nature communications



Article

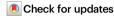
https://doi.org/10.1038/s41467-025-64125-3

Single-pixel infrared imaging thermometry maps human inner canthi temperature

Received: 16 September 2024

Accepted: 10 September 2025

Published online: 06 October 2025



Cheng Jiang^{1,2}, Patrick Kilcullen ^{1,2}, Yingming Lai¹, Tsuneyuki Ozaki¹ & Jinyang Liang ¹⊠

Efficiently and accurately mapping the temperature of human inner canthi is crucial for disease diagnostics and monitoring. The specific anatomical location of the inner canthi precludes temperature screening methods that are invasive, require tissue contact, and/or demand active illumination. Camerabased thermography, although capable of passive and non-contact temperature mapping, suffers from low efficiency in pixel allocation to the inner canthi as well as from measurement inaccuracies due to background blending and moderate pixel sensitivity. In response to these challenges, we develop singlepixel infrared imaging thermometry (SPIRIT). We design diagonally aggregated two-dimensional transmissive encoding masks using a cyclic S-matrix, which supports compressed data acquisition in a single scan and high image quality through non-iterative reconstruction. SPIRIT maps the temperature distribution of human inner canthi with a resolution of 0.3 °C, which enables human temperature mapping via single-pixel imaging. Using SPIRIT, we reveal sub-degree temperature differences induced by daily physical activities and the glasses-wearing habit. These findings shed light on SPIRIT's contribution to improving evaluation criteria for public health, including COVID-19 febrile screening.

Efficient and accurate determination of core body temperature is of paramount importance for diagnostics and monitoring of many diseases, such as COVID-191,2, sepsis3, neurological injuries4, and hyperthermia⁵. Nonetheless, the measurements at the wrist, forehead, and armpit may diverge from core body temperature due to external influences (e.g., airflow and coverings) as well as variations in blood flow, heat dissipation, and insulation⁶. By contrast, the inner canthi of the human eyes, shielded within the eye sockets, are situated in a more stable environment. Meanwhile, they have efficient and rapid heat exchange with blood via a prominent vascular network, which enables a sensitive response to core body temperature⁸. The thin, transparent mucous membranes of the inner canthi also facilitate an accurate temperature reading⁹. To determine the inner canthi temperature, international standards recommend spatially resolved measurements with at least 3×3 pixels, each being $1 \times 1 \text{ mm}^2$ in size¹⁰⁻¹².

Existing techniques confront challenges in accurately mapping the inner canthi temperature. The requirement for fast and efficient screening excludes all invasive methods that rely on target labeling (e.g., using rare-earth doped nanoparticles^{13,14} or thermochromic liquid crystals¹⁵) or target coupling (e.g., using ultrasound gel¹⁶). Moreover, for eye safety concerns, techniques necessitating active illumination, including laser thermometers¹⁷, photoacoustic imagers^{18,19}, and Raman spectrometers^{20,21}, are also inapplicable. Furthermore, to avoid clinical complications, techniques requiring direct tissue contact, such as thermocouples²² and thermistors²³, are disqualified.

Passive, non-contact techniques represent the most promising approach to mapping temperature distribution in the inner canthi. Among them, imaging thermography using a two-dimensional (2D) CCD or CMOS sensor, which can offer temperature information over a large spatial format, has become mainstream^{24,25}. Narrow-gap semiconductors, such as mercury cadmium telluride (HgCdTe), are

¹Centre Énergie Matériaux Télécommunications, Institut National de la Recherche Scientifique, Université du Québec, Varennes, QC, Canada. ²These authors contributed equally: Cheng Jiang, Patrick Kilcullen. ⊠e-mail: jinyang.liang@inrs.ca

particularly sensitive to long-wave infrared (LWIR) radiation²⁶ and hence are used as the photosensitive component in each sensor pixel to detect the thermal radiation emitted from humans²⁷. Though safe and easy to implement, this technique faces challenges in practicality and measurement accuracy. In particular, to minimize thermal noise, 2D LWIR sensors require cooling systems with efficient heat dissipation^{28,29}. However, commonly used technologies, including Stirling cryo-coolers and liquid coolers, introduce complexities in operation and considerably increase the cost³⁰. Moreover, vibration induced by the compressor assembly adversely affects the imaging quality and sensor durability³¹. In practice, LWIR thermography is typically configured to image the entire human face. The small size and spatial separation of the inner canthi result in a considerably low portion of allocated pixels (e.g., <0.5%) for temperature mapping^{12,32}. Consequently, the measurements are unavoidably affected by the background blending with the targeted areas. Considering the temperature-signal correlation, any perturbation in the signal inevitably compromises the accuracy of temperature readings³³.

Single-pixel imaging (SPI) offers a promising alternative to overcome the limitations of imaging thermography. This technique encodes the object's radiation by using pre-determined spatial patterns, with subsequent data acquisition performed by a point detector. During the ensuing image reconstruction, these encoding patterns serve as prior knowledge, enabling the accurate retrieval of spatial information^{34,35}. The throughput and multiplexing advantages embedded in SPI improve signal-to-noise ratios (SNRs) in measurements and reconstructed images, enabling it to detect fine intensity gradients and spatial details³⁶.

SPI leverages the advantages of single-pixel detectors with their wider availability in the sensing spectrum to fill the void where the 2D-sensor-based counterparts are not available or impractical³⁷, including infrared^{38,39}, terahertz⁴⁰, and acoustic⁴¹ bands. Moreover, compared to the complex heat-management systems required for 2D sensors, high-quality cooling of a single-pixel detector is more feasible, reliable, and economical, which enhances the accessibility and maintainability of the SPI systems. Furthermore, the masks used in SPI can be designed to image customized fields of view (FOVs). This resource-conserving paradigm allows for focusing on the regions of interest, which thus avoids capturing and processing irrelevant information.

Despite these attractive advantages, to date, SPI has not yet been demonstrated for human thermometry. A pivotal challenge lies in the ineffectiveness of available spatial light modulators (SLMs) in modulating LWIR light⁴²⁻⁴⁵. These pixelated devices, whose pixel size is comparable to the wavelength of LWIR light, bring in considerable loss in diffraction⁴⁶. Their protective windows are opaque to LWIR light⁴⁷. Efforts of re-windowing with zinc selenide (ZnSe) have enabled LWIR modulation using a digital micromirror device⁴⁸. However, window replacement can induce physical stress, contamination, and thermal mismatch, which can lead to misalignment, warping, or cracking of the delicate micromirrors⁴⁹. As an alternative, SLMs are capable of delivering patterned visible or near-infrared pulses to a semiconductor to induce transient absorption for LWIR light modulation⁵⁰. However, the required high-energy pump sources⁵¹ largely reduce its application scope. Besides the inherent limitations in SLMs, the coding strategy in most existing systems, which is based on the Walsh-Hadamard matrix⁵², exhibits drawbacks from time-consuming, compressed-sensing-based image reconstruction using a large number of iterations⁵³. Thus far, instant temperature mapping is still beyond reach.

To surmount these limitations, we develop single-pixel infrared imaging thermometry (SPIRIT) for temperature mapping in humans. This system passively detects thermal radiation using a transmissive encoding scheme. Diagonally aggregated physical masks constructed from a cyclic S-matrix are designed for 2D spatial encoding via a single compressed-sensing-compatible scan, ensuring that 100% of the

imaging pixels are dedicated to the targeted regions in the system's FOV. The encoded thermal radiation in the LWIR spectral range is then integrated by a cooled HgCdTe single-pixel detector. Leveraging spatial multiplexing, thermoelectric cooling, lock-in amplification, and non-iterative image processing, SPIRIT instantly maps the temperature distribution with a resolution of 0.3 °C. We apply SPIRIT to human inner canthi temperature mapping and monitor diurnal temperature variations. SPIRIT is also used to investigate the influence of gender and glasses wearing on the temperature readings at the inner canthi.

