Healthy volunteers | Completed (exhibits good tolerability, pharmacokinetic and pharmacodynamic effects) | Phase I | Placebo-controlled | NCT03228433 ⁶³⁸ |
| | TAK-418 | LSD1 | Healthy volunteers | Terminated (due to business decision) | Phase I | Placebo-controlled | NCT03501069 ⁶³⁸ |
| | TAK-418 | LSD1 | Healthy volunteers | Terminated (due to administrative reasons) | Phase I | In combination with [18 F] MNI-1054 (radiotracer) | NCT04202497 |
| | LH-1802 | LSD1 | AML, MDS | Ongoing | Phase I | — | CTR20222026 |
| | SYHA1807 | LSD1 | SCLC | Unknown | Phase I | — | NCT04404543 |
| WDR domain inhibitor | MAK683 | EED | DLBCL | Active, not recruiting | Phase I | — | NCT02900651 |

ACC adenoid cystic carcinoma, AD Alzheimer's disease, ADHD attention deficit hyperactivity disorder, ALL acute lymphoblastic leukemia, AML acute myeloid leukemia, APL acute promyelocytic leukemia, ARDS acute respiratory distress syndrome, ASD autism spectrum disorder, ATRA all-trans-retinoic acid, BC breast cancer, BPD borderline personality disorder, CHOP Cyclophosphamide, Hydroxydoxorubicin, Oncovin, and Prednisone, CLL chronic lymphocytic leukemia, CMML chronic myelomonocytic leukemia, CNS central nervous system, DLBCL diffuse large B cell lymphoma, DOT1L disruptor of telomeric silencing 1-like, EED embryonic ectoderm development, EZH2 enhancer of zeste homolog 2, FL follicular lymphoma, GBM glioblastoma multiforme, HDAC histone deacetylase, HER2 human epidermal growth factor receptor 2, HL Hodgkin lymphoma, HPV human papillomavirus, HR hormone receptor, KDM lysine demethylase, KMT lysine methyltransferase, LSD1 lysine specific demethylase 1, MCL mantle cell lymphoma, MDS myelodysplastic syndrome, MF mycosis fungoides, MM multiple myeloma, MS multiple sclerosis, MTAP methyl-thioadenosine phosphorylase, NHL non-Hodgkin lymphoma, NSD nuclear receptor binding SET domain protein, NSCLC non-small cell lung cancer, PDAC pancreatic ductal adenocarcinoma, PRMT protein arginine methyltransferase, PTCL peripheral T cell lymphoma, RCC renal cell carcinoma, SCLC small cell lung cancer, SETD2 SET domain-containing histone lysine methyltransferase 2, TCC transitional cell carcinoma of the urinary system, TNBC triple-negative breast cancer, WDR WD40 repeat

provide sufficient data to confirm its antitumor efficacy. CPI-0209 (turmimetostat), an oral, next-generation dual EZH2/EZH1 inhibitor developed by the same company, is currently under clinical trial for treating both solid tumors and hematological malignancies (NCT05944562, NCT05942300, NCT04104776). SHR2554, a highly selective EZH2 inhibitor, has demonstrated potent efficacy both in vitro and in vivo.⁵³⁹ Its first-in-human, dose-escalation, and dose-expansion phase 1 trial conducted at 13 hospitals in China in 2018

indicated good tolerance and promising antitumor activity in patients with R/R lymphomas.^{540,541} A pharmacokinetic study revealed that combining itraconazole, an inhibitor of CYP3A4-metabolizing enzymes, with SHR2554 improves its plasma concentration while maintaining a favorable safety profile, suggesting a new therapeutic strategy.⁵⁴² PF-06821497, a lactam-derived EZH2 inhibitor, was optimized from a series of similar compounds using ligand-based and physicochemical-

property-based strategies, showing optimal inhibitory and therapeutic effects in mouse models.⁵⁴³ It also has demonstrated synergistic effects in combination with HDAC inhibitors, inhibiting proliferation and inducing apoptosis in cancer cell lines.⁵⁴⁴ Currently, two clinical studies are exploring appropriate administration methods for PF-06821497, such as intravenous injection or oral intake, and whether it can be consumed with food (NCT06392230, NCT05767905). In addition to demonstrating a strong therapeutic effect in tumor treatment in animal models, GSK126 has also achieved significant breakthroughs in enhancing β -like cell regeneration among patients with T1DM.^{111,545} However, a terminated phase I clinical trial revealed insufficient evidence of clinical activity for GSK126 at tolerable doses.⁵⁴⁶ XNW5004 (NCT06022757) and AXT-1003 (NCT05965505) are innovative EZH2 inhibitors currently in clinical trials, reflecting ongoing advancements in this therapeutic area. Moreover, astemizole, originally an antiallergy medication inhibiting histamine receptor H1, has recently been shown to disrupt the EZH2-embryonic ectoderm development (EED) PPI within the PRC2, offering new perspectives in developing EZH2/PRC2 inhibitors.⁵⁴⁷

DOT1L inhibitors: EPZ-5676 (pinometostat), EPZ004777, and SGC0946 are three selective inhibitors of DOT1L that are currently under extensive research. EPZ-004777 was the first SAM-competitive inhibitor of DOT1L to demonstrate *in vivo* efficacy.⁵⁴⁸ Despite showing promising therapeutic effects in various subtypes of AML through cell experiments and animal models, EPZ-004777's preclinical application has been largely constrained by its pharmacokinetic characteristics.^{549,550} EPZ-5676 has been developed to improve selectivity and inhibition effects, showing potential as a therapeutic agent for mixed lineage leukemia (MLL).⁵⁵¹ Early investigations using patient-derived xenografts and mouse models have indicated that EPZ-5676 exhibits potent antileukemic activities, facilitating further evaluation.^{552,553} In three completed clinical trials (NCT01684150, NCT02141828, NCT03701295), EPZ-5676 has been assessed for safety, tolerability, and preliminary antitumor activity in pediatric patients with MLL, with the combination of EPZ-5676 and azacytidine in a phase Ib/II study expected to show synergistic antiproliferative activities (NCT03701295). SGC0946, a brominated analog, serves as another selective inhibitor of DOT1L. Its therapeutic potential, either as monotherapy or in combination with other inhibitors such as HDACs and the mitogen-activated protein kinase pathway, has been observed in various solid tumors, setting the groundwork for clinical trials of SGC0946.^{554–556}

Beyond EZH2 and DOT1L, several inhibitors targeting other subfamilies are being investigated in clinical studies, including EZM0414 and KTX-1001. EZM0414, a novel inhibitor of SETD2 derived from the optimization of EPZ-719, exhibits improved pharmacokinetic properties and potent pharmacodynamic activity in mouse xenograft models.⁵⁵⁷ A phase I/Ib clinical trial is currently underway to explore the safety, tolerability, and therapeutic efficacy of EZM0414 in patients with R/R MM and R/R diffuse large B-cell lymphoma (NCT05121103). KTX-1001, a selective NSD2 inhibitor, has been FDA-approved for clinical trials since 2022 and is being studied in a phase I trial to treat patients with R/R MM (NCT05651932). These meticulously organized clinical trials focusing on KMTs are drawing increasing attention, leading to significant breakthroughs in understanding the relationship between human diseases and aberrant histone methylation.

PRMT inhibitors: Significant progress has been made in developing inhibitors for type I PRMTs (PRMT1-4, 6, and 8) and a selective inhibitor targeting PRMT5, with several agents entering the early phases of clinical trials.

Two type I PRMTs inhibitors are already in clinical stages, including GSK3368715 and CTS-2190. GSK3368715 (EPZ019997), an oral, reversible inhibitor of PRMT1/6/8 developed for treating

tumors and pulmonary disorders.^{558–560} GSK3368715 underwent a phase 1 clinical trial for treating solid tumors and diffuse large B-cell lymphoma in 2018. However, the first clinical application of a PRMT1 inhibitor did not meet expectations and was terminated early in 2022 due to its ineffectiveness.⁵⁶¹ Given the adverse events potentially caused by high and sustained concentrations of the inhibitor *in vivo*, research into PROTAC-based degraders of GSK3368715 has intensified, potentially offering therapeutic benefits at lower doses and reducing adverse effects.⁵⁶² CTS-2190, another inhibitor targeting PRMT1/3/4/6, received clinical trial approvals from the US FDA and China NMPA in February and April 2023, respectively. A phase I/II clinical trial is being conducted to evaluate its tolerability and preliminary antitumor activity in healthy participants and patients with solid tumors (NCT06224387).

Thirteen PRMT5 inhibitors have advanced to phase I and II clinical trials. Among these, GSK3326595, JNJ64619178, and PF06939999 were the earliest selective PRMT5 inhibitors to receive clinical trial approvals. GSK3326595 is a substrate-competitive inhibitor, while JNJ64619178 and PF-06939999 function as SAM-competitive agents.^{563–565} The efficacy and understanding of GSK3326595 primarily rely on animal model data due to a lack of published results from completed clinical trials. This inhibitor has been shown to induce DNA damage in cancer cells and enhance the antiproliferative effects of poly ADP-ribose polymerase inhibitors, such as niraparib;⁵⁶⁶ however, long-term or chronic use of GSK3326595 is associated with potential liver-related adverse effects.⁵⁶⁷ A completed phase I clinical trial of JNJ-64619178 determined that a daily dose of 0.5 mg was better tolerated by participants with R/R B cell non-Hodgkin lymphoma, though it demonstrated limited therapeutic effects.⁵⁶⁸ Conversely, PF-06939999 has shown an acceptable safety profile and clinical efficacy in its phase I trial.⁵⁶⁹ TNG908 and MRTX1719, both brain-penetrant PRMT5 inhibitors, have shown promise in selectively targeting cancer cells deficient in methylthioadenosine phosphorylase in both preclinical models and clinical trials.^{570,571} Phase I/II clinical trials for these drugs recruit participants to assess their therapeutic effects on various solid tumors (NCT05245500). Prelude Therapeutics has developed PRT543 and PRT811, leading oral PRMT5 inhibitors whose safety profiles and preliminary therapeutic potential have been evaluated in phase I clinical trials (NCT04089449, NCT03886831). PRT543 has demonstrated good tolerance and efficacy among patients with adenoid cystic carcinoma, warranting further advanced clinical testing.⁵⁷² A phase I/II clinical trial of SKL27969 began in 2022 to evaluate its safety, tolerability, pharmacokinetics, pharmacodynamics, and preliminary efficacy in patients with advanced solid tumors. However, this study was terminated in 2024 due to portfolio prioritization, with no significant safety trends or issues identified during its execution (NCT05388435). Other PRMT5 inhibitors, such as AMG193, SH3765, TNG462, SCR6920, and SYHX-2001, are currently under investigation in clinical trials and are in the “recruiting” phase. Given that most of the small molecules or core scaffolds of PRMT5 inhibitors have been examined only in cellular experiments, there remains a significant gap in knowledge regarding their efficacy and therapeutic effects *in vivo*. Therefore, it is imperative to bridge the crucial divide between fundamental research and clinical application.

