

<https://doi.org/10.1038/s42003-025-07813-6>

Convergent evolution of complex adaptive traits modulates angiogenesis in high-altitude Andean and Himalayan human populations



Giulia Ferraretti^{1,18}, Aina Rill^{1,14,18}, Paolo Abondio^{2,15,18}, Kyra Smith¹, Claudia Ojeda-Granados^{1,16}, Sara De Fanti³, Marta Alberti¹, Massimo Izzi⁴, Phurba T. Sherpa⁵, Paolo Cocco⁶, Massimiliano Tiriticco⁶, Marco Di Marcello⁶, Agnese Dezi⁷, Guido Alberto Gnecci-Ruscone^{8,17}, Luca Natali^{6,9}, Angela Corcelli¹⁰, Giorgio Marinelli⁶, Paolo Garagnani^{11,12}, Davide Peluzzi⁶, Donata Luiselli², Davide Pettener¹, Stefania Sarno^{1,19} & Marco Sazzini^{1,13,19} ✉

Convergent adaptations represent paradigmatic examples of the capacity of natural selection to influence organisms' biology. However, the possibility to investigate the genetic determinants underpinning convergent complex adaptive traits has been offered only recently by methods for inferring polygenic adaptations from genomic data. Relying on this approach, we demonstrate how high-altitude Andean human groups experienced pervasive selective events at angiogenic pathways, which resemble those previously attested for Himalayan populations despite partial convergence at the single-gene level was observed. This provides additional evidence for the drivers of convergent evolution of enhanced blood perfusion in populations exposed to hypobaric hypoxia for thousands of years.

Convergent evolution refers to an evolutionary scenario in which different species or populations independently develop the same (or similar) biological trait instead of inheriting it from a common ancestor. In particular, convergent adaptation occurs when distantly related species/populations, which live in distinct geographical areas or epochs, occupy comparable ecological settings and are subjected to the same selective pressures, thus being similarly targeted by natural selection¹. Several cases of convergent evolution have been described so far in a number of animal and plant taxa^{1–4},

with fewer examples (e.g., skin pigmentation⁵, malaria resistance⁶, lactose tolerance⁷ and metabolic adaptations to cold climates⁷) being instead reported in the human species when comparing adaptive traits among populations of different ancestry. Nevertheless, experimental approaches to accurately characterise genomic variability within and between species/populations, as well as inferential statistical methods suitable to investigate how far phenotypic convergence is achieved by means of genetic convergence, have been only recently conceived. This is especially the case of

¹Laboratory of Molecular Anthropology & Centre for Genome Biology, Department of Biological, Geological and Environmental Sciences, University of Bologna, Bologna, Italy. ²Department of Cultural Heritage, Ravenna Campus, University of Bologna, Ravenna, Italy. ³IRCCS Istituto delle Scienze Neurologiche di Bologna, Bologna, Italy. ⁴Complex Operative Unit of Endocrinology and Diabetes Care, Department of Medical and Surgical Sciences, University of Bologna, Bologna, Italy. ⁵Mount Everest Summitters Club, Rolwaling, Dolakha, Nepal. ⁶Explora Nunaat International, Montorio al Vomano, Teramo, Italy. ⁷Department of Precision and Regenerative Medicine and Ionian Area, University of Bari Aldo Moro, Bari, Italy. ⁸Department of Archaeogenetics, Max Planck Institute for Evolutionary Anthropology, Leipzig, Germany. ⁹Italian Institute of Human Paleontology, Rome, Italy. ¹⁰Department of Translational Biomedicine and Neuroscience, University of Bari Aldo Moro, Bari, Italy. ¹¹IRCCS Azienda Ospedaliero-Universitaria di Bologna, Bologna, Italy. ¹²Department of Medical and Surgical Sciences, University of Bologna, Bologna, Italy. ¹³Interdepartmental Centre Alma Mater Research Institute on Global Changes and Climate Change, University of Bologna, Bologna, Italy. ¹⁴Present address: Josep Carreras Leukaemia Research Institute, PhD Programme in Biomedicine, University of Barcelona, Barcelona, Spain. ¹⁵Present address: Department of Biology, University of Rome Tor Vergata, Rome, Italy. ¹⁶Present address: Department of Medical and Surgical Sciences and Advanced Technologies "GF Ingrassia", University of Catania, Catania, Italy. ¹⁷Present address: Archaeo- and Palaeogenetics, Institute for Archaeological Sciences, Department of Geosciences & Senckenberg Centre for Human Evolution and Palaeoenvironment, University of Tübingen, Tübingen, Germany. ¹⁸These authors contributed equally: Giulia Ferraretti, Aina Rill, Paolo Abondio. ¹⁹These authors jointly supervised this work: Stefania Sarno, Marco Sazzini. ✉e-mail: marco.sazzini2@unibo.it

complex (i.e., polygenic) adaptive traits, whose evolution is regulated by changes at several genes that are functionally related and contribute to the modulation of the same biological function⁸. Development of methods able to test the occurrence of selective events under a realistic approximation of a polygenic adaptation model has thus opened new possibilities to assess whether natural selection acted on the same genetic variants or genes or functional pathways in species/populations showing shared or similar adaptive traits⁹.

To this end, human adaptation to the high-altitude environment represents a valuable case study because the main selective pressure acting on populations dwelling at altitude (i.e., hypobaric hypoxia) cannot be mitigated by cultural adaptations and thus acts with the same intensity on human groups living at comparable altitudes irrespectively to their ancestry, geographical locations and socio-cultural contexts. We can thus hypothesise that Himalayan and Andean high-altitude populations, which represent the main human groups whose ancestors have had to cope with such a selective pressure, experienced convergent evolution at least to some extent. Despite this assumption, limited overlapping of potential adaptive loci (e.g., entailing variants at the *EGLN1*, *EDNRA* and *EPAS1* genes) has been described so far between them^{10–13}. This may be partially due to the limitations imposed by studies that to date focused mainly on selective sweeps. Indeed, these adaptive mechanisms are mediated by the action of natural selection on single/few genetic variants at the same gene, which implies that these loci can exert a large effect on a given biological trait⁸. That being so, selective sweeps have low probability to occur in human populations, who have long maintained low effective population sizes and have accumulated low genetic variability and thus need several thousands of years to evolve¹⁴. Coupled with the considerably more recent human colonisation of the high-altitude ecological niche in the Andes than in the Himalayas, as supported by estimates at around 9 thousand years ago (kya) for the divergence between Andean and lowland South American populations¹⁵ and by archaeological evidence for occupation of the Tibetan Plateau since Palaeolithic times¹⁶, this suggests selective sweeps might be not the predominant drivers of adaptation of Andean people to such a challenging environment.