Results

System setup

A schematic of the SPIRIT system is depicted in Fig. 1. Thermal radiation emitted from the inner canthi is first imaged by a ZnSe lens (39-532, Edmund Optics) onto the intermediate image plane with a 4:1 magnification ratio. A mask plate and a window plate, each with a 2" diameter and a 150-um thickness, are placed at the intermediate image plane. The mask plate, attached to a motorized translation stage (MTS25-Z8, Thorlabs) moving vertically, contains two encoded stripes separated by 7.5 mm. Each encoded stripe comprises aggregated masks with encoding pixels of $250 \, \mu m \times 250 \, \mu m$ in size. The window plate has two open areas, measuring 2.75 mm × 1.75 mm and 2.75 mm × 1.50 mm, rotated to -45° and 45°, respectively, and separated by the same distance as the two encoded stripes (i.e., 7.5 mm). In this way, thermal radiation is encoded by two masks with sizes of 11×7 and 11 × 6 pixels, respectively (see the blue box in Fig. 1). Subsequently, the encoded thermal radiation in both areas is collected by another ZnSe lens (307B/1.0, Yoseen Infrared), passing through an optical chopper (MC2000B, Thorlabs), and finally focused onto an HgCdTe photodiode (PDAVJ10, Thorlabs) cooled by a Peltier thermoelectric cooler. SPIRIT's sensing spectrum, limited by the anti-reflection coating of the second ZnSe lens and the responsive range of the photodiode, is from 8 µm to 10.6 µm. As depicted in the yellow dashed box in Fig. 1, the FOV in this configuration contains two regions-11 mm × 7 mm (-45°-rotated) and 11 mm × 6 mm (+45°-rotated) in size—separated by 30 mm, which can cover both inner canthi of most humans⁵⁴. Meanwhile, each encoding pixel samples a 1mm × 1mm area in the targeted inner canthus, which satisfies the recommendation specified in the international standards^{10,11}. Details of the mask-window alignment are presented in Supplementary Note 1 and Supplementary Fig. 1.

In operation, the optical chopper provides an 850-Hz modulation. The acquired signals are sent to a lock-in amplifier (SR830, Stanford Research Systems) to enhance the SNR before being stored in a digitizer (ATS9625, Alazartech). Data acquisition is synchronized with the vertical scan of encoding masks. The resulting data points, termed "bucket signals", are then transferred to a computer for image reconstruction. Each scan, collecting 91 bucket signals (detailed in the next section), takes -15 s. A visible color camera (16-544-RCD-05P, Edmund Optics) with a camera lens (MVL8M23, Thorlabs) is also used to coincide the inner canthi with the FOV of the SPIRIT system. Details of system synchronization and FOV co-registration are provided in Supplementary Notes 2–3 and Supplementary Figs. 2–3. A performance comparison highlighting the impact of the lock-in amplifier on SNR enhancement is presented in Supplementary Note 4 and Supplementary Fig. 4.

Coding strategy

SPIRIT's encoding patterns are meticulously designed to efficiently measure temperature distribution from both inner canthi with a balance between data acquisition time and reconstructed image quality. The bucket signals, represented by an *m*-element vector *y*, can be interpreted as the inner products computed optically between an image comprising *n* pixels, represented by *x*, and the collection of 2D encoding patterns. This procedure can be succinctly articulated as

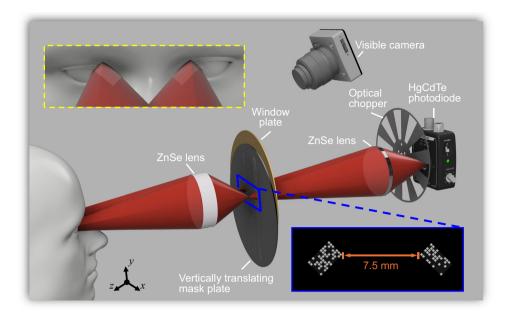


Fig. 1 | **Schematic of single-pixel infrared imaging thermometry (SPIRIT).** Yellow dashed box: Close-up view of the field of view. Blue box: Close-up view of an encoding mask passing the open areas on the window plate. The mask plate and the window plate, although made from the same material, are

depicted in different colors for display purposes. The CAD sketches of the optical chopper, photodiode, and visible camera lens in this figure are courtesy of Thorlabs, Inc. The 3D head model is reproduced with permission under a CC BY 4.0 license from Sketchfab.

y = Sx, where the measurement matrix S incorporates each encoding pattern represented in row form.

SPIRIT's encoding patterns are derived from a cyclic S-matrix (detailed in Methods, Supplementary Notes 5–6, Supplementary Fig. 5, Supplementary Table 1, Supplementary Movie 1, and Supplementary Software Package), which is known to offer superior performance over random-matrix encoding in terms of noise reduction and computation efficiency⁵⁵. As depicted in Fig. 2a, SPIRIT's measurement matrix S operationalizes a binary cyclic S-matrix with twin-prime factors of p=11, q=13, yielding $n=p\times q=143$. As an example, the first 2D encoding pattern, produced by row-major reshaping the first row of S, is shown in the blue shaded area of Fig. 2b. Then, several replicas of this pattern are tiled with shifts (shown in red, yellow, and green colors in Fig. 2b). This construction encapsulates the 2D encoding patterns reshaped from any row of S.

We develop a diagonal scanning approach for a compact aggregation of the encoding patterns compatible with compressed sensing in data acquisition and supporting non-iterative image reconstruction. As the first step, the tiled pattern is further duplicated and shifted (Supplementary Fig. 5d). Starting from the top-left corner, an aggregate pattern is generated by cropping out rectangles with $p \times q$ in size (delineated by the red dashed box in Supplementary Fig. 5d) along the diagonal direction across this matrix. As illustrated by the five colored boxes in Fig. 2b, this aggregate pattern, although corresponding to disconnected rows in the measurement matrix, supports continuous scanning (Supplementary Movie 1). More importantly, this approach supports a highly uniform sampling distribution of the set of 2Dshaped bucket signals. Figure 2c depicts the maximum distance of each unsampled signal to its sampled neighbors for m = 91 patterns. Compared to the conventional linear scan counterpart³⁷, SPIRIT substantially narrows the gaps between the unsampled signals. Considering the similarity between adjacent encoding patterns and thus their corresponding bucket signals, the unsampled data are estimated by using discrete Laplace interpolation⁵⁶, which enables forming an *n*-element vector \tilde{y} . Finally, by direct matrix inversion, the image is retrieved as $\tilde{x} = S^{-1}\tilde{y}$. This non-iterative compressed-sensing image reconstruction enables SPIRIT's online processing for instant data retrieval.

To implement the designed aggregate patterns for inner canthi, each encoding pattern is divided into two parts with the sizes of $p \times [(q+1)/2]$ and $p \times [(q-1)/2]$, respectively. Subsequently, each part undergoes flipping and/or rotation (detailed in Supplementary Note 6, Supplementary Fig. 5f, and Supplementary Movie 1), which better accommodates variations in the intercanthal distance among different individuals (Fig. 2d). By following the same arrangement procedure for each encoding pattern and exploiting the aggregation between consecutive encoding patterns, two encoded stripes are formed (Fig. 2e). A simulation of this coding strategy is presented in Supplementary Note 7 and Supplementary Fig. 6. Besides the m = 91 encoding patterns, four auxiliary L-shaped run-way patterns are appended at the beginning and the end of both encoded stripes to indicate the onset and completion of encoding as well as to facilitate the mask-window alignment. Finally, to ensure vertical alignment in encoding-mask scanning, one pixel is placed at the same horizontal position on the top and bottom of the mask plate (Fig. 2e).