Targeting the eraser of histone methylation: KDM. KDMs are enzymes that remove histone and nonhistone methylation. They can be divided into two categories based on their molecular structures: flavin adenine dinucleotide-dependent KDM (KDM1) and Fe(II)- and α -KG-dependent KDM (KDM2-7), also called Jumonji C (JmjC)-KDMs.⁵⁷³ Both upregulation and downregulation of KDMs can affect the expression of pathological genes in cancers or other disorders. Currently, representative inhibitors of

diverse KDM proteins are being investigated. Based on catalytic mechanisms, lysine specific demethylase 1 (LSD1/KDM1A) inhibitors can be divided into irreversible and reversible inhibitors; KDM2-7 inhibitors are classified into four types: α -KG cofactor mimics or inhibitors of α -KG oxygenases (such as N-oxalylglycine), metal cofactor disruptors, histone substrate competitive inhibitors, and other substrate- and cofactor-independent inhibitors.⁵⁷⁴

KDM2/7 inhibitors: KDM2 and KDM7 proteins, which belong to the JmjC-KDM subfamilies, share high similarity in their Fe(II)- and α -KG-binding residues.⁵⁷⁵ The development of inhibitors for KDM7 and KDM2 has typically co-occurred. In 2013, a series of hydroxamate analogs featuring an alkyl chain were identified. These compounds demonstrated antiproliferative activity in cancer cells by inhibiting KDM2A, KDM7A, and KDM7B.⁵⁷⁶ Similarly, Gerken et al.⁵⁷⁷ developed a series of novel KDM2A/7A inhibitors characterized by saturated indoline ring systems. These indoline-containing compounds exhibited potent and selective effects on KDM2A/7A at low micromolar concentrations, with notable cellular activity. Nonetheless, addressing limitations such as cytotoxicity and off-target effects remains challenging for future research. Other selective inhibitors have also been identified, including a cyclic peptide inhibitor, OC9, designed to target the PHD finger domain of KDM7. This inhibitor results in the inhibition of KDM7B and the activation of KDM7A.⁵⁷⁸ Through virtual screening of α -KG oxygenases, daminozide, a plant growth regulator, was found to selectively inhibit KDM2A. The therapeutic potential of daminozide was observed in mouse models of osteoarthritis, pointing to new directions for developing 2KG-competed inhibitors with enhanced selectivity, although its use in humans is unlikely.^{579,580}

KDM3 inhibitors: In addition to IOX1, various inhibitors of the KDM3 family have been identified, most exhibiting a pan-inhibitory effect across all family members. Through virtual screening of natural products and traditional Chinese medicine components, compounds JDI-4, JDI-12, and JDI-16 selectively bind to the JmjC domains of KDM3B and KDM3C.⁵⁸¹ Subsequent in vitro and in vivo studies confirmed the inhibitory effect and antitumor potential of JDI-16 in a KDM3-dependent manner.⁵⁸¹ Another compound, JDM-7, also identified from this screening, inhibits KDM3A and KDM3B in AML cell lines, although initial observations indicated limited effects on the KDM3 family.⁵⁸² Additionally, through high-throughput screening of benzhydryl amine derivatives, CBA-1 was found to be a potent inhibitor of KDM3A, exhibiting antiproliferative effects on colorectal cancer cell lines.⁵⁸³ The use of CBA-1 in zebrafish models also showed minimal toxicity, suggesting its potential as a promising drug for clinical application.⁵⁸³

KDM4 inhibitors: Given the critical roles of KDM4s in cancers and the inherent complexity of the KDM4 subfamily, significant efforts have been dedicated to developing KDM4 inhibitors. These inhibitors are categorized into four previously reviewed classes: α -KG cofactor mimics, Metal cofactor disruptors, histone substrate competitive inhibitors, and inhibitors targeted reader domains, having been summarized extensively in previous work.^{584,585} In addition to these established categories, we emphasize the progress in novel inhibitors that have not yet been summarized, further expanding the scope of therapeutic options against KDM4-related cancers.

TACH101 is a novel pan-inhibitor of KDM4A-D, competitively inhibiting α -KG without affecting other KDM subfamilies. The therapeutic effects of TACH101 have been demonstrated in organoids and xenograft models, suggesting its potential as an anticancer agent worthy of further investigation in animal studies.⁵⁸⁶ SD49-7, a derivative of SD70, is another novel KDM4

inhibitor. It has shown a stronger effect than SD70 in suppressing the proliferation of AML cell lines and enhancing the progression of resistant tumors in mouse models.⁵⁸⁷ Based on virtual screening, 2-(methylcarbamoyl)isonicotinic acid has been identified as an initial active fragment specifically inhibiting KDM4A by preventing its binding to H3K9me3 in a substrate-competitive manner.⁵⁸⁸ Molecular docking and dynamics approaches have recently revealed that a series of natural products containing sugars, aromatic rings, and OH or O⁻ groups can interact with KDM4 and inhibit its activities.⁵⁸⁹ However, the mechanisms of these interactions remain unclear, underscoring the need for further development of these potential drugs.

KDM5 inhibitors: KDM5 inhibitors have shown significant therapeutic potential, though many compounds still lack sufficient evidence to confirm their efficacy and safety in vivo.

A prevailing approach in KDM5 inhibition involves designing small molecules that compete with α -KG for binding sites.⁵⁹⁰ Among these, KDOAM-25 is a potent and selective inhibitor affecting MM and triple-negative breast cancer cells, with minimal adverse effects observed in vivo applications. Nevertheless, its poor cell membrane permeability hinders its efficacy.⁵⁹¹ RS3195 exhibits inhibitory effects on KDM5B and KDM5D in vitro. Due to potential toxicity, RS5033 was developed as an alternative, featuring a phenyl ring instead of a pyrrole nucleus to improve tolerance.⁵⁹² KDM5-C49, an analog of 2,4-PDCA, binds to KDM5B in vitro and inhibits its enzymatic activities.⁵⁹³ To enhance cell membrane permeability and selectivity, derivatives KDM5-C48 and KDM5-C70 have been developed.^{593–595} Through high-throughput virtual screening, a series of cyclopenta[c]chromen derivatives targeting KDM5A have been identified as promising drugs due to their potent inhibitory effects and low toxicity.⁵⁹⁶ N70, a thienopyridine-based selective KDM5A inhibitor, displays α -KG-competitive inhibition, while its analog, N71, binds irreversibly to KDM5A through covalent modifications.⁵⁹⁷

Numerous compounds, identified through virtual screening and optimization of reported inhibitory molecules, employ different mechanisms of action.^{598,599} Among these, KDM5-inh1 and CPI-455 are broadly studied pan-inhibitors of KDM5. Using either KDM5-inh1 or CPI-455 has demonstrated therapeutic effects on cancer cell lines and has facilitated synergistic interactions with conventional antitumor agents.^{600,601} Further research should explore the potential for this synergy in animal models. GS-5801, designed from GS-080—one of the most potent KDM5 inhibitors—shows significant anti-HBV activity. Despite its promise, the in vivo effects of GS-5801 have not met expectations, underscoring the need for additional studies to enhance its efficacy.⁶⁰² Utilizing the AlphaScreen method, ryuvidine was identified as an inhibitor of KDM5A/B/C, exhibiting substantial therapeutic impact on drug-tolerant cells.⁶⁰³ Dexmedetomidine, recently identified as a KDM5 inhibitor, is utilized to manage acute kidney injury in a KDM5-dependent manner.⁶⁰⁴ A novel approach was introduced by Yang et al.,⁶⁰⁵ who reported the first selective metal-based KDM5A inhibitor, rhodium(III) complex1. This compound disrupts the interaction between KDM5A and H3K4me2/3, offering a new scaffold for optimizing KDM5A-targeted drugs. The screening of imidazopyridine-analogs of zolpidem led to the discovery of O4I3, a novel chemical inhibitor of KDM5A. O4I3 generates and sustains patient-specific induced pluripotent stem cells in vitro.⁶⁰⁶ Additionally, TK-129, a pyrazole-based KDM5B inhibitor, is applied in treating cardiovascular diseases.⁶⁰⁷ High-throughput screening technology has facilitated the identification of PBIT, another novel KDM5B inhibitor. Despite its promising attributes, PBIT exhibits unstable therapeutic effects across different cell lines, necessitating careful consideration of its application in treatment.⁶⁰⁸ Similarly, several pyrazole derivatives that inhibit KDM5B have been recognized, with several demonstrating potent activity in cells, suggesting new therapeutic strategies.⁶⁰⁹ Furthermore, lida

et al.⁶¹⁰ designed a selective KDM5C inhibitor with a triazole scaffold and subsequently synthesized a KDM5C degrader using PROTAC techniques. This selective degrader shows enhanced inhibitory effects on prostate cancer cell lines compared to its prodrug, thus expanding the possibilities for anticancer agent design.

KDM6 inhibitors: The KDM6 subfamily has gained attention as a therapeutic target for various diseases. GSK-J1 and GSK-J4 are two well-studied classical inhibitors of KDM6B, showing significant potential in treating autoimmune diseases, metabolic disorders, and tumors, and enhancing the effectiveness of traditional antitumor agents.^{611–614} Using optimized delivery systems for GSK-J1 has further advanced the development of effective in vivo strategies.⁶¹⁵ Beyond these compounds, novel inhibitors have been introduced. For instance, KDOBA67, a hydroxyl derivative of GSK-J4, demonstrates favorable cell permeability in chordoma cell lines and inhibits the progression of chordoma.⁶¹⁶ Employing a virtual fragment screening approach, Giordano et al.⁶¹⁷ identified a series of benzoxazole scaffold compounds that bind to the KDM6B subfamily with high affinity, showing therapeutic promise in melanoma cell lines. Zhang et al.⁶¹⁸ developed a simple capillary electrophoresis method for screening KDM6B inhibitors, leading to the identification of salvianic acid A and puerarin 6"-O-xyloside as effective agents. Additionally, Jones et al.⁶¹⁹ used computational methods to develop an optimized peptide derived from the H3 C-terminus, which may enhance selectivity when linked with known inhibitors.