Consistent with this scenario, Andeans indeed evolved less effective genetically regulated physiological adjustments than populations of Tibetan/Sherpa ancestry, especially to counteract long-term detrimental effects of hypobaric hypoxia^{17,18}. When compared with acclimatized Native American lowlanders, they exhibit traits contributing to enhanced efficiency of respiratory functions and oxygen utilisation, such as larger lung volumes and chest dimensions, narrower alveolar to arterial oxygen gradients, less hypoxia-induced pulmonary vasoconstriction, greater uterine artery blood flow during pregnancy, less altitude-related decrement in maximal exercise oxygen consumption and increased cardiac oxygen utilisation^{17,18}. However, when genetic determinants of these physiological characteristics have been searched for, most of the inferred selective sweeps failed to be replicated by multiple studies, suggesting that their relationships with the observed adaptive traits are far from being fully elucidated^{15,18–21}.

Results and discussion

To test whether polygenic adaptations played a role in shaping the Andean adaptive phenotype, and to extend to complex traits the investigation of the genetic bases of potential convergent evolution between Andean and Himalayan populations, we assembled a dataset representative of genomic variation at several high-altitude ethnic groups from the Andean region (Supplementary Table 1). After stringent quality control (QC) filtering (see ‘Methods’), 7,966,198 Single Nucleotide Variants (SNVs) from 24 Bolivian Aymara whole genome sequences (WGS)¹⁵ were used to perform multiple selection scans (i.e., H12, nSL and LASSI) that collectively are able to test for the occurrence of a range of selective events (see ‘Methods’ for further details concerning the rationale behind the choice of these inferential approaches). The obtained results then informed gene-network analyses aimed at identifying sets of functionally related loci enriched for signatures of natural selection. Furthermore, we applied the same pipeline of analyses on: (i)

imputed genotype data for 8,024,216 SNVs from 130 individuals belonging to additional Bolivian Aymara groups, as well as from Aymara, Quechua, and Uros high-altitude Peruvian populations^{22,23} used as validation datasets and (ii) a control dataset including 24 individuals from three different Amazonian low-altitude groups (i.e., Ashaninka, Cashibo, and Shipibo)²³ to filter out adaptive loci shared between high- and low-altitude South American populations, with the aim of focusing exclusively on those typical of Andean peoples.

Identification of Native American low-altitude control populations

Setting the considered Bolivian Aymara WGS into the genomic landscape of indigenous groups from Central and South America enabled us to confirm them as well representative of the genetic variation observable across Andean high-altitude populations (Fig. 1a, Supplementary Information). Comparable results were obtained also when considering imputed data for Andeans, thus providing evidence for consistency between non-imputed and imputed datasets (Supplementary Fig. 1). Such a consistency was further attested by the high and significant correlation observed between pre- and post-imputation identity-by-state (IBS) matrices of individual pairwise differences in the genome-wide proportion of shared alleles (Mantel correlation = 0.95, $P < 10^{-4}$).

The outgroup-f3 statistic was then computed on the assembled reference dataset to select the most suitable low-altitude control populations of South American ancestry to be contrasted with Andean groups in terms of evolved biological adaptations (see ‘Methods’ for a detailed description of the adopted selection criteria). According to this approach, while each Andean population was confirmed to share the greatest genetic similarity with the other Andean groups, Amazonian populations from Peru appeared as the next most affine ones to Andeans (Fig. 1b, Supplementary Fig. 2 and Supplementary Information). Among these putative control populations, eight Ashaninka, six Cashibo and 10 Shipibo subjects were specifically selected to constitute the low-altitude control group according to their clustering patterns and negligible Andean-specific genetic component, as pointed out by CHROMOPAINTER/fineSTRUCTURE and ADMIXTURE estimates respectively of haplotypes sharing and individual ancestry proportions (Fig. 1c and Supplementary Information).

Genomic signatures ascribable to polygenic adaptive events in Andean populations

At first, genome-wide distributions of nSL and H12 statistics, which are informative of both strong and weak selective events, were calculated for Andean and control groups relying on both WGS and imputed data. Then, we used these distributions as input for the *signet* algorithm to infer gene networks made up of loci belonging to the same functional pathway and that have been concurrently targeted by natural selection (see ‘Methods’). By considering only pathways confirmed by each selection statistic and by both WGS and imputed datasets, we shortlisted the most plausible biological functions having evolved adaptively in the considered populations.

Significant gene-networks identified for the low-altitude control group were found to participate primarily to immune-related pathways (e.g., *Staphylococcus aureus* infection, *Human papillomavirus* infection and *Arachidonic acid* metabolism), suggesting the evolution of adaptations able to mediate immune and inflammatory responses to pathogens (Supplementary Table 2). Similar selective pressures seem to have acted also on Andean groups, as attested by their putative adaptive loci belonging to the *Purine metabolism* and *Herpes simplex virus 1* infection pathways (Fig. 2a and Supplementary Table 3).

Conversely, adaptive events plausibly contributing to the response to stresses imposed by the high-altitude environment were inferred only in Andeans and were mediated by genes involved in the *Focal adhesion* pathway, which regulates cells adhesion to extracellular matrix and subsequent signal transduction at the intracellular level (Fig. 2a, b and Supplementary Table 3). Most of the loci constituting the significant networks identified by analysing both Aymara WGS and Andean imputed datasets