Quantification of the SPIRIT system's performance

To experimentally demonstrate LWIR imaging using SPIRIT, we captured thermal radiation penetrating through a visible-light blocker. As depicted in Fig. 3a, temporally stable and spatially uniform thermal radiation was emitted from a blackbody radiator (YSHT-35, Yoseen Infrared) with a temperature of 37.0 °C. Within SPIRIT's FOV, the thermal radiation was shaped by two metal plates marked with the hollow letters "X" and "Z", respectively. Moreover, a piece of black plastic cover, opaque to visible light but partially transparent to LWIR light, was placed in front of the SPIRIT system. A pre-calibrated thermal camera (M384D, Yoseen Infrared) was implemented to provide the ground truth of the thermal signals. The experimental results are shown in Fig. 3b. While the visible camera in the SPIRIT system captured no information due to the cover's opacity in the visible spectrum, the LWIR imaging provided by SPIRIT correctly discerned both letters.

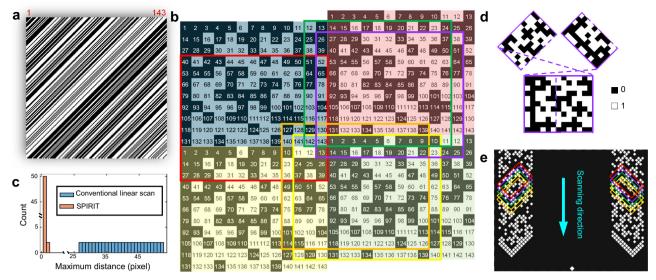


Fig. 2 | **Coding strategy of SPIRIT. a** Binary cyclic S-matrix of order n = 143 (i.e., p = 11 and q = 13). **b** Tiled matrix derived from (a). The numbers represent the corresponding row indices of each element of the first column of the matrix shown in (a), with shaded colors highlighting its tiling property. The colored boxes outline five consecutive encoding patterns corresponding to isolated rows in (a) indexed by their top-leftmost corner elements. **c** Histogram of the maximum neighboring

distance of the conventional linear scan and SPIRIT with the same scanning steps. **d** Separation and rearrangement of a representative encoding pattern marked by the purple box in (**b**). **e** Beginning section of the mask plate. The colored boxes outline the masks generated by the corresponding encoding patterns marked in (**b**).

To investigate the spatial resolution of SPIRIT, we imaged a constant thermal signal from the blackbody radiator through a 0.5-mm slit rotated to four angles— 0° , 90° , 45° , and -45° (Fig. 3c–f). Line profiles of normalized intensity along the direction of the slit width were then extracted for each rotation position. Spatial resolution was determined by counting the number of pixels along the line profile whose normalized intensity exceeded 0.5. This evaluation reveals that SPIRIT's spatial resolution is determined by the finite size of the encoding pixels. All four orientations exhibit the same spatial resolution of one SPIRIT pixel despite the anisotropy caused by the pixel shape. Further details on the number of data points are provided in Supplementary Table 2.

After validating the imaging capacity of SPIRIT, we performed temperature calibration using the blackbody radiator whose spatially uniform temperature was tuned from 31.7 °C to 39.8 °C with a step of 0.9 °C. For each preset temperature on the blackbody radiator, five measurements were conducted. The intensity in the reconstructed image is correlated with the temperature by 57

$$\widetilde{x} = C\varepsilon\sigma T_s^4 + \Delta. \tag{1}$$

Here, $T_{\rm s}$ is the preset temperature of the blackbody radiator, ε = 0.98 is the emissivity of the blackbody radiation source and of human⁵⁸. σ = 5.67 × 10⁻⁸ W m⁻² K⁻⁴ denotes the Stefan–Boltzmann constant. C takes into account the emission area and light-to-voltage conversion, and Δ is the measurement error, both of which were determined by a pixel-by-pixel linear fitting using Eq. (1). The measured temperature map, $T_{\rm m}$, was calculated as

$$T_{\rm m} = \sqrt[4]{\frac{\widetilde{x}}{C\varepsilon\sigma}}.$$
 (2)

The result is shown in Fig. 3g, which shows SPIRIT's temperature resolution of 0.3 $^{\circ}$ C.

Upon completion of temperature calibration, we used SPIRIT to reconstruct structured features and temperature distributions using Eq. (2). The blackbody radiator's temperature was set to 32.0 °C, 35.0 °C, and 38.0 °C, respectively. For each temperature, we placed

two metal plates with engraved letters in the FOV. The resulting reconstruction images, presented in Fig. 3h, i, display excellent agreement with the ground truth, demonstrating SPIRIT's ability to robustly map the temperature of fine features.

Temperature screening using SPIRIT

To demonstrate SPIRIT of the human inner canthi, we recruited 39 volunteers from diverse ethnic backgrounds (detailed information is included in Supplementary Table 3). The thermal camera was deployed as the gold standard to monitor the facial temperature of the volunteers during SPI. Figure 4a shows the representative temperature screening results for four volunteers. SPIRIT's results overlay the eye contour of the volunteers generated from the images captured by the co-registered visible camera. Using images captured by the visible camera, the inner canthi are identified as the areas formed by the inner (or nasal) junction of the upper and lower eyelids⁵⁹. SPIRIT's reconstructions reveal that the inner canthi regions exhibit higher temperatures relative to the adjacent areas covered by skin. The ground truth results are provided in Supplementary Note 8 and Supplementary Fig. 7, which show a good agreement with SPIRIT.

The mean temperatures within the left and right inner canthi were calculated by using only the pixels residing in these areas in the images reconstructed by SPIRIT (see the number of data points in Supplementary Table 4). Figure 4b shows that the 39 volunteers exhibit mean inner canthi temperatures ranging from 34.7 °C to 37.2 °C, aligning well with the ground truth. It is noteworthy that the mean temperatures of the two inner canthi can differ for the same individual, possibly due to asymmetrical blood flow, physiological stress, or emotional state³³. Moreover, considering the recommended threshold temperature for fever screening based on inner canthi to be 37.1 °C⁸, one volunteer (V16) was classified as fever-positive while the remaining 38 volunteers were fever-negative, all of which were consistent with the gold-standard measurements.