LSD1 inhibitors: Extensive research has been conducted on the biological and pathological functions of LSD1 and its inhibitors. Compared to other KDM subfamilies, LSD1 inhibitors have seen significant advances.⁶²⁰ Currently, several LSD1 inhibitors such as tranylcypromine (TCP), ORY-1001 (ladademstat), ORY-2001, GSK-2879552, IMG-7289 (bomedemstat), INCB059872, SP-2577 (seclidemstat), CC-90011 (pulrodemstat), 4SC-202 (domatinostat), JBI-802, TAK-418, LH-1802, and SYHA1807, are undergoing clinical trials.

TCP, an irreversible inhibitor, is being used in clinical practice among patients with AML and MDS, showing promising effects either alone or in combination with all-trans-retinoic acid in phase I/II clinical trials, with overall response rates exceeding 20%.^{621,622} Building on TCP's structure, novel inhibitors like ORY-1001, ORY-2001, GSK-2879552, INCB059872, and IMG-7289 have been developed, which also bind irreversibly to LSD1.⁶²³ These advancements have broadened the spectrum of treatable diseases with LSD1 inhibitors. Notably, ORY-1001 and ORY-2001, both orally administered, have been evaluated for their effectiveness in R/R hematologic malignancies and neurological disorders such as borderline personality disorder and AD.^{624–628} In completed phase I clinical trials, ORY-1001 exhibited a good safety profile without significant extrahematologic toxicity among healthy volunteers and patients with AML, indicating good therapeutic potential.^{624,627} GSK2879552 has shown antitumor efficacy in animal models,⁶²⁹ yet several clinical trials have been terminated due to a high incidence of adverse events.^{630,631} Similarly, clinical trials for INCB059872 were halted due to business decisions, among other reasons (NCT02959437, NCT03514407, NCT03132324, NCT02712905). Greater attention must be dedicated to evaluating the tolerability and efficacy of novel treatments. According to completed clinical trials, IMG-7289 demonstrates potential in ameliorating several blood disorders (NCT04254978, NCT03136185, NCT02842827), with numerous recent registrations for novel clinical trials concerning this drug. SP-2577 and CC-9001 are reversible LSD1 inhibitors,⁶³² which, compared to their irreversible counterparts, exhibit enhanced safety profiles and have been extensively studied in both solid tumors and hematological malignancies.^{633,634} Domatinostat and JBI-802, dual

inhibitors targeting LSD1 and HDAC, selectively interact with class I HDAC isoenzymes and HDAC6, respectively.⁶²⁰ Although promising antitumor effects have been observed in cancer cell lines, the therapeutic potential of domatinostat requires further exploration due to its unfavorable toxicity.^{424,635} TAK-418, a novel LSD1 inhibitor noted for its effective brain penetration, is considered a potential treatment for central nervous system disorders.^{636,637} The administration of TAK-418 was well tolerated by healthy volunteers in a phase I clinical trial, laying a solid foundation for further investigation.⁶³⁸ LH-1802 and SYHA1807, novel inhibitors, are currently under clinical trial evaluation for metastatic prostate cancer and small-cell lung cancer, respectively (NCT03678025, NCT04404543). The encouraging outcomes from these clinical-stage applications have spurred greater interest in the development of LSD1 inhibitors, with ongoing efforts to discover effective and tolerable agents.

Targeting the reader of histone methylation: reader domains. The identification of histone lysine and arginine methylation is attributed to proteins possessing malignant brain tumor (MBT) domains, chromodomains, Tudor domains, proline-tryptophan-tryptophan-proline (PWWP) domains, PHD fingers, and WD40 repeat (WDR) domains.⁶³⁹ Notably, enzymes that serve as writers or erasers for histone methylation may also contain these reader modules, such as PHD fingers and Tudor domains, aiding in recognizing residues they catalyze.⁶⁴⁰ Although numerous inhibitors targeting reader domains have been discovered, nearly half of these originate from structure-based virtual screenings and lack in vivo evaluation of their inhibitory effects and therapeutic activity.⁶⁴¹ Encouragingly, MAK683, an inhibitor targeting EED—a representative histone methylation reader containing the WDR domain—has entered clinical trials.⁶⁴² Currently, MAK683 is in a phase I clinical trial for treating diffuse large B-cell lymphoma (NCT02900651). Inspired by this milestone, many potent and selective inhibitors of EED and other molecules targeting reader domain proteins are expected to advance into clinical trials as promising therapeutic strategies.

Epigenetics-targeted drugs and m6A

RNA m6A methylation, a prevalent and conserved modification in eukaryotic RNAs, is crucial in determining transcript fate at the post-transcriptional level through RNA processing, export, degradation, and translation. Dysregulated m6A regulators contribute to various pathological conditions, particularly in the pathogenesis of diverse tumors.⁶⁴³ With the identification of various enzymes involved in m6A modification—including writers, erasers, and readers—the reversibility of m6A modification has been increasingly recognized, providing a foundation for developing epigenetics-targeted drugs that regulate RNA m6A as a core mechanism.

Targeting the writer of m6A: METTL3. METTL3 plays a critical role in the m6A modification process by transferring methyl groups from SAM to target RNA, catalyzing the conversion of adenosine to methyladenosine. This function of METTL3, the most extensively studied m6A writer, has been linked to the development of various pathologies, notably various tumors.⁶⁴⁴ Recent research has highlighted a range of inhibitors and agonists targeting METTL3, with several epigenetic drugs demonstrating promising efficacy both in vitro and in vivo, thus reinforcing the significance of METTL3 regulation in disease pathology and its potential as a therapeutic target.

METTL3 inhibitors: The study of METTL3 inhibitors has attracted increasing attention due to their diverse roles in regulating gene expression across different diseases. These inhibitors are categorized into competitive and allosteric inhibitors and gene expression suppressors, each leveraging distinct mechanisms of action.

SAM analog is the dominant part of the competitive inhibitors, initially developed based on a fulfilled screening of compounds containing the adenosine moiety (the fragment responsible for the combination with METTL3 at the binding sites for SAM). In 2020, Bedi et al.⁶⁴⁵ performed a series of docking studies on over 4000 adenosine-moiety compounds, identifying seven molecules with potential binding affinity to METTL3; however, their inhibitory effects in vivo were minimal. Similarly, Moroz-Omori et al.⁶⁴⁶ and Dolbois et al.⁶⁴⁷ reported on adenine-based libraries, identifying UZH1a and UZH2 as compounds that occupy the catalytic pocket of METTL3, suggesting their role as potential competitive inhibitors in vitro. Cpd-564, an METTL3 inhibitor identified from ChemDiv and MCE screening libraries, has shown significant reno-protective effects in mouse models of acute kidney injury induced by cisplatin and ischemia-reperfusion.⁶⁴⁸ Coptisine chloride, identified via molecular docking-based virtual screening from the Vitas-M chemical library, displayed high affinity to METTL3, exerting competitive inhibitory effects by occupying the SAM binding pocket.⁶⁴⁹ STM2457 and STM3006 are novel small molecules that bind non-covalently to the catalytic center of METTL3, reducing its enzymatic activity.^{650,651} Specifically, STM2457 has demonstrated promising antitumor effects and tolerability in mouse models, improving drug resistance to chemotherapy.^{650,652–654} In comparison, although STM3006 exhibits enhanced cellular potency, its in vivo efficacy is constrained by its shorter half-life.⁶⁵¹ In 2023, STC-15, an oral inhibitor optimized from STM2457, became the first and the only RNA m6A target drug to be applied in phase I clinical trials (NCT05584111). Through detailed studies on the spatial structure of the catalytic domain of METTL3, a series of branched, Y-shaped molecules are designed. These were synthesized by integrating chemical fragments from the most effective inhibitors, resulting in molecules with selectivity and binding affinities surpassing those of STM2457, the only commercially available METTL3 inhibitor.⁶⁵⁵ This advancement not only underscores the potential of METTL3 as a therapeutic target but also guides future drug design. Additionally, several natural products with METTL3-inhibitory capabilities have been identified. Quercetin, known as a DNMT inhibitor, has been found to interact with the adenosine moiety pocket in METTL3, forming a stable complex that reduces its catalytic activity.⁶⁵⁶ This interaction decreases METTL3 hyperactivation and lowers m6A levels in protein kinase D2 mRNA, improving insulin sensitivity under palmitic acid stimulation—a benefit in hyperinsulinemia conditions.⁶⁵⁷ Other natural compounds like berberine and curcumin, also noted for DNMT/HDAC inhibition, have shown METTL3 inhibitory activity, though their mechanisms require further clarification.^{658,659} Moreover, molecules F039-0002 and 7460-0250 have been designed to target METTL3's catalytic pocket, showing potential in treating inflammatory bowel disease.⁶⁶⁰ Several candidates identified through silico analysis of South African natural products—SANCDB0370, SANCDB0867, and SANCDB1033—also exhibit METTL3 inhibitory properties, with further validation needed.⁶⁶¹ More recently, Li et al.⁶⁶² designed a stapled peptide inhibitor, RSM3, targeting the PPI at the METTL3-METTL14 interface. This inhibitor offers a unique approach compared to other small-molecule competitive inhibitors, providing a novel avenue for therapeutic intervention.

Allosteric inhibitors prevent METTL3/14-dependent m6A methylation in a non-competitive manner. To date, three allosteric inhibitors have been identified. The first two allosteric inhibitors are CDIBA and CDIBA-43n, which initially function as cytosolic phospholipase A2 inhibitors preventing inflammation. They show an inhibitory effect in the presence of METTL3/14 complex, instead of separate METTL3 and METTL14 subunits.⁶⁶³ The third compound, eltrombopag (previously mentioned as a TET agonist), is recently reported to bind with the METTL3 subunit at an allosteric site and has shown potential in treating AML.^{663,664}

Metformin, traditionally used as a first-line treatment for T2DM, has recently been found to inhibit METTL3 expression, possibly contributing to its beneficial effects in patients with malignant tumors.⁶⁶⁵ The role of metformin in inhibiting METTL3 expression, at the post-transcriptional level, in breast tumors, is first reported in breast cancer.⁶⁶⁶ Subsequently, the application of metformin is also found to inhibit METTL3 expression at the transcriptional level, mediated by the recruitment of DNMT.⁶⁶⁷ This dual action of metformin, combined with chemotherapy, offers potential benefits for patients resistant to traditional chemotherapy, potentially mitigating poor prognoses.^{667,668} Given its safety profile, metformin is a promising candidate as an epigenetic drug targeting METTL3.