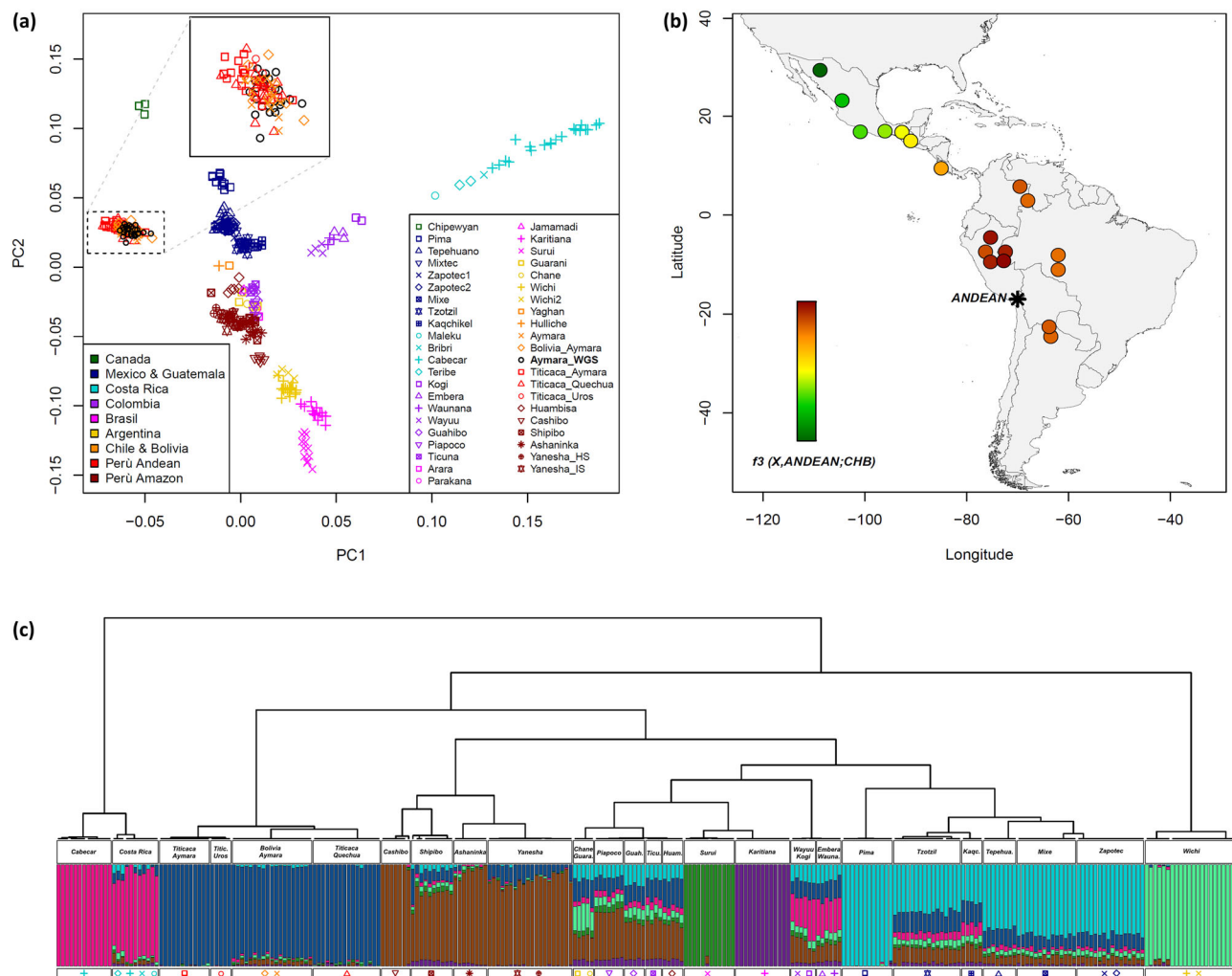


Fig. 1 | Population structure analyses performed to investigate genetic relationships between high-altitude Andean groups and low-altitude Central and South American populations. **a** PCA representing PC1 versus PC2 computed for the 43 un-admixed Native American groups reported in the legend at the bottom right of the plot. Individuals are colour-coded according to their country of origin as reported in the bottom-left legend. The box at the top of the plot details the position of Bolivian Aymara WGS in the considered genetic space with respect to the other high-altitude Andean populations. **b** Heat map of values for the outgroup- f_3 statistic representing the estimated amount of shared genetic drift between the Andean population cluster (marked by a star-like symbol) and each of the considered populations from Central and Southern America (indicated in their approximate

geographic locations) A gradient ranging from green to red is specifically used to indicate lower-to-higher levels of genetic affinity as reported by the corresponding colour scale. **c** Fine-scale patterns of genetic clustering and proportions of ancestry components inferred at the individual level. The fineSTRUCTURE hierarchical clustering dendrogram obtained for Andean, Central and South American individuals included in the assembled dataset is reported, along with ancestry components inferred for each subject with ADMIXTURE analysis at $K = 8$. For each genetic cluster pinpointed by the fineSTRUCTURE analysis, the corresponding individuals and their admixture proportions are specifically detailed, along with the symbols used in the PCA plot for the populations they belong to. The source data used to generate the figures are reported in Supplementary Data 2—Tables 1–3.

are indeed known to play a pivotal role in angiogenic processes (Fig. 2a, b). Especially five of these genes (i.e., *LAMA3*, *COL4A1*, *ITGB6*, *PRKCB* and *PTK2*) represent the most robust candidate targets of positive selection due to patterns ascribable to polygenic adaptive evolution identified according to both nSL-based and H12-based network analyses (Fig. 2b). Among them, *ITGB6*, *PRKCB* and *PTK2* have been proved to be the main regulators of vascular sprouting, which implies the transformation of blood vessels endothelial cells into cells able to migrate and proliferate out of the vessel to create new vascular structures^{24–31}. In fact, *PTK2* inactivation has been demonstrated to prevent the beginning of the angiogenic process, leading to a lack of development of vascular tissue in embryos and thus to their premature death^{32,33}. Although overall adaptive evolution of loci at the *Focal adhesion* pathway was highlighted by significant networks obtained according to both nSL and H12 statistics, the remaining 28 genes included in these networks were pointed out as potentially adaptive by a single statistic. Nevertheless, their functional

relationships with the highly confirmed loci described above have been well established and they are known to contribute to the regulation of blood vessels formation as well (Fig. 2a, b). For instance, multiple studies collected evidence supporting the interaction among *PTK2*, *VEGFA*, *PDGFA* and *PDGFD* as a key driver of blood vessels formation in placenta and some variants at these loci are found to be associated with reduced functionality of maternal-foetal circulation, ischaemic placental disease and impaired foetal growth^{34,35}.

To further validate these findings, we run the *signet* algorithm also on genome-wide distributions of values obtained by applying the LASSI method respectively on the WGS, imputed and low-altitude control datasets. This inferential approach was indeed proven to outperform H12 and nSL statistics in the detection of weak selective events, being also able to accurately distinguish them from hard selective sweeps³⁶ (see 'Methods' for further details). After having filtered out putative adaptive gene networks shared among high-altitude and low-altitude groups, LASSI-based network