Monitoring of temperature variation related to physiology and habits using SPIRIT

We further investigated the temperature variation of the inner canthi related by three key factors among participants; diurnality, gender,

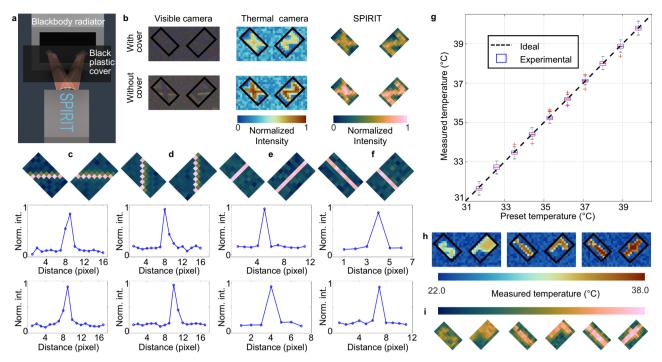


Fig. 3 | **Quantification of the SPIRIT system's performance. a** Experimental setup of the proof-of-concept demonstration. **b** Comparison of measurements using a visible camera (left), a thermal camera (middle), and SPIRIT (right), with (top row) and without (bottom row) a black plastic cover. Black boxes indicate the SPIRIT's field of view. SPIRIT reconstructions of a slit rotated to 0° (**c**), 90° (**d**), 45° (**e**), and -45° (**f**). Line profiles in the middle and bottom rows show the normalized mean intensity along the lines parallel to the slit in the left region and the right region of the field of view. The number of data points ($n_{\rm s}$) in each line is provided in Supplementary Table 2. **g** Relationship between the measured and the preset

temperatures. For each temperature, $n_{\rm s}=715$. In the box plot, the center line represents the median, the top and bottom of the boxes represent the 75th and 25th percentiles, respectively, and the whiskers indicate the minimum and maximum values within the interquartile range. The red crosses refer to outliers, which are values more than 1.5 times the interquartile range away from the bottom or top of the box. ${\bf h}, {\bf i}$ Ground truth of structured features with different temperatures (top row) and the corresponding reconstructions of SPIRIT (bottom row). Norm. int.: Normalized intensity.

and glasses wearing^{8,60,61}. For volunteers who regularly wear glasses, measurements were conducted immediately after the glasses were removed. To examine the impact of diurnal changes on temperature, a total of 29 volunteers (the remaining 10 volunteers provided only one measurement due to personal availability) were measured across four time windows (i.e., 9:00–11:00, 11:00–13:00, 13:00–15:00, and 15:00–17:00), as shown in Fig. 5a. Measurements from the fever-positive individual (i.e., V16) were excluded from the ensuing analyses. The 28 fever-negative participants had an equal representation in gender (i.e., 14 males and 14 females) and for each gender, seven wore glasses and seven did not.

Diurnal temperature variations were examined using SPIRIT measurements taken from individuals over a single day. As an example, Fig. 5b shows the imaging results of volunteer V13. The measured temperature maps across these time points reveal an undulating temperature increase of 0.9 °C with a fluctuation cycle of 4 h, characterized by a rise of 0.7 °C in the morning due to increased physical activity and hence a higher metabolic rate, a decline of 0.4 °C around noon as a result of energy dips associated with digestion and midday rest, and a subsequent rise of 0.6 °C in the afternoon by an increased level of activities. This trend, shared in the measured temperature variation in all 28 volunteers (Fig. 5a), reflects human physiology in response to the body's varying needs and activities.

We further analyzed temperature differences related to gender and glasses wearing across each measurement time window by dividing the participants into four groups. For volunteers who do not wear glasses, the measured temperatures for females were 35.5 ± 0.1 °C, 36.0 ± 0.1 °C, 35.6 ± 0.1 °C, and 36.0 ± 0.1 °C (mean \pm standard deviation) across the four measurement time windows, while the corresponding temperatures for males were 35.3 ± 0.1 °C, 35.8 ± 0.1 °C,

 $35.4\pm0.1\,^{\circ}\text{C}$, and $35.9\pm0.1\,^{\circ}\text{C}$, respectively. Similarly, for volunteers who wear glasses daily, the measured temperatures for females were $35.3\pm0.1\,^{\circ}\text{C}$, $35.7\pm0.1\,^{\circ}\text{C}$, $35.3\pm0.1\,^{\circ}\text{C}$, and $35.8\pm0.2\,^{\circ}\text{C}$, compared to $35.1\pm0.1\,^{\circ}\text{C}$, $35.6\pm0.1\,^{\circ}\text{C}$, $35.2\pm0.2\,^{\circ}\text{C}$, and $35.6\pm0.1\,^{\circ}\text{C}$ for males. As shown in Fig. 5c, under the same glasses-wearing condition, comparisons reveal a consistent mean temperature difference of $0.1-0.2\,^{\circ}\text{C}$ between male and female volunteers across the four time windows, which is likely attributed to metabolic rates and hormonal factors⁶³.

As for the glasses-wearing-related temperature differences, two examples are shown in Fig. 5d, and the gender-specific statistical analysis is presented in Fig. 5e. Both comparisons reveal that volunteers who do not wear glasses exhibit a 0.2-0.3 °C higher temperature than glasses-wearing volunteers. The observed discrepancy in inner canthi temperatures could be attributed to several factors. First, a frame of glasses can exert slight, continuous pressure on the skin and underlying tissues at the bridge of the nose and around the temples⁶⁴. Over time, this pressure could lead to minor reductions in blood flow to the area, slightly lowering the temperature of nearby regions including the inner canthi. Moreover, glasses can trap humidity in the ocular area, and as this moisture evaporates, it could cool the surrounding skin, affecting the inner canthi temperature⁶⁵. Finally, wearing glasses changes how people perceive and react to their environment, potentially altering typical eye movements⁶⁶. Reduced movement and the resulting decrease in muscular activity around the eyes might also have a minor effect on local temperature.

We performed statistical analyses of these measured results, shown in Fig. 5c and e. We first verified data normality with the Kolmogorov-Smirnov test. Next, a two-sample t-test was applied to establish significant differences (p < 0.05) between groups, followed by a right-tailed t-test to confirm that one group consistently exhibited

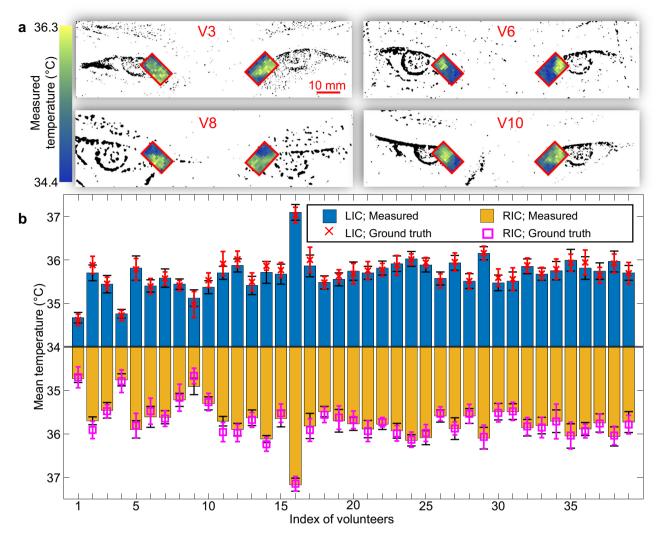


Fig. 4 | **Temperature mapping of human inner canthi by SPIRIT. a** Representative reconstruction results for four volunteers overlaying the corresponding eye contours. **b** Comparison of the mean temperature of pixels within the left inner canthus (LIC) and right inner canthus (RIC) of 39 volunteers, measured by

SPIRIT (shown as the bar heights) and the thermal camera as the ground truth (marked by the red crosses and the magenta squares). Error bars: standard deviation. The values of $n_{\rm s}$ vary for each volunteer and are provided in Supplementary Table 4.

higher temperatures. These analyses confirmed that the observed temperature differences related to diurnality, gender, and glasses wearing are statistically significant. Details of these statistical analyses are included in Supplementary Note 9, Supplementary Figs. 8–10, and Supplementary Tables 5–6.

Discussion

We have developed SPIRIT for efficient and accurate temperature mapping of human inner canthi. SPIRIT incorporates diagonal aggregation of 2D encoding patterns to uniformly fill the 2D-reshaped bucket signals, which allows non-iterative image reconstruction for instant data retrieval. SPIRIT has enabled passive temperature mapping of human inner canthi in 15 s with an overall frame size of 11×13 pixels and a temperature resolution of 0.3 °C. We have also applied SPIRIT to the monitoring of the diurnal temperature variations as well as an investigation of temperature differences related to gender and glasses wearing.