METTL3 agonists: While research has predominantly focused on METTL3 inhibitors, there is also interest in agonists, given their potential benefits in DNA damage repair, tumor therapies, and regenerative medicine.^{669–672} In 2019, Selberg et al.⁶⁷³ predicted interactions between four small-molecule ligands with METTL3 involving piperidine and piperazine rings, similar to SAM's binding. These interactions enhanced cell viability and promoted proliferation, although differing onset times among the compounds suggest the need for further development of more effective METTL3 complex activators.⁶⁷³ Melatonin seems to act as an agonist of METTL3. Lv et al.⁶⁷⁴ proposed that melatonin pretreatment can upregulate the expression level of METTL3, restore m6A levels in spermatogonial stem cells, and help them resist the destructive effect of Cr(VI) on reproductive function. However, this viewpoint has recently been questioned. In mouse models with colon inflammation, melatonin inhibits METTL3 expression through melatonin receptor 1B.⁶⁷⁵ Further research is necessary to clarify melatonin's role in METTL3 regulation.

Targeting the eraser of m6A: FTO and ALKBH5. FTO and alKB homolog 5 (ALKBH5) are established m6A erasers, each playing significant roles in epigenetic regulation. FTO is primarily involved in energy homeostasis, demethylating m6A in various RNA species, including cellular mRNA, which impacts multiple biological processes.⁶⁷⁶ ALKBH5 not only demethylates m6A-marked mRNA but also m6A-marked single-stranded DNA (ssDNA), influencing oncogenic or tumor-suppressive activities.⁶⁷⁷ Over the years, numerous small molecules targeting these m6A writers have been identified and designed, showing promising therapeutic efficacy in vitro and in vivo and advancing the development of epigenetic drugs.

FTO inhibitors: Current strategies for developing FTO inhibitors are multifaceted. Based on the spatial structure of FTO, competitive or non-competitive inhibitors that bind to FTO covalently or non-covalently have been developed. With a deeper understanding of FTO functionality, metabolites in vivo possibly related to FTO have been identified, represented by D-2-HG, a metabolite produced by mutant IDH.⁶⁷⁸ Furthermore, exploring the mechanisms underlying medical agents that treat FTO-related diseases provides a theoretical foundation for drug discovery.⁶⁷⁹ Subsequently, optimizing the molecular structures of these initially detected compounds will contribute to the development of FTO inhibitors with high selectivity and inhibitory effects, providing promise for the clinical application of FTO inhibitors in the future. Here, we summarize the typical drugs that inhibit FTO, which are the foundation for developing novel inhibitors through constant iterations.

In 2012, the first FTO inhibitor, rhein, was identified. Rhein impairs FTO activity by disrupting its interaction with ssDNA at the catalytic domain.⁶⁸⁰ Although rhein increases m6A levels in vitro, its weak selectivity and low inhibitory efficacy have limited its clinical potential, highlighting the need for more effective FTO inhibitors.⁶⁸⁰ New FTO inhibitors need to be designed to

overcome these drawbacks. Meclofenamic acid (MA), an FDA-approved nonsteroidal anti-inflammatory drug, binds selectively to similar sites on FTO.⁶⁸¹ MA and its prodrug, MA2, have shown promising results in reversing tumor progression and enhancing the efficacy of chemotherapeutic drugs, significantly prolonging survival.^{681,682} Inspired by MA, various compounds have been developed, such as the fluorescein derivative FL1, which retains the benzyl carboxylic acid structure critical for interaction with FTO. The complex formed between FTO and FL1 inhibits the enzyme's activity and facilitates the study of FTO-related signaling pathways through fluorescein labeling.⁶⁸³ Other optimization molecules include GNPIPP12-MA,⁶⁸⁴ 13a,⁶⁸⁵ FB23/FB23-2,⁶⁸⁶ Dac-51/Dac-85,⁶⁸⁷ ZLD115,⁶⁸⁸ and FTO-02/FTO-04/FTO-43.^{689,690} These compounds significantly improve MA in inhibitory activity, cell permeability, and biosafety while reducing off-target effects and potential resistance. Another similar mechanism inhibitor is diacerein, a structural analog of rhein, which has shown antitumor effects in breast cell lines.⁶⁹¹

In addition to interfering with the interaction between ssDNA and FTO, inhibitors of this enzyme also compete with cofactors such as α -KG and iron(II). For instance, fumarate hydrazide 2 and compounds with the aminohydroxyfuranone core exemplify this approach.^{692,693} Furthermore, the discovery of N-CDPCB, a competitive inhibitor that binds to non-conserved fragments of FTO, provides novel insights into the development of inhibitory agents. Mechanistically, compounds like benzene-1,3-diol and 4-Cl-1,3-diol are crucial in mediating and enhancing the specific interaction between FTO and N-CDPCB.⁶⁹⁴ Additional potential inhibitors, such as CHTB and radicicol, have been identified through virtual screening; these compounds have similar structures.^{695,696} However, related evidence is lacking to exhibit their efficacy. Moreover, Su et al.⁶⁹⁷ reported on CS1 and CS2, which tightly bind to the catalytic pocket of FTO, activating immune checkpoint genes and reversing immune evasion in tumor diseases. Clausine E, another FTO inhibitor, targets the enzyme's hydrophobic cavity, exhibiting antitumor activity.⁶⁹⁸

These ongoing discoveries provide a deeper understanding of the diverse structures of molecules interacting with FTO and their mechanisms of action, promoting large-scale virtual screenings to identify more potential inhibitors. For example, mupirocin, entacapone, compounds "18,077" and "18,079", several quinolone derivatives, and a series of 1,2,3-triazole analogs have been identified as potential FTO inhibitors.^{699–703} Notably, quinolone derivatives and their antitumor properties have shown the potential to improve symptoms in neurodegenerative diseases by inhibiting FTO activity.⁷⁰² These findings broaden the potential clinical applications of FTO inhibitors.

FTO agonists: Recent studies have identified that certain tricyclic antidepressants, such as imipramine and amitriptyline, exert their antidepressant effects by activating FTO in N2a cells.⁷⁰⁴ This emerging area of research highlights the potential therapeutic benefits of FTO activators and calls for more attention to their development and evaluation.

ALKBH5 inhibitors: The RNA demethylase ALKBH5 is recognized as a pro-oncogene, playing a vital role in the post-transcriptional regulation of various targets in cancer biology.⁶⁷⁷ Interest in targeting ALKBH5 for therapeutic purposes has significantly increased. We classify the identified ALKBH5 inhibitors into three main categories based on their mechanisms of action. The first category comprises typical competitive inhibitors that compete with cofactors for binding sites. These agents consist of IOX1 (also known as a TET/KDM inhibitor), MV1035, and Ena21, which exhibit the therapeutic potential of targeting ALKBH5 in antitumor therapies.^{705–707} Compounds that non-covalently interact with the active pocket of ALKBH5 are classified as the second group. Through structure-based virtual screening and optimization, the

current compounds include DDO-2728, 2-((1-hydroxy-2-oxo-2-phenylethyl)thio)acetic acid, and 4-((furan-2-ylmethyl)amino)tetrahydropyridazine-3,6-dione.^{708,709} The third category includes molecules that bind to the m6A-binding pocket of ALKBH5, directly disrupting the interaction between the enzyme and its substrates. For instance, compounds 20m and TD19 are representative of this type.^{710,711} Some compounds still exist whose potential mechanisms for inhibiting ALKBH5 have not been elucidated, such as ALK-04, Ena15, ZINC78774792, and ZINC00546946, although their antiproliferative effects have been revealed in vitro and in vivo.^{707,712,713} In brief, the continued development and in-depth research into ALKBH5 inhibitors hold significant potential for disease treatment, necessitating further efforts.

Targeting the reader of m6A: IGF2BP and YTH domain family. The discovery of m6A readers with specific motifs has spurred significant interest in developing drugs targeting these proteins, expanding the possibilities for therapeutic interventions.

IGF2BP inhibitors: Insulin-like growth factor 2 mRNA-binding proteins (IGF2BPs) are newly identified m6A readers that enhance the stability and maintenance of their target mRNAs.⁷¹⁴ IGF2BP plays an oncogenic role in various cancers, making its inhibition a promising strategy for antitumor therapy.⁷¹⁵

Six IGF2BP inhibitors have been developed, demonstrating antitumor effects by disrupting IGF2BP-RNA interactions. BTYNB, the first identified IGF2BP1 inhibitor, suppresses melanoma and ovarian cancer cell proliferation by blocking IGF2BP1's interaction with c-Myc mRNA.⁷¹⁶ BTYNB's therapeutic effects are being studied across various tumor models, including esophageal squamous carcinoma,⁷¹⁷ neuroblastoma,⁷¹⁸ and cholangiocarcinoma.⁷¹⁹ CWI1-2 and JX5 are novel IGF2BP2 inhibitors with antileukemic activities that inactivate the Notch1 signaling pathway. CWI1-2 forms a hydrophobic interaction with IGF2BP2's RNA-binding core, while JX5 binds tightly to the protein. Further research is needed to enhance their safety and reduce cytotoxicity.^{720,721} Cucurbitacin B, a natural product, exerts a pharmacological allosteric effect on IGF2BP1. In hepatocellular carcinoma mouse models, it modifies IGF2BP1's configuration, reducing its efficacy.⁷²² Another compound, "7773," specifically disrupts the IGF2BP1-Kras mRNA interaction, effectively inhibiting IGF2BP1's pro-oncogenic activity.⁷²³ Isoliquiritigenin is the only small molecule identified targeting IGF2BP3.⁷²⁴ Derived from the Chinese herb licorice, it downregulates IGF2BP3 expression, showing promise in treating non-small cell lung cancer.⁷²⁴

YTH domain family inhibitors: YTH domain family (YTHDF) comprises a group of readers featuring a YTH domain at the C-terminus. This domain forms a hydrophobic pocket essential for recognizing m6A modifications.⁷²⁵ Elevated levels of YTHDF proteins have been associated with the progression of various cancers.⁷²⁶ Conversely, reducing these proteins can synergistically enhance the effectiveness of ionizing radiation and anti-PD-L1 therapies in reducing cancer burdens.^{727,728} This underscores the potential of YTHDF inhibitors as a promising direction for improving antitumor treatments.