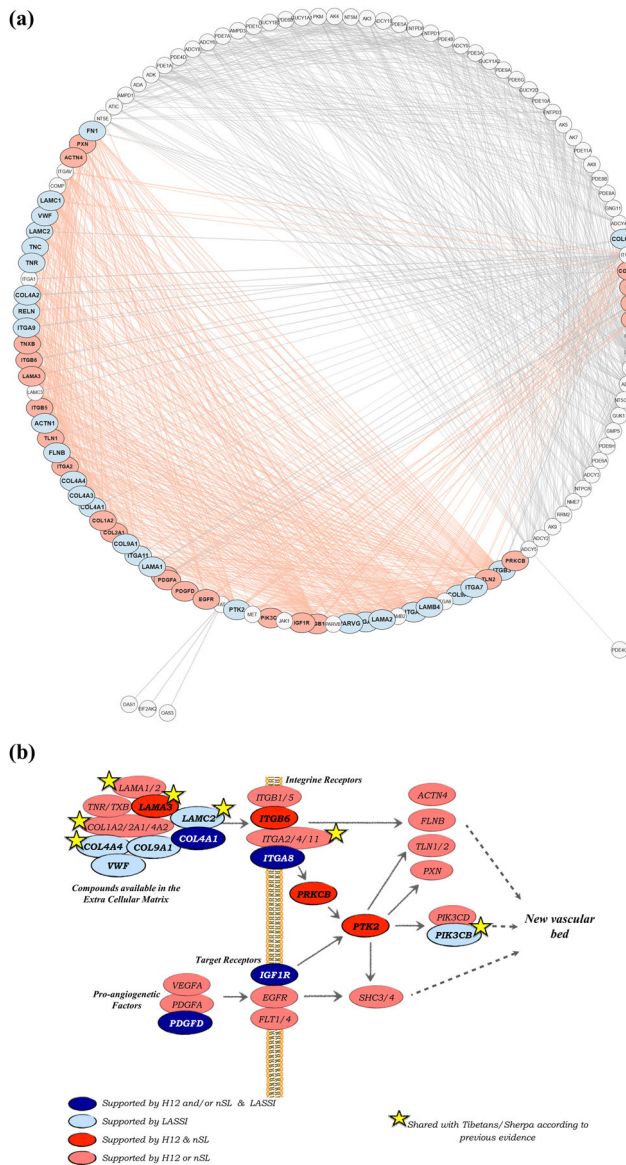


Fig. 2 | Functional relationships between genes putatively mediating polygenic adaptations evolved by Andean populations and their role in angiogenic processes. **a** Circular network build with Cytoscape v3.10.3 and displaying protein-protein functional interactions inferred for the entire set of adaptive genes belonging to significant pathways identified by H12-based and nSL-based *signet* analyses according to both WGS and imputed datasets. Genes pointed out by LASSI-based *signet* analyses as participating to the *Focal Adhesion/PI3K-Akt signalling* pathways are also reported. Gene products that establish similar connections in the network are placed next to each other, comporting the delineation of two well-distinguishable functional clusters. In detail, non-angiogenesis related genes (e.g., belonging to the *Herpes simplex virus 1 infection* pathway) are reported as light-grey circles, while angiogenesis-related genes (i.e., belonging to the *Focal adhesion/PI3K-Akt signalling* pathways) are displayed with pink (when supported by H12 and/or nSL) and light blue (when supported by LASSI) ellipses. The source data used to generate the plot is reported in Supplementary Data 2—Table 4. **b** Scheme displaying functional interactions between *PI3K-Akt signalling* and *Focal adhesion* adaptive genes contributing to improved angiogenesis in Andean populations according to the KEGG database. During the early phases of angiogenesis molecular compounds made available in the extracellular matrix, such as those building the basement vascular membrane (e.g., *COL4A1*) and angiogenic factors (e.g., *VEGFA* and *PDGFD*), interact with integrin and target receptors (e.g., *IGF1R*) stimulating protein kinases (e.g., *PRKCB* and *PTK2*), which subsequently activate those signalling cascades essential for migration of endothelial cells and formation of new vascular structures. The yellow stars mark those genes previously identified as loci putatively mediating polygenic adaptation to hypobaric hypoxia in Tibetan/Sherpa populations^{52,61,62}, thus providing evidence for partial genetic convergence between Andean and Himalayan adaptive traits in addition to the remarkable convergence observed at the biological function/pathway level.

analyses pointed out 39 genes plausibly targeted by positive selection and supported by results for both WGS and imputed datasets (Supplementary Fig. 3). Among them, 27 loci were found to belong to functional pathways related to angiogenesis promotion, cell proliferation and vasodilatation (Supplementary Fig. 3), with four of them (i.e., *COL4A1*, *IGF1R*, *PDGFD*, *ITGA8*) being included in the previously described significant networks belonging to the *Focal adhesion* pathway pointed out by H12-based and nSL-based analyses (Fig. 2b). Another significant *Focal adhesion* network was supported only by the Aymara WGS dataset (Fig. 2b and Supplementary Table 4), with some genes (i.e., *COL4A1*, *COL4A2*, *COL9A1*, *LAMC2*, *VWF* and *ITGA8*) participating also to a network belonging to a functionally related pathway (i.e., *PI3K-Akt signalling*) detected according to imputed data (Supplementary Fig. 3 and Supplementary Table 4). These results contribute to corroborate the identification of the *Focal adhesion pathway* and of associated molecular cascades as complex biological functions that have been shaped by the action of natural selection in Andean highlanders.

Among the adaptive genes validated by such an approach, *COL4A1* encodes a type IV collagen alpha protein that is one of the main constituents of the vascular basement membrane^{29,37}, whose degradation and remodeling represent the very first steps of angiogenesis induced by *VEGF*, *FGF* and *PDGF* growth factors³⁷. Interestingly, mutant *COL4A1* mice present retinal

and subretinal neovascular lesions with respect to wild-type ones³⁸, while human *COL4A1* and *COL4A2* variants have been associated with cerebrovascular disease²⁹ and intracerebral haemorrhages due to systemic small-vessel disease³⁹, suggesting that *COL4A* proteins play a crucial role in modulating the formation of new vascular structures. The *IGF1R* gene instead encodes for the insulin like growth factor 1 receptor that binds *IGF1* angiogenic factors and is known to be overexpressed in most malignant tissues where it acts as an anti-apoptotic agent by enhancing cell survival^{29,40}. In particular, remodelling of tumour blood vessels mediated by *IGF1* was observed in the melanoma mouse model⁴⁰, with *IGF1* and *IGF1R* deficiency being also associated to severe postnatal growth retardation in mice^{41–43}. Similarly, homozygous partial *IGF1* deletion in humans causes intrauterine growth retardation, which is accompanied by low average placental weight⁴⁴. Finally, the platelet-derived growth factor *PDGF-D* represents another angiogenic factor whose activity promotes tumour vasculogenesis in mice due to *VEGF* upregulation⁴⁵. Moreover, *PDGFD* down-regulation was also observed in the placenta of women with preeclampsia, a syndrome characterised by high resistance utero-placental circulation, placental hypoperfusion and elevated expression of antiangiogenic factors⁴⁶.

In line with this evidence, two additional significant gene networks participating to the *Apelin signalling* and *Vascular smooth muscle contraction* pathways were proposed to have adaptively evolved in Andeans, but not in the low-altitude control group, according to both WGS and imputed data. However, these signatures did not reach statistical significance in H12/nSL-based network analyses, being detected only by LASSI-based ones and including the *GNG7*, *PRKCE*, *RYR2* and *RYR3* loci (Supplementary Table 4 and Supplementary Fig. 4). In particular, *RYR2* and *RYR3* codify for two ryanodine receptors (RyR): the former is expressed in the sarcoplasmic reticulum of the cardiac muscle²⁹ and is essential for contraction regulation mediated by calcium release^{47,48}, while the latter is mainly found in skeletal muscle, as well as in the heart Purkinje tissue, thus possibly contributing to electric stability of these fibres in such an organ⁴⁹. When compared with acclimatised lowlanders, native Andean individuals are found to maintain heart rate unaltered during apnoea⁵⁰, similarly to what observed for Himalayan people⁵¹ and suggesting that adaptive modulation of heart frequency and cardiac output might result in a decreased risk of developing arrhythmias in these high-altitude populations.