SPIRIT possesses several technical advantages. First, its transmissive masks with a relatively large encoding pixel size reduce diffraction loss and the number of required optical components, which enhances light throughput and hence the system's sensitivity compared to SLM-based SPI. SPIRIT also employs a single compressed

sensing-compatible scan of diagonally aggregated 2D masks generated from a cyclic S-matrix. This coding strategy not only accommodates variations in inner canthi structures and intercanthal distances but also facilitates lightweight, non-iterative image reconstruction. Moreover, in contrast to camera-based 2D thermography techniques, SPIRIT dedicates its entire FOV to the inner canthi, which enhances the measurement efficiency and reduces inaccuracies in temperature readings due to the background blending.

SPIRIT also holds practical merits. Its passive and non-contact operation is eye-safe and avoids sanitizing complications, making it suitable for large-scale temperature screening. Moreover, considering the international regulation and control of 2D thermal sensors, SPIRIT offers an open-source solution⁶⁷. Its ability to detect sub-degree temperature differences related to daily physical activities, gender, and the glasses-wearing habit sheds light on setting more precise evaluation criteria for public health (e.g., for COVID–19 febrile screening). Notably, SPIRIT unexpectedly revealed the onset and development of fever in one volunteer (V16), who started with the fever-negative condition (Fig. 5a). As another example, the glasses-wearing-related temperature difference provides experimental evidence for the under-explored impact of glasses-established microclimate between the inner canthi and the spectacle lenses⁶⁸. Its economic setup and instant data

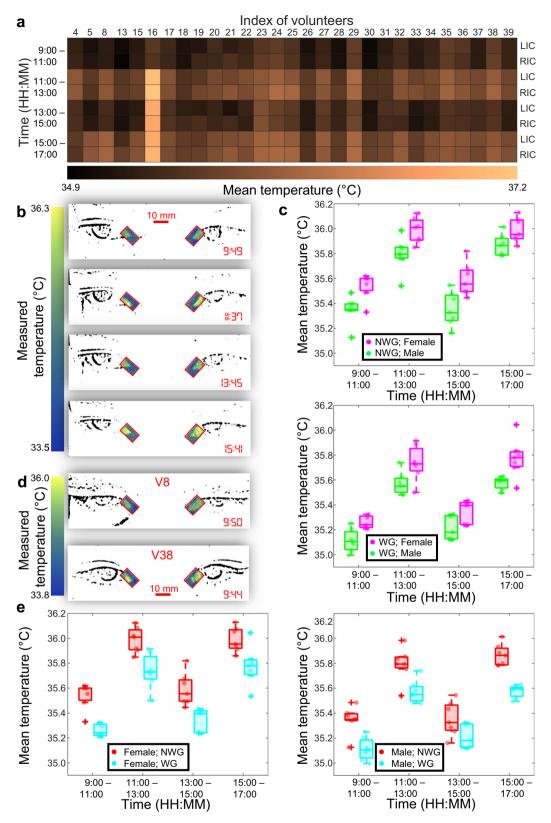


Fig. 5 | **SPIRIT of human temperature variation. a** Heatmap of inner-canthus temperature across four measurements for 29 volunteers on a single day. **b** Temperature maps of volunteer V13 at four time points on a single day. **c** Gender-related temperature differences among volunteers, subdivided into those not wearing glasses (NWG, top) and those wearing glasses (WG, bottom). For both plots

in (c), n_s = 7 for both female and male. **d** Representative results of volunteers with (top) and without (bottom) glasses. **e** Glasses-wearing-related temperature differences among volunteers, subdivided into females (left) and males (right). For both plots in (e), n_s = 7 for both WG and NWG. The box plots of (c) and (e) follow the same convention as that used in Fig. 3g.

processing further strengthen its implementation perspective in public places and clinics.

As a generic platform, SPIRIT holds promises for future development and applications. Currently, the SPIRIT system possesses two technical challenges. First, compared to infrared thermography, SPIRIT requires precise alignment of various components for data acquisition and relies on image reconstruction to retrieve temperature maps. Second, the 15-s data acquisition time requires volunteers to keep their eyes open for the entire duration, which sometimes caused eye blinking, leading to unsuccessful data acquisition. To overcome these limitations, several technical improvements are envisioned. First, an automated calibration module using fiducial markers and real-time motorized adjustments for mask-window alignment could simplify SPIRIT's component alignment^{69,70}. Second, a fully automated innercanthus detection algorithm could be developed to enhance the temperature measurement process⁷¹⁻⁷⁵. Third, a high-speed rotating mask could be employed to increase frame size and reduce data acquisition time⁷⁶⁻⁷⁸. Fourth, advanced interpolation methods could be explored to potentially improve reconstructed image quality without compromising processing time^{79,80}. Finally, a zoom ZnSe lens⁸¹ could be engineered to adjust the magnification ratio so that the intercanthal area projected onto the mask plate would maintain a constant length (i.e., 7.5 mm), enhancing SPIRIT's adaptability to a wide range of facial geometries. Besides temperature screening, SPIRIT is readily applicable to perceiving abnormal temperature signatures from diseases (e.g., inflammation⁸², circulatory disorders⁸³, and breast cancer⁸⁴) and detecting objects in obscured conditions (e.g., fog85, smoke86, and occlusion⁸⁷). Other potential applications also include infrared astronomy, atmospheric observations⁸⁸, and property characterization of advanced materials89.

Methods

Human study

The human study was conducted in strict adherence to the protocol (Project CÉR-20-578) approved by the Human Ethics Research Committee at the Institut National de la Recherche Scientifique, Université du Québec. In total, 39 volunteers were recruited for this study, including 19 females and 20 males, as determined by self-reported gender. Informed consent was obtained from every participant. Additional information in each experiment presented in this work is illustrated in Figs. 4–5 and stated in Main Text. Details of the protocol are provided in Supplementary Note 8.

Cyclic S-matrix

An S-matrix of order n can be defined as any matrix within the class of $\{0,1\}$ -valued $n \times n$ matrices that maximizes the determinant⁹⁰. It can be shown that for this S-matrix to exist, n must be either equal to 1, or of the form 4w-1 for some positive integer w^{55} . Moreover, it can be shown that for any S-matrix denoted by S^{90} ,

$$S^{-1} = \frac{2}{n+1} \left(2S^T - J \right), \tag{3}$$

where *J* denotes the all-ones matrix with size $n \times n$, and S^T denotes the matrix transpose of *S*.

Cyclic S-matrices possess an additional cyclic structure, whereby the initial row determines all subsequent rows via left-wise circular shifts. In particular, letting $S_{i,j}$ denote the element of S with row index i and column index j, $S_{i,j}$ satisfies

$$S_{i,i} = S_{0,i+i}$$
, (4)

where $\{i,j\} = 0, \ldots, n-1$, and i+j is interpreted modulo n. For imaging, it is desirable that n be factorizable into two parts of approximately equal size.