The binding sites between YTHDF proteins and m6A modifications are primary targets for most YTHDF inhibitors.⁷²⁹ The successful elucidation of the crystallographic structures of the YTH domains in YTHDF proteins has provided critical opportunities for drug design.⁷³⁰ High-throughput screening technology has identified three small molecules—ebesen, DC-Y13, and DC-Y13-27—as effective YTHDF inhibitors.^{727,731} Ebesen targets YTHDF1 and YTHDF2, either covalently or non-covalently binding to the YTH domain.⁷³¹ DC-Y13 and DC-Y13-27, particularly the latter, act as selective inhibitors of YTHDF2, offering therapeutic benefits.⁷²⁷ Additionally, studies have shown that disrupting O-GlcNAcylation

Table 6. Summary of chromatin remodeling-targeted drugs for different diseases in clinical trials

| Drug | Target(s)/ mechanisms(s) | Condition(s) | Status/outcome(s) | Phase(s) | Other intervention(s)/ drug(s) | Study ID |
|---------|--------------------------------|---|--|------------|---|-------------|
| FHD-286 | SMARCA4/2 allosteric inhibitor | Metastatic uveal melanoma | Terminated (due to business reasons) | Phase I | — | NCT04879017 |
| FHD-286 | SMARCA4/2 allosteric inhibitor | AML, MDS, CMML | Recruiting | Phase I | With or without low-dose Cytarabine or Decitabine | NCT04891757 |
| PRT3789 | PROTACs-based SMARCA2 degrader | NSCLC and other solid tumors with SMARCA4 gene mutation | Recruiting | Phase I | With or without Docetaxel | NCT05639751 |
| FHD-609 | PROTACs-based BRD9 degrader | Advanced synovial sarcoma | Terminated (due to sponsors' decision) | Phase I | — | NCT04965753 |
| CFT8634 | PROTACs-based BRD9 degrader | Synovial sarcoma and other SMARCB1-perturbed soft tissue sarcomas | Terminated (no significant clinical activity with CFT8634 as a single agent) | Phase I/II | — | NCT05355753 |

AML acute myeloid leukemia, BRD9 bromodomain containing 9, CMML chronic myelomonocytic leukemia, MDS myelodysplastic syndrome, NSCLC non-small cell lung cancer, PROTAC proteolysis-targeting chimeras, SMARCA4 SWI/SNF-related, matrix-associated, actin-dependent regulator of chromatin A4

of YTHDF proteins can also decrease their stability and enzymatic activities, providing new avenues for identifying YTH-inhibiting small molecules.^{732,733}

Epigenetics-targeted drugs and chromatin remodeling

SWI/SNF complexes are intricate multimeric structures composed of diverse, variable subunits that play distinct roles, emphasizing the importance of personalized characteristics and frequent mutations in these subunits in various human diseases. Recently, the design of small molecules targeting different components of the SWI/SNF complex has expanded, yielding numerous potential therapeutic interventions. Those that have progressed to clinical trials are detailed in Table 6.

The active DNA-dependent ATPase A domain inhibitor (ADAADI) was the first discovered inhibitor targeting the SWI/SNF complex. It was identified during studies on mammalian cell resistance to certain antibiotics in vitro.⁷³⁴ ADAADI binds to specific motifs in the enzyme complex, inducing conformational changes that inhibit SWI2/SNF2's catalytic activities.⁷³⁴ Currently, ADAADI shows promising therapeutic effects in prostate cancer in preclinical studies, laying the groundwork for further development of SWI/SNF-targeted epigenetic drugs.⁷³⁵

Research has also focused on specific inhibitors targeting the SWI/SNF-related, matrix-associated, actin-dependent regulator of chromatin A4 (SMARCA4) and its paralog SMARCA2, which are DNA-stimulated ATPases within the SWI/SNF complexes. SMARCA4, commonly mutated in various tumors, is associated with reduced sensitivity to traditional cancer treatments.^{736,737} Inhibiting SMARCA4/2 is an effective strategy for curbing tumor growth and improving patient outcomes. Papillon et al.⁷³⁸ reported the earliest selective allosteric inhibitors of the SMARCA4/SMARCA2 subunits, with confirmed effects on pediatric H3K27M diffuse midline glioma and AML in both in vivo and in vitro settings.^{739,740} FHD-286, a novel orally bioavailable SMARCA4/SMARCA2 allosteric inhibitor, has shown preclinical efficacy. Combined treatment with FHD-286 and other epigenetic drugs, such as decitabine, BET inhibitors, and menin inhibitors, has demonstrated synergistic effects in reducing AML burden without significant toxicity.⁷⁴¹ Notably, FHD-286 has entered clinical development for treating various malignant tumors, including metastatic uveal melanoma (NCT04879017) and several malignant hematological disorders (NCT04891757).

An alternative approach to inhibiting SMARCA4/SMARCA2 involves using specific inhibitors that target their BDs. This

strategy extends to other BD-containing proteins within the SWI/SNF complexes, such as polybromo-1 (PBRM1), BD containing 7 (BRD7), and BRD9, also considered promising targets for epigenetic drug development. Notably, SMARCA4/SMARCA2 and BRD9/BRD7 each contain one BD, whereas PBRM1 contains six tandem BDs, providing numerous potential interaction points for inhibitors.⁷⁴² Current research primarily focuses on inhibitors for family VIII BD in SMARCA4/SMARCA2 and PBRM1, with four major classes of inhibitors reported: salicylic acid fragment hits such as PFI-3;^{743–745} aminopyridazines represented by GNE-064;⁷⁴⁶ quinoxalines represented by LM146,^{747,748} and dihydroquinazolinones represented by compound16 and GNE-235.^{749,750} These inhibitors are categorized as either pan-inhibitors, affecting multiple proteins, or selective inhibitors, targeting specific proteins. PFI-3, its analogs, and GNE-064 are pan-inhibitors, whereas LM146 shows a higher affinity for SMARCA2, and compound16 and GNE-235 are selective for PBRM1. The therapeutic applications of these inhibitors, particularly the pan-inhibitors, have been extensively studied in various diseases.^{751–755} However, the efficacy of PFI-3 as a standalone treatment for malignancies has been less satisfactory. The application of compound16, on the other hand, demonstrates promising therapeutic effects in PBRM1-dependent prostate cancer, suggesting its potential as a foundational treatment for PBRM1-driven cancers.⁷⁴⁹ As for the other molecules, though their binding ability and inhibitory effects have been validated at the molecular level, sufficient evidence is still lacking in vivo or in vitro to demonstrate their clinical value. Furthermore, targeting family IV BD of BRD9 and BRD7 has led to the development of many selective inhibitors. Current research includes inhibitors like BI-7271,⁷⁵⁶ BI-7273,⁷⁵⁶ BI-9564,⁷⁵⁶ I-BRD9,⁷⁵⁷ iBRD9,⁷⁵⁸ GNE-375,⁷⁵⁹ and newly identified inhibitors developed through integrated computational approaches.⁷⁶⁰ Selective inhibitors for BRD7, such as 1-78 and 2-77,⁷⁶¹ and molecules like LP99, TP-472, 4-acylpyrroles, and GSK6776, which inhibit both BRD9 and BRD7, are being evaluated for their therapeutic effects in various pathologies.^{762–765}

PROTAC technology plays a significant role in developing SWI/SNF inhibitors, with novel agents such as AU-24118 and AU-15330 being tested in preclinical studies and clinical trials. AU-24118 and AU-15330 are degraders targeting family VIII BD in SMARCA4/SMARCA2 and PBRM1, which are valuable tools in castration-resistant prostate cancer treatment.^{766,767} AU-24118 has shown promise in inducing tumor regression at therapeutic doses.⁷⁶⁶

Table 7. Summary of non-coding RNA drugs for different diseases in clinical trials

| Drug | Target/ Mechanism | Condition(s) | Status/outcome(s) | Phase(s) | Other intervention(s)/ drug(s) | Study ID/references |
|-----------|-------------------------|--|---|----------------|--|---------------------------------|
| MRG-106 | MiR-155 inhibitor | CTCL, MF, CLL, DLBCL, and ATCL | Completed (unpublished) | Phase I | Stable background therapy (simultaneously applied in few participants) | NCT02580552 |
| MRG-106 | MiR-155 inhibitor | CTCL, MF | Terminated (due to business reasons) | Phase II | Vorinostat (active comparator) | NCT03713320 |
| MRG-106 | MiR-155 inhibitor | CTCL, MF | Terminated (due to eligible subjects receiving treatment in a crossover arm of NCT03713320) | Phase II | — | NCT03837457 |
| MRG-110 | MiR-92a-3p inhibitor | Healthy volunteers | Completed (unpublished) | Phase I | Placebo-controlled | NCT03603431 |
| MRG-110 | MiR-92a-3p inhibitor | Healthy volunteers | Completed (significant inhibition on targeted miRNA in vivo) | Phase I | Placebo-controlled | NCT03494712 ¹¹⁵¹ |
| RG-012 | MiR-21 inhibitor | AS | Completed (unpublished) | Phase I | — | NCT03373786 |
| MRG-201 | MiR-29b mimic | Healthy volunteers | Completed (unpublished) | Phase I | Placebo-controlled | NCT02603224 |
| MRG-201 | MiR-29b mimic | Keloid | Completed (exhibits therapeutic effects and manageable adverse events) | Phase II | Placebo-controlled | NCT03601052 |
| MRX34 | MiR-34a mimic | Liver cancer, SCLC, lymphoma, melanoma, MM, RCC, NSCLC | Terminated (due to serious immune- related adverse events) | Phase I | — | NCT01829971 ^{804,1152} |
| MRX34 | MiR-34a mimic | Melanoma | Withdrawn (due to immune-related serious adverse events in the phase I study) | Phase I/ II | Dexamethasone premedication | NCT02862145 |
| TargomiRs | MiR-16 mimic | MPM, NSCLC | Completed (exhibits acceptable safety profile and early signs of therapeutic activity) | Phase I | — | NCT02369198 ¹¹⁵³ |
| INT-1B3 | MiR-193a- 3p mimic | Advanced solid tumors | Terminated (due to the insufficient funding) | Phase I | — | NCT04675996 |

AS Alport syndrome, ATCL adult T cell leukemia/lymphoma, CLL chronic lymphocytic leukemia, CTCL cutaneous T cell lymphoma, DLBCL diffuse large B cell lymphoma, MF mycosis fungoides, MiR microRNA MM multiple myeloma, MPM malignant pleural mesothelioma, NSCLC non-small cell lung cancer, RCC renal cell carcinoma, SCLC small cell lung cancer

However, long-term treatment at high doses can lead to mutations in the BD and overexpression of ATP-binding cassette subfamily B member 1 (ABCB1), which contributes to drug resistance development.⁷⁶⁶ Combining these treatments with ABCB1 inhibitors could potentially mitigate resistance to SMARCA4/SMARCA2 inhibitors in vivo. Additionally, applying PROTACs to previously reported inhibitors can enhance their selectivity and reduce off-target effects. For instance, the linkage of BI-7273 with an E3 ubiquitin ligase has led to the design of dBRD9-A, the first BRD9-directed degrader, which is undergoing optimization.^{768,769} The current focus on dBRD9-A, which is being tested for its efficacy in AML, MM, and interferon-induced inflammation in animal models, highlights its potential as a promising therapeutic for both tumor and non-tumor conditions.^{768,770,771} Additional PROTACs-based SWI/SNF inhibitors, such as those derived from dihydropyrrolo-quinazolin scaffolds (targeting SMARCA4, SMARCA2, and PBRM1),⁷⁴⁷ A947 (targeting SMARCA2),⁷⁷² VZ-185 (targeting BRD9 and BRD7),⁷⁷³ CFT8634 as well as FHD-609 (targeting BRD9), further illustrate the breadth of ongoing research.⁷⁷⁴ Notably, FHD-609 have recently advanced to clinical trials for synovial sarcoma (NCT04965753), underscoring the critical role of drug development targeting mutations in SWI/SNF complexes. Moreover, CFT8634 was originally planned for investigation in a phase 1/2 clinical trial of synovial sarcoma and other SMARCB1-perturbed soft-tissue sarcomas (NCT05355753). However, the clinical trial was terminated because of the less significant clinical activity of CFT8634 as a single agent.