Convergent adaptive evolution between Andean and Himalayan populations

Interestingly, when we previously applied a gene network-based approach to WGS from Tibetan and Sherpa populations, several genes included in Andean significant networks obtained from H12/nSL analyses (e.g., *LAMA1*, *COL4A2* and *ITGA2*) were similarly found to participate to functional pathways targeted by natural selection (Fig. 2b, Supplementary Table 5)⁵². This pattern was basically confirmed by results from LASSI-based analyses. First, this approach recapitulated previous evidence supporting convergent adaptations between Andean and Tibetan highlanders involving the *EPAS1*¹⁰ and *EGLN1*¹³ genes, with genomic windows at these loci being classified as targets of selective sweeps and showing likelihood values falling in the top 1% of the distribution obtained for the Aymara WGS dataset.

As regards instead genes included in significant networks, *LAMC2* was already proposed to have contributed to polygenic adaptive events also in Tibetan/Sherpa populations⁵² (Fig. 2b, Supplementary Table 5). This gene codifies for the laminin gamma 2 subunit, which is an epithelium-specific basement membrane protein highly expressed in different types of cancers and tumour metastasis^{53–55}. Silencing *LAMC2* expression was proved to cause cell cycle arrest and significantly suppresses migration and invasion of tumour cells, as well as in vivo angiogenesis of the malignant tissue due to partially blocked activation of the *EGF* receptor and its downstream pathway^{54,55}. Moreover, *LAMC2* expression in the placenta predominantly interests syncytiotrophoblasts and cytotrophoblasts villous and was found to be substantially upregulated in Placental Accreta Spectrum (PAS) tissues, stimulating trophoblast over-invasion via *PI3K/Akt/MMP2/9* signalling pathways⁵⁶. In fact, PAS is a condition causing pathologic adherence of the placenta due to abnormal trophoblast neovascularization and invasion into the uterine wall^{57,58}. This invasion has been related to improved stimulation of pro-angiogenic factors not only in the trophoblast embryonic tissue, but also in the maternal basal plate of the placenta^{58,59}. We can thus hypothesise that adaptive evolution at *LAMC2* and functionally related loci observed in the considered high-altitude populations might result in promoting angiogenesis in the placental tissues.

Other genes that we propose to have been targeted by natural selection in Andeans (i.e., *COL4A4*, *PLCE1*, *PIK3CB* and *PRKCE*) were previously described as plausible adaptive loci in Tibetan groups^{13,60–62} (Fig. 2b and Supplementary Table 5). Again some of them are known to be associated to inhibition of apoptosis and angiogenesis in tumours (*PLCE1*)⁶³ or to cardiovascular traits (*PRKCE*)⁶⁴. In particular, *PRKCE* has been demonstrated to exert a cardio-protective role against ischaemic injury⁶⁴ and presents variation patterns ascribable to both adaptive evolution and archaic introgression in Tibetan populations^{12,60,65}. Other genes instead seem to be involved in the modulation of decidual or in the decidualization phase in preparation to pregnancy^{66,67}. For instance, *COL4A4* expression, along with that of other proteins belonging to the collagen type IV family, increases during decidualization of the human endometrium⁶⁷ participating to the regulation of those morphological and functional changes (e.g., increased vascular permeability, oedema, invasion of leucocytes, vascular remodelling and angiogenesis) that are essential for the establishment of a successful pregnancy^{68,69}. Similarly, *PIK3CB* turned out to be up-regulated in the decidua of early-onset pre-eclampsia versus normal pregnancies, plausibly playing a role in abnormal placental development⁶⁶.

According to this picture, the obtained results revealed that changes at functional pathways enabling generalised and/or placental improvement of angiogenesis, as well as cardiovascular protection, seem to have characterised the evolutionary history of both Andean and Himalayan populations^{51,52,65}. Nevertheless, an incomplete genetic convergence between these high-altitude peoples was observed. This is not unexpected within the framework of an evolutionary scenario largely characterised by polygenic adaptive events, in which single loci play a negligible role with respect to their overall synergic interactions. Overall, we provided evidence for the same biological functions having adaptively evolved in these populations, plausibly in response to the selective pressure imposed by hypobaric hypoxia.

Putative adaptive genes associated with complex traits and eQTLs

To finally test whether the identified candidate adaptive genes are associated to complex physiological and/or pathological traits possibly implicated in the adaptive responses evolved by the considered high-altitude populations, we first consulted the Common Metabolic Disease Knowledge Portal (CMDKP) (Supplementary Table 6) (see ‘Methods’ for further details). Furthermore, we searched for expression quantitative trait loci (eQTLs) that could be able to modulate the expression of these genes in specific tissues. For this purpose, we relied on information stored in the GTEx portal and we verified if eQTLs at each gene are located in genomic windows showing large positive values of the likelihood statistic calculated by LASSI (i.e., those chromosomal regions representing the most robust targets of natural selection). The proportions of eQTLs affecting the expression of candidate genes in tissues potentially involved in the modulation of biological adaptations to high altitudes (i.e., HA-eQTLs) were reported in Supplementary Table 7, while HA-eQTLs falling in LASSI candidate adaptive windows were shortlisted in Supplementary Table 8.

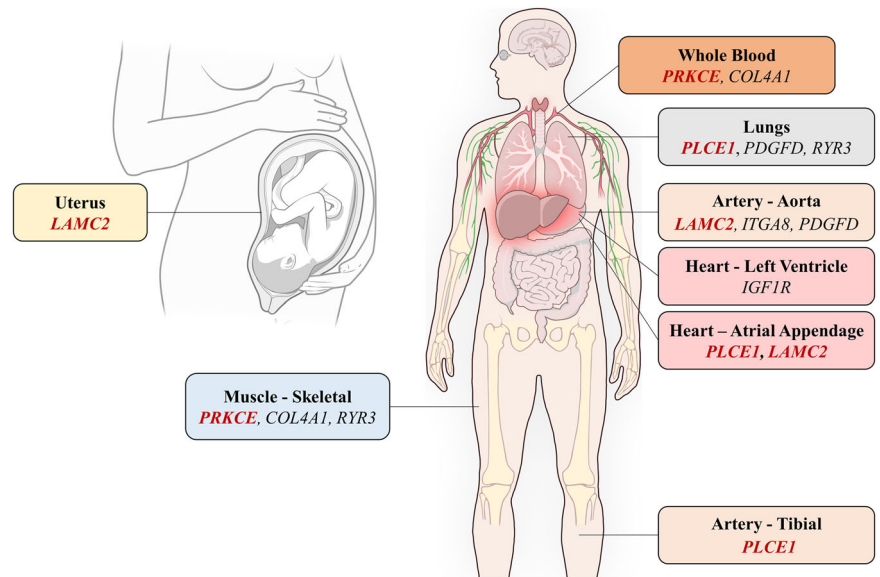
According to such an approach, the most compelling results were obtained for the *COL4A1*, *IGF1R*, *PDGFD*, *ITGA8*, *RYR3*, *LAMC2*, *COL4A4*, *PLCE1* and *PRKCE* genes, which turned out to be associated to haematological (e.g., haemoglobin concentration and haematocrit) and/or cardiovascular (e.g., heart rate and stroke) traits. Moreover, they included several HA-eQTLs that have been proved to influence gene expression in the aorta, heart, lung, skeletal muscle, blood and uterus (Supplementary Tables 6–8 and Fig. 3). Among them, *LAMC2*, *PLCE1* and *PRKCE* showed the greatest proportion of HA-eQTLs with respect to the total number of annotated eQTLs (Supplementary Table 7). Remarkably, 100 eQTLs in the *LAMC2* gene, 44 in *PLCE1* and 22 in *PRKCE* were included in LASSI candidate adaptive windows, modulating their expression respectively in the uterus/aorta/atrial appendage, in lungs/artery tibial/atrial appendage and in the skeletal muscle and whole blood tissues (Fig. 3, Supplementary Table 8). Similarly, 51 eQTLs on the *RYR3* gene, 11 on *COL4A1*, eight on *IGF1R* and four on *PDGFD* and *ITGA8* were included in adaptive genomic windows and regulate the expression of these loci in skeletal muscle, lungs, aorta, heart and blood tissues (Fig. 3, Supplementary Table 8).