For the pattern design in SPIRIT, the "twin-prime construction method" was used^{37,55}. Given any pair of twin primes p and q = p + 2, it is possible to construct a cyclic S-matrix of order n = pq. The following functions can then be defined:

$$f(j) = \begin{cases} +1 & \text{if } j \text{ is a quadratic residue } (\text{mod } p) \\ 0 & \text{if } j \equiv 0 \, (\text{mod } p) \\ -1 & \text{otherwise} \end{cases}$$
 and (5)

$$g(j) = \begin{cases} +1 & \text{if } j \text{ is a quadratic residue } (\text{mod } p) \\ 0 & \text{if } j \equiv 0 \pmod{p} \\ -1 & \text{otherwise} \end{cases}$$
 (6)

from which the elements of S are computed, with i+j interpreted modulo n, by

$$S_{i,j} = \begin{cases} 0 & \text{if } [f(i+j) - g(i+j)]g(i+j) = 0\\ +1 & \text{otherwise} \end{cases}$$
 (7)

Data interpolation and image reconstruction

The image reconstruction procedure of SPIRIT leverages the fact that the multiplication of a vector by a cyclic S-matrix of appropriate order can be viewed as an operation of 2D discrete convolution^{37,56}. In particular, the following equivalence of indexed sums may be observed for expressing the matrix-vector product of an underlying image x with a cyclic S-matrix S:

$$y_u = \sum_{v=0}^{n-1} S_{u,v} x_v = \sum_{k=0}^{p-1} \sum_{l=0}^{q-1} S_0(i+k,j+l) x(k,l) = y(i,j).$$
 (8)

Here, the notations y(i,j), x(k,l), and $S_0(i+k,j+l)$ are used as respective shorthands for y_u , x_v , and $S_{0,u+v}$ (i.e., showing the conversion between 2D and 1D indexing schemes via row-major ordering), where u=qi+j, v=qk+l, and all resulting indices are interpreted as modulo n. The salient feature of Eq. (8) is that, since both horizontal and vertical index shifts of ± 1 result in a high degree of similar spatial overlap between encoding patterns and the underlying image, bucket signals collected from the SPIRIT system exhibit 2D smoothness when organized according to row-major ordering. In terms of linear indexing, this feature implies that for any individual bucket signal y_u , a high degree of correlation should be expected between values such as of $y_{u\pm 1}$ and $y_{u\pm q}$ that neighbor y_u after 2D reshaping.

This expected 2D smoothness gives rise to redundancy that is exploited to achieve a reduction in the size of physical encoding masks used in SPIRIT^{36,55}. In particular, the physical masks are designed in such a way that a continuous scan acquires the elements of y in a diagonal fashion within a 2D array, i.e., y(0,0), y(1,1), ..., and y(m-1,m-1), in a compressed data collection (i.e., with m < n). This design also allows the measured elements of y to be distributed in an approximately uniform manner (Supplementary Fig. 5e), well-suited for the estimation of non-measured elements via 2D interpolation. More details about the design procedure used to select the desired matrix order n and scan size m are provided in Supplementary Note 5.

Discrete Laplace interpolation⁹¹ was used to estimate the non-measured data points in the 2D-shaped bucket signals by

$$y_{u} = \frac{1}{4} \left[y_{u+1} + y_{u-1} + y_{u+q} + y_{u-q} \right].$$
 (9)

This approach allows a complete n-element interpolated data vector \tilde{y} to be constructed as a combination of all bucket signal values both measured and estimated. From the full interpolated data vector \tilde{y} , a final recovered image \tilde{x} is obtained via inversion of the full

measurement matrix. Finally, the vector \widetilde{x} may be reshaped according to both row-major ordering, as well as separation and rotation to produce images concurring with the two regions in the FOV defined by the windows on the window plate.

Reporting summary

Further information on research design is available in the Nature Portfolio Reporting Summary linked to this article.

Data availability

All data needed to evaluate the findings of this study are present in the paper and Supplementary Information. Raw data are provided with the Source Data Package (https://doi.org/10.5281/zenodo.16790723).

Code availability

A MATLAB software package is available as Supplementary Software Package (https://doi.org/10.5281/zenodo.16790730).

References

- Perpetuini, D., Filippini, C., Cardone, D. & Merla, A. An overview of thermal infrared imaging-based screenings during pandemic emergencies. Int. J. Environ. Res. Public Health 18, 3286 (2021).
- Eurosurveillance editorial team Updated rapid risk assessment from ECDC on the novel coronavirus disease 2019 (COVID-19) pandemic: increased transmission in the EU/EEA and the UK. Eurosurveillance 25, 2003121 (2020).
- Evans, S. S., Repasky, E. A. & Fisher, D. T. Fever and the thermal regulation of immunity: the immune system feels the heat. *Nat. Rev. Immunol.* 15, 335–349 (2015).
- Polderman, K. H. Induced hypothermia and fever control for prevention and treatment of neurological injuries. *Lancet* 371, 1955–1969 (2008).
- Nakamura, K., Nakamura, Y. & Kataoka, N. A hypothalamomedullary network for physiological responses to environmental stresses. Nat. Rev. Neurosci. 23, 35–52 (2022).
- Dushoff, J., Plotkin, J. B., Viboud, C., Earn, D. J. & Simonsen, L. Mortality due to influenza in the United States—an annualized regression approach using multiple-cause mortality data. Am. J. Epidemiol. 163, 181–187 (2006).
- Ring, E., McEvoy, H., Jung, A., Zuber, J. & Machin, G. New standards for devices used for the measurement of human body temperature. J. Med. Eng. Technol. 34, 249–253 (2010).
- Foster, J., Lloyd, A. B. & Havenith, G. Non-contact infrared assessment of human body temperature: the journal Temperature tool-box. *Temperature* 8, 306–319 (2021).
- Ghourabi, M., Mourad-Chehade, F. & Chkeir, A. Eye recognition by YOLO for inner canthus temperature detection in the elderly using a transfer learning approach. Sensors 23, 1851 (2023).
- Medical electrical equipment Part 2-56: Particular Requirements for Basic Safety and Essential Performance of Clinical Thermometers for Body Temperature Measurement. ISO 80601-2-56: 2017 (International Organization for Standardization, 2017).
- Medical electrical equipment—Part 2-59: Particular Requirements for the Basic Safety and Essential Performance of Screening Thermographs for Human Febrile Temperature Screening—Amendment
 IEC 80601-2-59:2017/Amd 1:2023 (International Electrotechnical Commission, 2023).
- Ring, E. & Ammer, K. Infrared thermal imaging in medicine. *Physiol. Meas.* 33, R33 (2012).
- Liu, X. et al. Fast wide-field upconversion luminescence lifetime thermometry enabled by single-shot compressed ultrahigh-speed imaging. Nat. Commun. 12, 6401 (2021).
- 14. Liu, M., Lai, Y., Marquez, M., Vetrone, F. & Liang, J. Short-wave Infrared photoluminescence lifetime mapping of rare-earth doped