Considering the prevalence of mutations in SWI/SNF complexes in cancers, continued research into the in vivo therapeutic effects, potential applications, and long-term risks of these drugs is essential for assessing their clinical utility.

Epigenetics-targeted drugs and non-coding RNA

A deep understanding of ncRNA's role in disease progression, particularly in various cancers, has led to innovative epigenetic strategies for disease management.⁷⁷⁵ RNA interference (RNAi) technologies, which utilize small double-stranded RNA to selectively interact and degrade specific intracellular RNAs, mimic gene deletion phenotypes.^{776,777} RNAi-based therapies targeting ncRNA are categorized based on their action mechanisms and intended outcomes: silencing overexpressed ncRNAs to curb disease-related expressions, restoring downregulated ncRNAs to regain lost functions, and blocking ncRNA localization to prevent ncRNA from functioning by interfering with its subcellular localization. RNAi-oriented drugs altering ncRNA patterns have been widely studied and applied in clinical practice (Table 7). Herein, we summarize the emerging technologies for ncRNA-targeted agent development, aiming at supplementing the current understanding of drug design.

Significant advancements in molecular editing and delivery systems have bolstered the clinical viability of these innovative ncRNA-targeted therapies.^{778,779} One pivotal development has been the chemical modification of synthetic nucleic acids to enhance their stability and delivery efficiency. The initial focus on

replacing phosphodiester bonds with phosphorothioate has been a cornerstone in numerous FDA-approved oligonucleotide therapies,⁷⁸⁰ although concerns about inflammation and toxicity in vivo have prompted research into alternative modifications for RNA-targeted based on RNAi.⁷⁸¹ Over the years, various modification strategies have emerged, including those based on 2'-O-methyl, 2'-O-methoxyethyl, 2'-fluoro, and n-acetylgalactosaminy, aiming to preserve the therapeutic attributes of nucleic acids while enhancing their stability.⁷⁸² Locked nucleic acids (LNAs) represent a notable innovation, linking the 2' and 4' carbons of ribose rings with methylene bridges, thus improving hybridization affinity and resistance to nucleases.⁷⁸³ Cobomarsen, an LNA-based inhibitor targeting miR-155, exemplifies this technology's potential, having shown promising results in preclinical studies for hematological malignancies and solid tumors and exhibiting positive effects in mycosis fungoides patients.^{784–788} In addition, the therapeutic efficacy of cobomarsen has been further validated in patients with mycosis fungoides, indicating well-tolerated and positive clinical potentials (NCT03713320).

Furthermore, integrating nanomedicine-based delivery systems, such as lipid-based, polymeric, inorganic, and biomimetic nanoparticles, has significantly advanced the development of RNAi drugs. These delivery techniques enhance the stability and bioavailability of oligonucleotide drugs and improve their efficacy in modulating target ncRNA expression and functionality.⁷⁸⁹ The recent emphasis on nanoparticles designed for targeted delivery of therapeutic nucleic acids to specific subcellular organelles marks a significant advancement in ncRNA therapy. As ncRNAs are present not only in the cytoplasm but also the nucleus and various organelles, targeting these subcellular locations can enhance the efficacy of treatments.^{790,791} Researchers are exploring opportunities to integrate subcellular organelle-targeting signals into nanoparticle delivery systems. Current studies have reported RNAi nanoparticle systems designed to target the nucleus, mitochondria, endoplasmic reticulum, and Golgi apparatus.^{792–795} Though nucleus-targeted nanoparticles have been studied in regulating ncRNAs, only a few of them have proposed the incorporation of nucleus-targeting TAT peptide and nucleus-targeting peptide amphiphile into nanoparticle delivery systems to achieve active transportation.⁷⁹² Many researchers have only reported the high concentrations of therapeutic oligonucleotides in the nucleus without elucidating the mechanisms involved in nucleus-targeting. This approach leaves a gap in our understanding of the complex and precise intracellular delivery processes that involve biomembrane systems and cytoskeletal interactions.⁷⁹⁶ The variability in cell types, delivery materials, and therapeutic nucleic acids means that successful results in specific contexts may not universally apply, highlighting the challenges in translating these strategies from experimental to clinical settings.

Additionally, the integration of clustered regularly interspaced short palindromic repeats (CRISPR)/CRISPR-associated system (CRISPR/Cas) technologies, particularly CRISPR/Cas9, into ncRNA research offers novel avenues for manipulating ncRNA expression, including lncRNAs and microRNAs (miRNAs).^{797–800} CRISPR/Cas9's flexibility and high specificity make it an advantageous tool for gene-targeted cancer therapies, such as CRISPR interference, activation, and knockout strategies, now moving into preclinical trials. This gene-editing technology promises to enhance the efficacy of traditional cancer treatments and aims to minimize off-target effects, thus fostering the development of personalized medicine.^{801,802}

However, several challenges persist despite the growing number of ncRNAs identified as potential therapeutic targets. Technological advancements are still needed, and safety concerns must be rigorously addressed.⁸⁰³ The premature termination of the Phase I clinical trial for MRX34, a liposomal mimic of miR-34a, due to severe immune reactions and fatalities, underscores the

critical need for comprehensive preclinical data and cautious progression to clinical trials.⁸⁰⁴

PRACTICAL CHALLENGES IN THE EPIGENETIC DRUG DEVELOPMENT AND APPLICATION

Massive efforts have been made in epigenetics-targeting drug development, whether approved in different countries for the treatment of specific indications or currently identified and further evaluated in fundamental experiments or clinical trials, exhibiting notable potential. However, there are still practical issues that should be dealt with when applying them to a large scale in clinical practice.

First of all, a thorough understanding of epigenetic mechanisms is crucial for successfully applying epigenetic-targeted drugs. Alterations in epigenetic enzymes can change DNA, histones, or chromatin structures, impacting cellular processes like transcription, replication, and DNA repair. Even minor modifications in epigenetic enzyme activity can significantly affect cellular functions. Therefore, deepening our understanding of epigenetic biology is essential. However, gaps remain in our knowledge, particularly regarding the roles of epigenetic regulation and its proteins in mammals, such as new DNA modifications like m6A, RNA modifications beyond m6A, novel reader domains, and PPI networks related to epigenetic regulation. These areas represent potential targets for novel epigenetic drugs but require further exploration.^{805–810} Second, many molecules identified through virtual screening and molecular docking techniques are primarily used as molecular probes to study enzyme localization and function. However, transitioning from a potent molecular probe to a viable therapeutic agent involves rigorous in vivo evaluation to confirm their inhibitory effects and therapeutic potential. This step is crucial as it determines whether these compounds can be safely and effectively used in clinical settings.^{811,812} These small molecules are potent probes to detect the localization and function of targeted enzymes, and their potential for medical use needs confirmation. Notably, another challenge that needs to be overcome is associated with the precise localization and targeted activity of epigenetics-targeted drugs is critical for enhancing their clinical efficacy and reducing potential side effects. The need for precise delivery of epigenetic drugs to subcellular structures is underscored by the diverse localizations and functions of their target enzymes and ncRNAs. For instance, enzymes responsible for histone modifications are found both in the nucleus, where they modify histones, and in the cytoplasm, where they regulate non-histones and related signaling pathways at the post-translational level.^{813,814} Similarly, the function of ncRNAs depends significantly on their localization and distribution within the cell.⁷⁹⁰ To address these challenges, a deeper understanding of the intracellular trafficking mechanisms is required. Additionally, developing sophisticated drug delivery systems, such as lipid-based nanoparticles or targeted delivery vehicles, can enhance the specificity and efficiency of these therapies by directing them to their precise intracellular sites of action. Furthermore, the challenge of achieving selective inhibition within the HDAC family exemplifies the broader issue of specificity in drug design. Many drugs targeting epigenetic enzymes exhibit pan-inhibitory effects, leading to significant off-target effects and adverse reactions, particularly when the drugs indiscriminately affect multiple members of an enzyme family.⁴⁰⁸ This necessitates the development of more selective inhibitors that specifically target single proteins or subfamilies of proteins. By focusing on selective inhibition, researchers can potentially improve the safety and efficacy of these treatments, minimizing unwanted interactions and enhancing their therapeutic impact.

Importantly, the possibility of developing resistance to epigenetic-targeted drugs, which is another factor limiting their further application, cannot be ignored. Several studies have

investigated the mechanisms involved in the development of resistance to epigenetic agents in different cancers.^{815–817} The activation of certain signaling pathways and gene mutations in tumor cells play an indispensable role in inducing the development of drug resistance to epigenetic agents. Research has shown that the activation of enhanced Wingless/Integrated (Wnt)/ β -catenin signaling contributes to developing resistance to BET inhibitors in leukemia cells. However, inhibition of this pathway helps rescue drug sensitivity in vitro and in vivo.^{818,819} Furthermore, enhanced activity of the protein kinase B (AKT)/mTOR complex 1 (mTORC1) signaling pathway is also responsible for drug resistance to BET inhibitors in prostate cancer.⁸²⁰ In another study on HDAC resistance in solid tumors, the potential role of the activation of some kinases and downstream pathways was also reported.⁸²¹ Additionally, altered TME may be one of the culprits in promoting drug resistance. Tumor-associated macrophage (TAM), a pivotal mediator in inducing tumor cells to develop resistance to traditional antitumor therapies, has recently been proposed to be involved in the occurrence of epigenetic drug resistance.^{822,823} A 2020 study on triple-negative breast cancer reported that interleukin-6 (IL-6) and IL-10 derived from TAM activated STAT3 signaling in tumor cells, conferring them with drug resistance to BET inhibitors.⁸²³ Additionally, these findings pave a theoretical foundation for the combination of epigenetics-targeted drugs and other pharmaceutical molecules to optimize their long-term efficacy.⁸²⁴ The relationship between drug resistance to epigenetic agents and mutations in certain genes is observed in tumor cells, especially in the case of applying EZH2 and IDH inhibitors.^{59,825,826} Based on this idea, the loss-of-function mutation of specific genes by CRISPR/Cas techniques may provide better platforms for coping with drug resistance.