Overall, this evidence further reinforces the functional links observed between the inferred candidate adaptive genes and complex mechanisms that have been long proposed to be finely regulated in both Andean and Tibetan populations^{70–72}. Additionally, they highlight possible associations of these adaptive changes with decreased risk of native Andean and Tibetan/Sherpa populations of developing pathological conditions that are instead more frequent in lowlanders exposed to the high-altitude stresses^{50–52,65}.

Conclusions

The identified genetic signatures might contribute to the increase in the density of blood vessels that results in an improved blood flow and oxygen delivery to tissues despite the hypoxic stress, especially leading to increased uterine blood flow during pregnancy, which is an adaptive trait qualitatively (but not quantitatively) similar between Andean and Himalayan groups^{52,73,74}. Although such a trait appears to be more enhanced in Tibetan/Sherpa populations with respect to Andean ones^{13,17,75}, the fact that several candidate loci identified in the present study (e.g., *PTK2*, *VEGFA*, *PDGFA*, *PDGFD*, *COL4A4*, *PIK3CB*, *IGF1R* and *LAMC2*) play a pivotal role in the development of vascular tissue specifically in placenta and embryo, and thus in the establishment of efficient maternal-foetal circulation^{32–35,56,66,67}, seems to support such an observation. Moreover, it might suggest that convergent adaptive evolution succeeded in optimising angiogenesis in these human groups mainly in early life and/or at the reproductive stage, thus leading to improved foetal development during intrauterine life (e.g., in terms of maturation of the respiratory system), which represents a key aspect to reduce neonatal mortality at high altitudes where efficiency of respiratory functions is crucial to ensure the individual’s survival. In addition, results obtained for the *PRKCE*, *RYR2* and *RYR3* genes

Fig. 3 | Candidate genes presenting eQTLs in putative adaptive genomic windows identified by the LASSI method and able to influence their expression in specific tissues. Schematic representation of the body districts in which the expression of the *COL4A1*, *IGF1R*, *PDGFD*, *ITGA8*, *RYR3*, *LMC2*, *PLCE1* and *PRKCE* candidate adaptive genes is modulated by eQTLs annotated in the GTEX portal. Genes reported in bold/red are those previously proposed to play an adaptive role also in populations of Tibetan/Sherpa ancestry^{52,60,62,65}. Illustration from NIAID NIH BIOART Source (<https://bioart.niaid.nih.gov/bioart/519>; <https://bioart.niaid.nih.gov/bioart/420>).



suggest that also some cardiovascular traits might have been shaped by natural selection in both Andean and Himalayan peoples^{50,51}. That being so, these findings highlighted polygenic mechanisms mediated by adaptive evolution of several focal adhesion and/or functionally related genes as the possible drivers of enhanced angiogenesis, oxygen transport and cardiovascular protection in Andean human groups similarly to what was previously observed in Tibetan/Sherpa populations, thus providing additional evidence for the molecular bases of their convergent adaptation to hypobaric hypoxia.

Methods

Assembled datasets

A WGS dataset including 9,439,267 SNVs for 24 Bolivian Aymara individuals who were born and live at around 3900 m of altitude was assembled from previously generated data¹⁵ and used to perform selection scans. Genome-wide genotype data were also collected for 141 individuals who were born and live between 3800 and 4200 m above sea level and belong to different Andean high-altitude populations that have been characterised by a largely shared genetic history^{22,23}. A written informed consent for data treatment was signed by each participant as attested in previous works^{15,22,23} and all ethical regulations relevant to human research participants were followed. Moreover, on 09/12/2019 the Ethics Committee of the University of Bologna released approval (prot. 205142) for the present study within the framework of the project ‘Genetic adaptation and acclimatisation to high altitude as experimental models to investigate the biological mechanisms that regulate physiological responses to hypoxia’ funded by Fondazione Cassa di Risparmio in Bologna (grant n. 2019.0552). In detail, 713,014 genetic variants were retrieved for 26 Bolivian Aymaras, as well as for 21, 22 and nine subjects respectively from Aymara, Quechua and Uros populations settled in the Peruvian Titicaca Lake area²³. Genotypes for 364,470 loci were collected also for 40 Quechua and 23 Aymara additional individuals²².

Data curation and imputation of the validation dataset

WGS data were submitted to QC procedures to filter for high-quality genotypes and to exclude possible closely related individuals. Specifically, the PLINK package v2.0⁷⁶ was used to retain only SNVs on autosomal chromosomes, variants/samples showing missing data <5% and loci with non-significant deviations from the Hardy-Weinberg equilibrium ($p > 1.059 \times 10^{-9}$). We also removed variants showing ambiguous A/T or G/C alleles and we calculated pairwise identity-by-descent (IBD) statistics by estimating the genome-wide proportions of shared alleles for each pair of individuals. Then, to exclude subjects related to the second degree, we

filtered out one individual from each pair showing an IBD coefficient >0.270, as previously proposed for populations of Native American ancestry⁷⁷ (Supplementary Information). This led to the generation of a high-quality WGS dataset including 7,966,198 SNVs and 24 individuals.

The described QC filters were applied also to the two assembled genome-wide high-altitude Andean datasets^{22,23} making available for validation analyses 681,932 and 364,413 variants, respectively. These data were merged by selecting only the 210,540 variants shared between both datasets and were phased by means of the Eagle algorithm v2.4⁷⁸ implemented in the Michigan Imputation Server (MIS) v1.2.4⁷⁹ using the 1000 Genomes Project Phase 3 and WGS datasets as reference panels. The MIS Minimac4 algorithm was then used to impute genotypes for 10,715,833 variants, which were submitted to the previously described QC procedures. In addition, the *bcftools* package v1.8⁸⁰ was used to retain only loci showing a squared Pearson correlation coefficient (r^2) between imputed genotype probability and true genotype call >0.95⁸¹. Finally, IBD kinship coefficients were calculated for each pair of individuals in the imputed dataset using the same approach described for WGS data. According to these post-imputation QC steps, a high-density imputed dataset including 130 Andean individuals characterised for 8,024,216 variants was obtained.