- nanoparticles using all-optical streak imaging. Adv. Sci. 11, 2305284 (2024).
- Smith, C., Sabatino, D. & Praisner, T. Temperature sensing with thermochromic liquid crystals. Exp. Fluids 30, 190-201 (2001).
- Daw, J., Rempe, J. & Wilkins, S. Ultrasonic Thermometry for in-pile Temperature Detection (Idaho National Lab.(INL), Idaho Falls, 2002).
- Sollai, S. et al. Performance of a non-contact infrared thermometer in healthy newborns. BMJ Open 6, e008695 (2016).
- Pramanik, M. & Wang, L. V. Thermoacoustic and photoacoustic sensing of temperature. J. Biomed. Opt. 14, 054024–054027 (2009).
- Shah, J. et al. Photoacoustic imaging and temperature measurement for photothermal cancer therapy. J. Biomed. Opt. 13, 034024–034029 (2008).
- Ellis, D. I., Cowcher, D. P., Ashton, L., O'Hagan, S. & Goodacre, R. Illuminating disease and enlightening biomedicine: Raman spectroscopy as a diagnostic tool. *Analyst* 138, 3871–3884 (2013).
- Han, X. X., Rodriguez, R. S., Haynes, C. L., Ozaki, Y. & Zhao, B. Surface-enhanced Raman spectroscopy. *Nat. Rev. Methods Prim.* 1, 87 (2021).
- Rykaczewski, K. & Dhanote, T. Analysis of thermocouple-based finger contact temperature measurements. J. Therm. Biol. 108, 103293 (2022).
- 23. Hughes-Riley, T., Lugoda, P., Dias, T., Trabi, C. L. & Morris, R. H. A study of thermistor performance within a textile structure. *Sensors* **17**, 1804 (2017).
- 24. Lahiri, B. B., Bagavathiappan, S., Jayakumar, T. & Philip, J. Medical applications of infrared thermography: a review. *Infrared Phys. Technol.* **55**, 221–235 (2012).
- Meola, C. & Carlomagno, G. M. Recent advances in the use of infrared thermography. Meas. Sci. Technol. 15, R27 (2004).
- Hanna, S. et al. MCT-based LWIR and VLWIR 2D focal plane detector arrays for low dark current applications at AIM. *J. Electron. Mater.* 45, 4542–4551 (2016).
- Carlomagno, G. M. & Cardone, G. Infrared thermography for convective heat transfer measurements. *Exp. Fluids* 49, 1187–1218 (2010).
- Barela, J., Kastek, M., Firmanty, K. & Krupiński, M. Comparison of parameters of modern cooled and uncooled thermal cameras. SPIE Proc. 10433, 23–30 (2017).
- Dhumal, A. R., Kulkarni, A. P. & Ambhore, N. H. A comprehensive review on thermal management of electronic devices. *J. Eng. Appl. Sci.* 70, 140 (2023).
- Singh, M., Sadana, M., Sachdev, S. & Pratap, G. Development of miniature Stirling cryocooler technology for infrared focal plane array. *Def. Sci. J.* 63, 571 (2013).
- 31. Yang, B. et al. A novel vibration control system for Stirling cryocoolers. SPIE Proc. **6835**, 78–88 (2008).
- 32. Teunissen, L. P. J. & Daanen, H. A. M. Infrared thermal imaging of the inner canthus of the eye as an estimator of body core temperature. J. Med. Eng. Technol. 35, 134–138 (2011).
- Usamentiaga, R. et al. Infrared thermography for temperature measurement and non-destructive testing. Sensors 14, 12305–12348 (2014).
- 34. Sun, M.-J. et al. Single-pixel three-dimensional imaging with time-based depth resolution. *Nat. Commun.* **7**, 12010 (2016).
- 35. Gibson, G. M., Johnson, S. D. & Padgett, M. J. Single-pixel imaging 12 years on: a review. *Opt. Express* **28**, 28190–28208 (2020).
- Kilcullen, P., Jiang, C., Ozaki, T. & Liang, J. Camera-free threedimensional dual photography. Opt. Express 28, 29377–29389 (2020)
- Kilcullen, P., Ozaki, T. & Liang, J. Compressed ultrahigh-speed single-pixel imaging by swept aggregate patterns. *Nat. Commun.* 13, 7879 (2022).

- Tao, C. et al. Shortwave infrared single-pixel spectral imaging based on a GSST phase-change metasurface. Opt. Express 30, 33697–33707 (2022).
- Wang, Y. et al. Mid-infrared single-pixel imaging at the singlephoton level. Nat. Commun. 14, 1073 (2023).
- Stantchev, R. I., Yu, X., Blu, T. & Pickwell-MacPherson, E. Real-time terahertz imaging with a single-pixel detector. *Nat. Commun.* 11, 2535 (2020).
- 41. Liang, J., Gao, L., Li, C. & Wang, L. V. Spatially Fourier-encoded photoacoustic microscopy using a digital micromirror device. *Opt. Lett.* **39**, 430–433 (2014).
- Dudley, D., Duncan, W. M. & Slaughter, J. Emerging digital micromirror device (DMD) applications. SPIE Proc. 4985, 14–25 (2003).
- 43. Zhu, J. et al. Ultrabroadband spectral amplitude modulation using a liquid crystal spatial light modulator with ultraviolet-to-near-infrared bandwidth. *Appl. Opt.* **49**, 350–357 (2010).
- Xu, Q., Schmidt, B., Pradhan, S. & Lipson, M. Micrometre-scale silicon electro-optic modulator. *Nature* 435, 325–327 (2005).
- 45. Liu, F. et al. Acousto-optic modulation induced noises on heterodyne-interrogated interferometric fiber-optic sensors. *J. Lightwave Technol.* **36**, 3465–3471 (2018).
- Chen, D., Chen, B., Shao, Q. & Chen, S.-C. Broadband angular dispersion compensation for digital micromirror devices. Opt. Lett. 47, 457–460 (2022).
- Texas Instruments, DLP650LNIR 0.65 NIR WXGA S450 DMD (accessed on: 2025/02/17) https://www.ti.com/product/DLP650LNIR.
- Pan, Y., Xu, X. & Qiao, Y. Design of two-DMD based zoom MW and LW dual-band IRSP using pixel fusion. *Infrared Phys. Technol.* 91, 90–100 (2018).
- Gallego, G. B. et al. Front-window replacement and performance characterization of a commercial digital micro-mirror device for use in the infrared spectrum. SPIE Proc. 12777, 3179–3188 (2022).
- Dong, J., You, P., Tomasino, A., Yurtsever, A. & Morandotti, R. Singleshot ultrafast terahertz photography. *Nat. Commun.* 14, 1704 (2023).
- 51. Liu, J. et al. Swept coded aperture real-time femtophotography. *Nat. Commun.* **15**, 1589 (2024).
- Craig, S. M. et al. Compressive sensing hyperspectral imager in the LWIR for chemical plume detection. SPIE Proc. 12094, 186–199 (2022).
- Russell, T. A., McMackin, L., Bridge, B. & Baraniuk, R. Compressive hyperspectral sensor for LWIR gas detection. SPIE Proc. 8365, 55–67 (2012).
- Hayat, N., Alkhairy, S., Cheema, A., Ehsan, M. & Khan, M. A. Normal interpupillary, inner canthal distance and outer canthal distance in a normal population of Pakistan. *Pak. J. Med. Sci.* 35, 50–54 (2019).
- Sloane, N. J. & Harwit, M. Hadamard Transform Optics, Ch. 4 (Academic Press, 1979).
- 56. Gopalsami, N. et al. Passive millimeter-wave imaging with compressive sensing. *Opt. Eng.* **51**, 091614–091614 (2012).
- 57. Kittel, C. & Kroemer, H. Thermal Physics, Ch. 2 (Macmillan, 1980).
- 58. Sanchez-Marin, F. J., Calixto-Carrera, S. & Villaseñor-Mora, C. Novel approach to assess the emissivity of the human skin. *J. Biomed. Opt.* **14**, 024006-024006-024006 (2009).
- Remington, L. A., & Goodwin, D. Clinical Anatomy and Physiology of the Visual System E-book: Clinical Anatomy and Physiology of the Visual System E-book, Ch. 2, Page 10 (Elsevier Health Sciences, 2021).
- Yanovich, R., Ketko, I. & Charkoudian, N. Sex differences in human thermoregulation: relevance for 2020 and beyond. *Physiology* 35, 177–184 (2020).
- 61. Morris, C., Atkinson, G., Drust, B., Marrin, K. & Gregson, W. Human core temperature responses during exercise and subsequent