In conclusion, for epigenetics-targeted drugs, it is crucial to balance pharmacokinetics (how the drug is processed in the body), tolerability (how well the drug is tolerated), and therapeutic efficacy (how to avoid off-target off-target toxicities and resistance). Developing optimal dosing regimens that maximize efficacy while minimizing side effects and resistance requires a thorough understanding of the drug's behavior in the body, including its absorption, distribution, metabolism, and excretion. Innovative dosing strategies, possibly involving controlled release formulations or real-time monitoring of drug levels, could play a vital role in achieving this balance. Continued research into the biological and pathological roles of targets for epigenetic drugs is essential.

DEVELOPING TRENDS AND FORTHCOMING PROSPECTS IN EPIGENETICS-TARGETED DRUGS

Research on epigenetics-targeted drugs has progressed rapidly, with a growing focus on their potential as next-generation clinical candidates. Current trends, illustrated in Fig. 5, emphasize the synergy between these agents and other therapeutic modalities such as chemotherapy, radiotherapy, kinase inhibitors, and immunotherapy. This integration promotes precision medicine and personalized treatment strategies and enhances the overall effectiveness of cancer therapies.^{827,828} Furthermore, developing epigenetic degraders, which can hydrolyze targeted proteins, complements the inhibitory functions of traditional epigenetic drugs.⁸²⁹ Notably, the swift advancement of sequencing technology has empowered the detection of epigenetic irregularities with growing efficacy, substantially enhancing the integration of epigenetics into personalized medicine.

Epigenetics-based combination therapy in cancer cells
The integration of epigenetics-targeted drugs with conventional cancer therapies—such as chemotherapy, targeted therapy, immunotherapy, and hormone therapy—is emerging as a promising strategy for cancer treatment. Increasing experimental

studies and clinical trials are assessing the safety and efficacy of various combination regimens. The benefits of these combinations can be categorized into two primary aspects:

Epigenetic drugs can synergize with other cancer therapies by modulating the metabolic and pathological characteristics of cancer cells, immune cells, and stromal cells within the TME.⁸³⁰ Although immune checkpoint inhibitors (ICIs), which target immune checkpoint proteins (ICPs), such as PD-L1, PD-1, and CTLA-4, show significant potential, their effectiveness may be limited by factors such as insufficient antigen presentation and suboptimal T cell responses in the TME.^{831,832} Epigenetic modifications can enhance the expression of tumor antigens and ICPs, overcoming these limitations. The use of epigenetic therapies not only disrupts immunosuppressive pathways but also enhances the recruitment of tumor-reactive immune cells, resulting in synergistic effects with ICIs.^{136,833–835}

Interfering with aberrant epigenetic features is crucial for combating drug resistance, a major challenge in oncology. Chemoresistance often correlates with changes in DNA methylation and histone acetylation, among other epigenetic characteristics.^{139,836,837} Combining chemotherapeutic agents with epigenetic drugs has become an important strategy to address resistance,^{838,839} also helping to mitigate chemotherapy-related side effects.^{840,841} Additionally, reversing epigenetic alterations in chemoresistant tumor cells can restore their sensitivity to conventional therapies, offering a renewed opportunity for treatment.⁸⁴² In the context of targeted therapy, inhibitors of mutant kinases initially provide rapid benefits but often lead to the development of resistance over time.⁸⁴³ The potential of epigenetic treatments, particularly HDAC and DNMT inhibitors, to reverse such resistance is currently being explored.^{844,845} The development of dual inhibitors, like CUDC907, CUDC101, and 4SC-202, shows promising results in overcoming resistance in kinase-driven cancers and warrants further investigation.⁸⁴⁶ For hormone-dependent cancers, such as estrogen receptor-positive breast cancer and androgen receptor-positive prostate cancer, endocrine therapy remains a crucial treatment option.⁸⁴⁷ However, epigenetic alterations can lead to resistance during endocrine therapy.^{848,849} Targeting these epigenetic changes can help sustain the effectiveness of endocrine therapies and reduce the proliferation of cancer cells.^{848,850}

At present, various combination regimens based on HMA and traditional anticancer drugs have entered clinical trials, gaining the potential to become an alternative for patients with certain diseases. For example, the combination of the oral B-cell leukemia/lymphoma 2 inhibitor venetoclax with HMAs has become a standard regimen among patients with AML or MDS who are ineligible for intensive chemotherapy. In November 2018, the FDA approved this combination for AML therapy.^{851–853} Further, triplet regimens that include HMAs, venetoclax, and other targeted agents are being developed for AML with specific gene mutations.^{229,854–856} Early results from these clinical trials have demonstrated a good safety profile and promising effects, with ongoing studies needed to confirm their efficacy and potential adverse events. In May 2022, the combination of ivosidenib, an IDH1 inhibitor, and azacitidine was approved by the FDA for older patients with newly diagnosed IDH1-mutated AML.²⁴⁰ Other drugs being combined with HMAs include HDAC inhibitors, polo-like kinase 1 inhibitors, T-cell immunoglobulin domain and mucin domain-3 antibodies, and PD-L1 antibodies.^{857–860} These combinations are currently under evaluation in ongoing registrational clinical trials across different stages, with promising results anticipated for updating clinical strategies. Additionally, the combination of HMAs with targeted therapy and immunotherapy, as well as chemotherapy, is showing promising application prospects, especially in hematologic malignancies with acquired chemoresistance caused by aberrant DNA methylation.^{861,862} Currently, azacitidine is approved in multiple countries and is

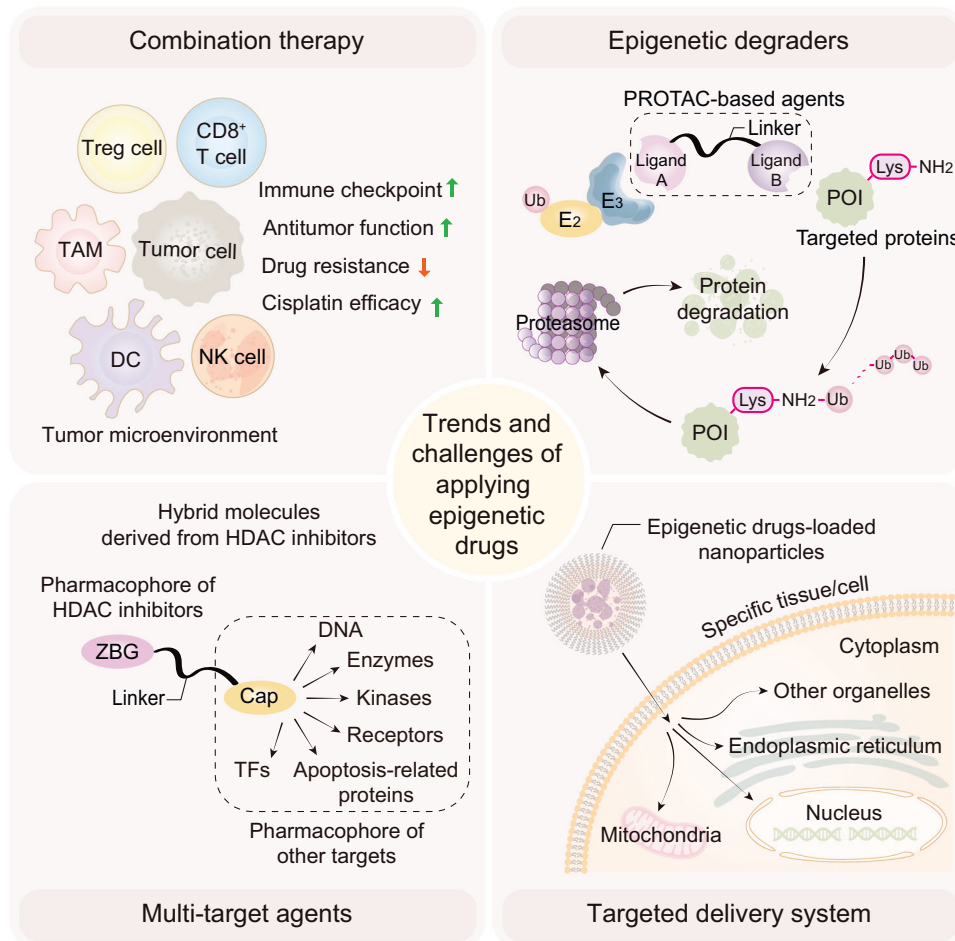


Fig. 5 The promising trends and practical challenges in the clinical application of epigenetics-targeted drugs. From the perspective of clinical practice, epigenetic agents are expected to become promising adjuvants in combination with traditional antitumor therapeutics, contributing to superior efficacy and decreased resistance. Based on this idea, multitarget anticancer agents inhibiting both HDAC and other pathological molecules have gained much attention as a new strategy. Further, epigenetic degraders based on PROTAC or other techniques responsible for TPD help supplement the catalytic function of epigenetic inhibitors. Notably, the successful transport of epigenetic regulators to specific tissues or cells, or even the finite subcellular structures, is the prerequisite for exerting therapeutic effects. The further optimization of different types of nanoparticles makes them inspiring tools for the delivery system

widely used in patients with myeloproliferative disorders, such as MDS, AML, chronic myelomonocytic leukemia (CMML), and juvenile myelomonocytic leukemia, whereas decitabine is approved for the treatment of MDS, AML, and CMML.^{863–865}

Small molecules serving as epigenetics-targeted degraders As an emerging therapeutic strategy, epigenetics-targeted degraders for targeted protein degradation (TPD), respected by molecules based on PROTAC, autophagy-targeting chimera (AUTAC), hydrophobic tagging (HyT), molecular glue (MG), and other novel techniques for drug discovery are worth trying as a remarkable alternative presenting pioneering approaches.⁸⁶⁶

PROTACs, first proposed in 2001, are considered revolutionary technologies in drug discovery. They consist of a ligand for targeted proteins, a ligand for E3 ubiquitin ligase, such as Von Hippel-Lindau and Cereblon (CRBN), and a linker connecting the two ligands.⁸⁶⁷ Many degraders targeting diverse epigenetic enzymes have been developed, including the newly designed or derived from the optimization of known selective inhibitors.^{868,869} This technology has led to the creation of various degraders targeting a wide range of epigenetic enzymes, enhancing their potency, duration of action, safety profile, and ability to counteract resistance mechanisms compared to conventional

inhibitors.^{867–869} For instance, dBET1, derived from the BRD inhibitor JQ-1, was the first PROTAC-based degrader targeting BET proteins. It has demonstrated superior anticancer effects in both AML cell lines and mouse xenograft models compared to JQ-1 alone.^{836,870} Further optimization led to dBET6, which increased cell permeability significantly improved survival rates in solid tumor models, and reduced the emergence of resistance.⁸³⁶ Innovative approaches to enhance the selectivity and safety of PROTACs include antibody-PROTAC technology, which combines monoclonal antibodies targeting specific pathological cells with degraders. This strategy facilitates targeted delivery, minimizing side effects while maximizing efficacy. Antibody-PROTACs have been developed for breast cancer cells overexpressing human epidermal growth factor receptor 2 and prostate cancer cells expressing six transmembrane epithelial antigens of the prostate 1, showing enhanced degradation specificity in these cell types.^{871,872} Compared with general degradation agents, these small molecules present the preferential degradation of target proteins in specific cell lines. Additionally, integrating control elements into PROTAC molecules allows for activation in specific physiological or pathological conditions, reducing potential off-target effects.⁸⁷³ Techniques such as photocaged PROTACs, photo-switchable PROTACs, and radiotherapy-triggered PROTAC

prodrugs represent cutting-edge strategies in this area. These methods ensure that the degradation activity of PROTACs can be spatially and temporally controlled, enhancing their clinical applicability and safety.^{874–876} Overall, the evolution of PROTACs and other PROTACs-oriented TPD strategies is shaping a promising future for epigenetics-targeted therapies, offering more precise and effective treatment modalities for various diseases, particularly cancer.