Assessing samples’ genetic ancestry and consistency between non-imputed and imputed data

To verify that the collected Bolivian Aymara WGS were representative of the genetic variation observable across Andean high-altitude populations, we first explored their distribution within the overall genomic landscape of Native American populations. For this purpose, the high-quality WGS dataset was merged with a reference panel made up of genome-wide genotypes from un-admixed individuals from Central and South American indigenous populations²³ and was submitted to Principal Component Analysis (PCA). Concurrently, to test also for consistency between non-imputed and imputed data, we performed PCA again by substituting the imputed data to the original ones as concerns the Andean populations included in the used Native American reference panel. In doing so, the *smartpca* method implemented in the EIGENSOFT package v6.0.1⁸² was used after having filtered the dataset for variants in high linkage disequilibrium (LD). LD pruning was obtained by considering sliding windows of 50 nucleotides sites in size and advancing by five loci at the time, as well as by removing variants for each pair showing $r^2 > 0.2$. Moreover, IBS-based matrices of individual pairwise differences in the genome-wide proportion of shared alleles were computed on both the non-imputed and imputed LD-pruned Andean datasets. These matrices were then compared by means of

the Mantel correlation test⁸³ using functions implemented in the R *vegan* package and empirically assessing statistical significance by running >10,000 permutations.

Selecting low-altitude control populations

To assess whether selection signatures detected in Andean people are informative about adaptations in response to high-altitude selective pressures, we selected low-altitude control populations of South American ancestry from the previously described reference panel of Native American groups²³ to be used for replicating selection scans. In doing so, our rationale was to identify South American groups that: (i) share an ancient common origin with Andeans, (ii) do not show evidence of remarkable admixture with Andeans occurred after the divergence from their common ancestors, (iii) did not experience high-altitude-related selective pressures during their evolutionary history. For this purpose, we calculated outgroup-*f*₃ statistic in the form of *f*₃(X, Andean; CHB) to formally assess the extent of shared drift between Andean populations and the other Central and South American populations (X). This was done using the *qp3Pop* function implemented in the ADMIXTOOLS package v3.0⁸⁴ by including only groups with more than five individuals and first by considering Andean populations separately (i.e., Aymara, Bolivian Aymara, Titicaca Aymara, Titicaca Quechua, and Titicaca Uros, respectively), and then by considering them as a whole (i.e., ANDEAN group).

Moreover, to accurately select the individuals to be included in the control group, we further investigated the genetic relationships between Andeans and the putative control populations pointed out by outgroup-*f*₃ analyses. To this end, we inferred individual-based genetic relationships by reconstructing haplotype sharing patterns by means of the CHROMOPAINTER/fineSTRUCTURE clustering approach⁸⁵ and by estimating ancestry proportions for each subject using the ADMIXTURE model-based clustering algorithm⁸⁶ (Supplementary Information). According to these procedures, we retained 24 individuals from low-altitude populations (i.e., eight Ashaninka, six Cashibo and 10 Shipibo), which have been characterised for 612,305 SNVs.

Inferring polygenic adaptive events from WGS and imputed data

A combination of selection scans (i.e., nSL, H12 and LASSI methods)^{36,87,88} suitable to detect both strong and weak selective events was applied to WGS, imputed and low-altitude control datasets. Collectively, these statistics were chosen to investigate genomic signatures left by the action of natural selection on the haplotype diversity of the examined populations, assuming that those potentially ascribable to polygenic adaptive mechanisms are represented by subtle changes in their haplotype structure and/or frequency spectrum rather than outstanding shifts in the frequency of single genetic variants. In detail, the nSL statistic was computed for each variant using *selscan* v.1.1.0b⁸⁹ and a 20,000 bp threshold for gap scale, 200,000 bp as the maximum gap length and 4500 consecutive loci as the maximum extension parameter⁸⁸. The H12 statistic was instead calculated with an ad hoc script using sliding windows of 200,000 bp and advancing by one variant at the time as previously suggested⁸⁷. Unstandardised values for each statistic were then normalised across multiple frequency bins by subtracting to each of them the bin average score and by dividing the resulting value by the related standard deviation. Finally, also the LASSI likelihood statistic was computed across sliding windows, but after having estimated the average number of SNVs included in all the haplotype blocks inferred for each dataset. For this purpose, we run the *--blocks* function implemented in the PLINK package v2.0⁷⁶ separately on WGS, imputed and low-altitude control datasets. Then, the LASSI algorithm was used to calculate a distorted haplotype frequency spectrum for each window and to associate to them a likelihood statistic informative of its conformity to a model of adaptive evolution (i.e., hard/soft sweep model) or neutral evolution. Calculation of the *m* parameter indicating the number of sweeping haplotypes (i.e., those carrying putative adaptive variants) was used to discriminate between hard (i.e., *m* = 1) and soft selective sweeps (i.e., *m* > 1). Such a method was used to validate results from H12/nSL analyses, because it has been proved to outperform them in

the identification of weak selective events according to simulations performed by considering a vast range of selection coefficients and demographic models³⁶.

In order to test a realistic approximation of a polygenic adaptation model, we retained as the representative scores of a given gene, the highest normalised nSL and H12 values, as well as the highest LASSI score computed among those obtained for all variants/windows annotated on that gene. In doing so, the overall distribution of the LASSI statistic was filtered to select windows showing *m* ≥ 2 to consider only weak selective events as potential contributors to polygenic adaptations. The resulting genome-wide distributions of selection scores were then submitted to gene-network analyses using the R *signet* algorithm⁹ and by setting 20,000 iterations to generate null distributions of the highest scoring subnetwork (HSS) for each gene-network of a given size to be compared with the obtained HSS. This provided a *p*-value for all the identified gene networks and those <0.05 after controlling the false discovery rate (FDR) were used to point out combinations of multiple genes that participate to the same functional pathway (according to information from the KEGG database²⁴) and that have been targeted by natural selection.

The most plausible biological functions having evolved adaptively in the Andean high-altitude groups were finally identified as those (i) supported by at least two out of the three selection scans implemented (i.e., nSL-based, H12-based and LASSI-based significant networks), (ii) replicated by applying the same pipeline of analyses to both WGS and imputed data, and (iii) not represented in significant networks obtained from the analysis of the low-altitude control dataset. Finally, to evaluate convergent evolution between Andean and Himalayan populations at a single-gene level, loci included in the supported networks were crosschecked with lists of genes whose variation patterns were previously proposed to have been shaped by the action of natural selection in Tibetan/Sherpa peoples^{13,52,65}.