- recovery: an important interaction between diurnal variation and measurement site. *Chronobiol. Int.* **26**, 560–575 (2009).
- Atkinson, G. & Davenne, D. Relationships between sleep, physical activity and human health. *Physiol. Behav.* 90, 229–235 (2007).
- Vardasca, R. The influence of angles and distance on assessing inner-canthi of the eye skin temperature. *Thermol. Int.* 27, 130–135 (2017).
- 64. Jarosz, J., Molliex, N., Chenon, G. & Berge, B. Adaptive eyeglasses for presbyopia correction: an original variable-focus technology. *Opt. Express* **27**, 10533–10552 (2019).
- Korb, D. R. & Blackie, C. A. Using goggles to increase periocular humidity and reduce dry eye symptoms. Eye Contact Lens 39, 273–276 (2013).
- Müller, C., Stoll, W. & Schmäl, F. The effect of optical devices and repeated trials on the velocity of saccadic eye movements. *Acta Oto-Laryngol.* 123, 471–476 (2003).
- 67. Påhlsson, P. et al. Preliminary validation results of an ASIC for the readout and control of near-infrared large array detectors. *SPIE Proc.* **9451**, 672–686 (2015).
- Zhang, X. et al. Dry eye management: targeting the ocular surface microenvironment. *Int. J. Mol. Sci.* 18, 1398 (2017).
- Mondéjar-Guerra, V., Garrido-Jurado, S., Muñoz-Salinas, R., Marín-Jiménez, M. J. & Medina-Carnicer, R. Robust identification of fiducial markers in challenging conditions. *Expert Syst. Appl.* 93, 336–345 (2018).
- 70. Wang, S., Jiang, C., Peters, R., Aktary, M. & Liang, J. Double-illumination all-optical nanoalignment for stacking X-ray Fresnel zone plates. *Adv. Devices Instrum.* **6**, 0088 (2025).
- 71. Lazri, Z. M., Zhu, Q., Chen, M., Wu, M. & Wang, Q. Detecting essential landmarks directly in thermal images for remote body temperature and respiratory rate measurement with a two-phase system. *IEEE Access* **10**, 39080–39094 (2022).
- Jiang, C. et al. Real-time high-speed three-dimensional surface imaging using band-limited illumination profilometry with a CoaX-Press interface. Opt. Lett. 45, 964–967 (2020).
- 73. Jiang, C., Kilcullen, P., Lai, Y., Ozaki, T. & Liang, J. High-speed dual-view band-limited illumination profilometry using temporally interlaced acquisition. *Photonics Res.* **8**, 1808–1817 (2020).
- Jiang, C. et al. Multi-scale band-limited illumination profilometry for robust three-dimensional surface imaging at video rate. Opt. Express 30, 19824–19838 (2022).
- 75. Jiang, C. et al. Fringe Projection Profilometry. *Coded Optical Imaging*, 241–286 (Springer, 2024).
- Oreopoulos, J., Berman, R. & Browne, M. Spinning-disk confocal microscopy: present technology and future trends. *Methods Cell Biol.* 123, 153–175 (2014).
- 77. Hahamovich, E., Monin, S., Hazan, Y. & Rosenthal, A. Single pixel imaging at megahertz switching rates via cyclic Hadamard masks. *Nat. Commun.* **12**, 4516 (2021).
- 78. Diaz, N. et al. Single-mask sphere-packing with implicit neural representation reconstruction for ultrahigh-speed imaging. *Opt. Express* **33**, 24027–24038 (2025).
- 79. Chai, X. et al. Deep learning for regularly missing data reconstruction. *IEEE Trans. Geosci. Remote Sens.* **58**, 4406–4423 (2020).
- 80. Marquez, M. et al. Deep-learning supervised snapshot compressive imaging enabled by an end-to-end adaptive neural network. *IEEE J. Sel. Top. Signal Process.* **16**, 688–699 (2022).
- 81. Yan, A. Q. et al. Design of compact infrared zoom lens system. *Adv. Mater. Res.* **571**, 324–327 (2012).
- Halonen, J. I., Zanobetti, A., Sparrow, D., Vokonas, P. S. & Schwartz,
 J. Associations between outdoor temperature and markers of inflammation: a cohort study. *Environ. Health* 9, 1–9 (2010).
- 83. Cheshire, W. P. Jr Thermoregulatory disorders and illness related to heat and cold stress. *Auton. Neurosci.* **196**, 91–104 (2016).

- 84. Han, F., Shi, G., Liang, C., Wang, L. & Li, K. A simple and efficient method for breast cancer diagnosis based on infrared thermal imaging. *Cell Biochem. Biophys.* **71**, 491–498 (2015).
- Velázquez, J. M. R. et al. Analysis of thermal imaging performance under extreme foggy conditions: applications to autonomous driving. J. Imaging 8, 306 (2022).
- Tsai, P.-F., Liao, C.-H. & Yuan, S.-M. Using deep learning with thermal imaging for human detection in heavy smoke scenarios. Sensors 22, 5351 (2022).
- Komatsu, S., Markman, A., Mahalanobis, A., Chen, K. & Javidi, B. Three-dimensional integral imaging and object detection using long-wave infrared imaging. *Appl. Opt.* 56, D120–D126 (2017).
- Haworth, T. J., Glover, S. C., Koepferl, C. M., Bisbas, T. G. & Dale, J. E. Synthetic observations of star formation and the interstellar medium. New Astron. Rev. 82, 1–58 (2018).
- Kamble, V. B., Bhat, S. & Umarji, A. Investigating thermal stability of structural defects and its effect on d ferromagnetism in undoped SnO2. J. Appl. Phys. 113, 244307 (2013).
- 90. Sloane, N. J. & Harwit, M. Masks for Hadamard transform optics, and weighing designs. *Appl. Opt.* **15**, 107–114 (1976).
- 91. Hoeltgen L., Setzer S., Weickert J. An optimal control approach to find sparse data for Laplace interpolation. Proc. International Workshop on Energy Minimization Methods in Computer Vision and Pattern Recognition. 151–164 (Berline, 2013).

Acknowledgements

The authors thank Prof. My Ali El Khakani and Prof. Amar Mitiche for fruitful discussions and experimental assistance. The authors also thank all the volunteers for their participation and time. This work was supported in part by the Natural Sciences and Engineering Research Council of Canada (RGPIN-2024-05551, ALLRP 592389-23), the Canada Research Chairs Program (CRC-2022-00119), the INRS Chair in Nanobiophotonics, and the Fonds de Recherche du Québec-Nature et Technologies (203345-Centre d'Optique, Photonique, et Lasers) to J.L.

Author contributions

C.J. built the experimental setup, performed the experiments, and analyzed the data. P.K. developed the first generation of the reconstruction algorithm. Y.L. assisted in some experiments. J.L. and T.O. initiated the project. J.L. proposed the concept, contributed to experimental design, and supervised the project. C.J. and P.K. drafted the manuscript. All authors revised the manuscript.

Competing interests

J.L., C.J. and P.K. declare a competing interest related to a US Provisional Patent Application (No. 63/824,609). Y.L. and T.O. declare no competing interests.

Additional information

Supplementary information The online version contains supplementary material available at https://doi.org/10.1038/s41467-025-64125-3.

Correspondence and requests for materials should be addressed to Jinyang Liang.

Peer review information *Nature Communications* thanks Yuecheng Shen and the other anonymous reviewer(s) for their contribution to the peer review of this work. [A peer review file is available].

Reprints and permissions information is available at http://www.nature.com/reprints

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

© The Author(s) 2025