Other types of epigenetics-targeted degraders, such as AUTAC-based agents, HyT-based degraders, and MG, offer innovative alternatives to PROTACs for TPD. Each technology employs distinct mechanisms and offers unique advantages for therapeutic applications. Unlike PROTACs, which utilize the proteasomal degradation pathway, AUTAC agents promote lysosome-dependent degradation of target proteins.⁸⁷⁷ One of the key benefits of AUTAC degraders is their enhanced membrane permeability due to their typically low molecular weights, making them potent therapeutic candidates.⁸⁷⁸ For instance, AUTAC-based degraders targeting BRD4, developed from the covalent interaction between autophagy key proteins and JQ-1, have shown significant antiproliferative activity across multiple tumor cell lines. This demonstrates their potential as effective medical tools for treating various diseases.⁸⁷⁹ Introduced in 2011, HyT technology uses small molecules composed of a targeted protein ligand, a hydrophobic tag, and a linker. Unlike PROTACs that often target the ubiquitin-proteasome system, HyT-based degraders work by increasing the hydrophobicity of the target protein, facilitating its degradation.⁸⁸⁰ MS1943, an EZH2 HyT degrader, illustrates this technology's effectiveness.⁸⁸¹ It has shown superior inhibitory effects on tumor cell lines and greater selectivity towards cancer cells over normal cells, demonstrating significant tumor suppression and good tolerance in mouse models.⁸⁸¹ Other HyT-based degraders are also developed, including those targeting HDAC and YEATS domain readers, providing therapeutic strategies for diseases caused by mutations or dysfunction in specific proteins.^{882–884} Further development of HyT-based degraders is ongoing, with efforts to improve their bioavailability and therapeutic effects in vivo by exploring new hydrophobic labels.⁸⁸⁵ MG differs fundamentally from PROTACs and other degraders by inducing degradation through promoting tight binding between the target protein and proteasome components, leading to the protein's subsequent degradation.^{886,887} A notable example is the MG-based degrader DD-1-073, targeting HDAC1/3, derived from SAHA (a known HDAC inhibitor). This was among the first applications of MG in HDAC degrader development.⁸⁸⁸ Similarly, another MG-based agent targeting BRD4, termed JP-2-197, is further established as an optimal derivative of JQ-1.⁸⁸⁸ Due to their low molecular weights, DD-1-073 and JP-2-197 have favorable pharmacokinetic properties, enhancing cell permeability and drug-ability.⁸⁸⁸ Despite their potential, the development of MG-based agents faces challenges due to the lack of systematic strategies for their design and identification, making large-scale screening and optimization difficult.

Developing epigenetic-targeting degraders, especially those targeting HDACs and epigenetic readers, has made significant progress, opening new avenues for clinical practice. These novel small molecules offer promising therapeutic alternatives, but several challenges and limitations must be addressed to enhance their clinical applicability and effectiveness.

Combining epigenetics-targeted drugs and sequencing technology

Owing to the substantial relationship among epigenetic signatures, lifestyle choices, and environmental influences, drugs that target epigenetic mechanisms are highly promising for advancing personalized medicine.⁸⁸⁹ However, leveraging these drugs in this field is challenging. As disease research enters a new phase owing

to advancements in sequencing technologies, the potential for epigenetic therapies to enhance personalized healthcare is being progressively realized.⁸⁹⁰

The advent of these cutting-edge technologies, ranging from whole-genome sequencing to single-cell analysis, has facilitated the detection of gene mutations and expression alterations associated with epigenetic changes throughout disease progression.⁸⁹¹ This advancement significantly enhances our understanding of the heterogeneity in epigenetic modifications across various cell types, thereby revealing new therapeutic targets for clinical application. In recent years, advancements in single-cell methodologies have allowed researchers to further explore the multiple dimensions of the epigenome, including chromatin accessibility, DNA methylation patterns, histone modification profiles, and chromatin interaction networks.^{892–894} Collectively known as “single-cell epigenomics”, this burgeoning field offers an enhanced comprehension of epigenomic regulation in physiological and pathological settings at the level of individual cells from a more intuitive perspective.^{895–897} These comprehensive “omic” profilings support the distinct biological identity of individuals, thus providing a solid theoretical foundation for refining therapeutic strategies to achieve individual targeting.⁸⁹⁸ Furthermore, by integrating CRISPR/cas9 gene-editing technology with various sequencing, researchers can conduct high-throughput functional genomic screens and identify pathological genes that are responsive to epigenetic therapies. This step not only aids in identifying novel targets for epigenetic agents but also promotes the assessment of therapeutic responses of target tissues or cells.^{899,900} For example, these methods may help detect the occurrence of drug resistance and optimize the efficacy of epigenetic interventions.⁹⁰¹

The significant potential of epigenetics in customizing personalized medicine has generated high enthusiasm, and advancements in this field have been consistently focused in medical research. Notably, the introduction of sequencing technologies has enabled us to investigate the correlation between epigenetic markers and disease pathology more comprehensively, thus facilitating the development of targeted therapeutic strategies. Nonetheless, the current epigenetic technologies used in the laboratory present several technical challenges that require refinement, such as the necessity for high-demand algorithms,⁹⁰² sufficient amounts of training data,⁹⁰³ limited genome coverage per cell,^{904,905} and uncertain reproducibility.⁹⁰⁶ These techniques require significant improvement before they can be effectively used in clinical practice.

CONCLUSIONS AND PERSPECTIVES

Since the term “epigenetics” was first introduced in 1942, there has been a significant focus on elucidating the mechanisms and pivotal roles of epigenetic modifications and their associated enzymes in human physiology and pathology. Drugs targeting epigenetic enzymes have shown promising potential for treating diseases, particularly cancers. This review comprehensively examines the major epigenetic mechanisms involved in the pathogenesis and progression of various diseases. It also highlights recent advances in epigenetics-targeted drugs, underscoring their potential in clinical settings. Additionally, we explore the integration of novel technologies in drug development and the synergistic value of these drugs in conjunction with other cancer therapies, pointing to the future direction of epigenetics-oriented therapeutic strategies. Despite these advancements, significant challenges remain. For instance, certain enzymes that regulate the epigenetic landscape still lack effective targeted drugs, and those identified through virtual screening require further in vivo and in vitro investigation to validate their efficacy and safety profiles. Addressing these gaps

will be crucial for integrating epigenetics-targeted drugs into clinical practice.

Advances in in-depth understanding of epigenetics have largely enhanced possibilities of curing disease. To date, the application of epigenetics-modifying drugs in preclinical and clinical setting has provided promise for the beginning of the era of epigenetic-orientated therapeutic strategies. Given the future needs in this field, great attention should be focused on exploring the heterogeneity of epigenetic hallmarks in different diseases to design and develop epigenetics-targeted drugs with high selectivity and improved targeting efficiency, based on the elucidation for the biological and pathological roles of epigenetics. These results are imperative for designing and developing agonists and inhibitors of epigenetic enzymes with enhanced selectivity and bioactivity. Furthermore, apart from the knowledge based on experimental research and preclinical studies, assessing the therapeutic potential of epigenetic drugs for specific diseases, particularly in advanced clinical trials, is also vital for advancing this field. Concurrently, emphasis should be placed on the clinical potential of integrating innovative drug discovery technologies into developing epigenetic-based drugs. Moreover, while capitalizing on unique strengths of epigenetics, efforts should be made to combine these novel agents with traditional therapeutic modalities, with a view to achieving synergic effects in treating disease, especially in the case of tumors with genomic complexity. In conclusion, we believe that deepened research in this field will catalyze innovation in treatment approaches for diseases involving epigenetic mechanisms, offering new hope to patients.

ACKNOWLEDGEMENTS

This work was supported by the National Natural Science Foundation grant of China (No. 82371647, 82071607, 82071651, 82403489); National Key Research and Development Program (2022YFC3600304, 2022YFC2704700); Science and Technology Department of Sichuan Province (23ZDYF2489); Cadre Health Care Committee of Sichuan Province, China (2023-1701); Chengdu Science and Technology Bureau (2021-YF05-02106-SN, 2017-GH02-00030-HZ); Outstanding Scientific Fund of Shengjing Hospital (No. 202003); Sichuan University 0-1 Innovation Research project (2023SCUH0019); The Frontiers Medical Center, Tianfu Jincheng Laboratory Foundation (T FJC2023010001); China Postdoctoral Science Foundation (No. 2023M733902, 2023M743909); PHD Research Initiated Fund Project in Liaoning Province (No. 2023-BSBA-331, 2023-BSBA-352); Science and Technology Plan of Liaoning Province (2022JH2/20200066); 345 Talent Project of Shengjing Hospital of China Medical University (No. M1344).

AUTHOR CONTRIBUTIONS

D.L., X.X., and Z.N. proposed the topic and main idea. W.D., X.Q., and Y.F. wrote the original manuscript and drew the figures. W.D., R.G., and P.B. were responsible for the revision of the manuscript. W.D., X.Q., Y.F., and S.L. were responsible for collecting data and making tables. W.D., T.L., Y.J., and S.W. were responsible for the literature search. D.L., X.X., and Z.N. commented on and revised the manuscript. All authors have read and approved the article.

ADDITIONAL INFORMATION

Competing interests: The authors declare no competing interests.

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