Investigating association between candidate adaptive loci and complex traits/eQTLs

The CMDKP database (<https://hugeamp.org/>) was used to validate associations between biological functions/traits that have been previously proposed to be involved in adaptive responses to high-altitude selective pressures^{51,52,65,70–72} and the candidate adaptive genes identified by gene-network analyses.

Furthermore, we used the GTEx portal (<https://gtexportal.org/home/>) to search for eQTLs at the identified candidate adaptive loci and we retained only those variants affecting their expression in tissues and/or districts of the body that have been previously proposed to be involved in the modulation of high-altitude adaptations^{17,90}.

Statistics and reproducibility

Statistical analyses were performed using the softwares PLINK v2.0, SHAPEIT2 v2.r790, bcftools v 1.8, Eagle v2.4, MIS v1.2.4, EIGENSOFT v6.0.1, ADMIXTOOLS v3.0, CHROMOPAINTERv2, fineSTRUCTURE fs2.1, ADMIXTURE v.1.22, selscan v.1.1.0b, R v.3.6.3, LASSI GitHub repository available at <https://github.com/mdegiorgio/LASSI.git>. Phenotypic associations, eQTLs and pathways information in the present study were obtained consulting the CMDKP database (<https://hugeamp.org/>), the GTEx portal (<https://gtexportal.org/home/>) and the KEGG database (<http://www.genome.ad.jp/kegg/>). The sample sizes of each population used in the analyses are reported in Supplementary Table 1. The source data behind the graphs in the paper are listed in Supplementary Data 2.

Reporting summary

Further information on research design is available in the Nature Portfolio Reporting Summary linked to this article.

Data availability

WGS data that support the findings described in the present study are available at the NCBI Sequence Read Archive (accession n. PRJNA470966)¹⁵. Genotype data used for validation analyses are instead

available via figshare (https://figshare.com/articles/dataset/South_American_dataset_Gnecchi-Ruscone_et_al_2019_/7667174)³³. We provided all the Supplementary Tables cited in the paper in Supplementary Data 1. The source data behind the graphs in the main text are listed in Supplementary Data 2.

Code availability

All the codes used in the present study are available as functions that were either implemented in published R packages, GitHub public repositories or have been previously provided at https://github.com/paoloabondio/Ojeda-Granados_et_al_2021.

Received: 11 March 2024; Accepted: 25 February 2025;

Published online: 06 March 2025

References

1. Stern, D. L. The genetic causes of convergent evolution. *Nat. Rev. Genet.* **14**, 751–764 (2013).
2. Scott, R. J. & Spielman, M. Genomic imprinting in plants and mammals: how life history constrains convergence. *Cytogenet. Genome Res.* **113**, 53–67 (2006).
3. Wen, W., Alseikh, S. & Fernie, A. R. Conservation and diversification of flavonoid metabolism in the plant kingdom. *Curr. Opin. Plant Biol.* **55**, 100–108 (2020).
4. Chemes, L. B., de Prat-Gay, G. & Sánchez, I. E. Convergent evolution and mimicry of protein linear motifs in host–pathogen interactions. *Curr. Opin. Struct. Biol.* **32**, 91–101 (2015).
5. Edwards, M. et al. Association of the OCA2 polymorphism His615Arg with melanin content in East Asian populations: further evidence of convergent evolution of skin pigmentation. *PLoS Genet.* **6**. <https://doi.org/10.1371/journal.pgen.1000867> (2010).
6. Kwiatkowski, D. P. How malaria has affected the human genome and what human genetics can teach us about malaria. *Am. J. Hum. Genet.* **77**, 171–192 (2005).
7. Balentine, C. M. & Bolnick, D. A. Parallel evolution in human populations: a biocultural perspective. *Evolut. Anthropol.: Issues, N., Rev.* **31**, 302–316 (2022).
8. Pritchard, J. K. & Di Rienzo, A. Adaptation—not by sweeps alone. *Nat. Rev. Genet.* **11**, 665–667 (2010).
9. Gouy, A., Daub, J. T. & Excoffier, L. Detecting gene subnetworks under selection in biological pathways. *Nucleic Acids Res.* **45**. <https://doi.org/10.1093/nar/gkx626> (2017).
10. Lawrence E. S., et al. Humangenetics functional EPAS1/HIF2A missense variant is associated with hematocrit in Andean highlanders. **10**. <https://www.science.org> (2024).
11. Yu, J. J. et al. Time domains of hypoxia responses and—omics insights. *Front. Physiol.* **13**, 885295 (2022).
12. Zhang, X. et al. The history and evolution of the Denisovan-EPAS1 haplotype in Tibetans. *Proc. Natl. Acad. Sci. USA.* **118**, 1–9 (2021).
13. Bigham, A. et al. Identifying signatures of natural selection in Tibetan and Andean populations using dense genome scan data. *PLoS Genet.* **6**, e1001116 (2010).
14. Hernandez, R. D. et al. Classic selective sweeps were rare in recent human evolution. *Science* **331**, 920–924 (2011).
15. Lindo, J. et al. The genetic prehistory of the Andean highlands 7000 years BP though European contact. *Sci. Adv.* **4**, eaau4921 (2018).
16. Zhang, P. et al. Denisovans and Homo sapiens on the Tibetan Plateau: dispersals and adaptations. *Trends Ecol. Evol.* **37**, 257–267 (2022).
17. Beall, C. M. Two routes to functional adaptation: Tibetan and Andean high-altitude natives. *Proc. Natl. Acad. Sci. USA.* **104**(Suppl 1), 8655–8660 (2007).
18. Julian, C. G. & Moore, L. G. Human genetic adaptation to high altitude: evidence from the Andes. *Genes.* **10**, 150 (2019).
19. Bigham, A. W. Genetics of human origin and evolution: high-altitude adaptations. *Curr. Opin. Genet. Dev.* **41**, 8–13 (2016).
20. Crawford, J. E. et al. Natural selection on genes related to cardiovascular health in high-altitude adapted Andeans. *Am. J. Hum. Genet.* **101**, 752–767 (2017).
21. Jacovas, V. C. et al. Selection scan reveals three new loci related to high altitude adaptation in Native Andeans. *Sci. Rep.* **8**. <https://doi.org/10.1038/s41598-018-31100-6> (2018).
22. Reich, D. et al. Reconstructing Native American population history. *Nature* **488**, 370–374 (2012).
23. Gnecchi-Ruscone, G. A. et al. Dissecting the pre-Columbian genomic ancestry of Native Americans along the Andes–Amazonia divide. *Mol. Biol. Evol.* **36**, 1254–1269 (2019).
24. Kanehisa, M. & Goto S. KEGG: kyoto encyclopedia of genes and genomes. **28**. <http://www.genome.ad.jp/kegg/> (2000).
25. Jufri, N. F., Mohamedali, A., Avolio, A. & Baker, M. S. Mechanical stretch: physiological and pathological implications for human vascular endothelial cells. *Vasc. Cell.* **7**. <https://doi.org/10.1186/s13221-015-0033-z> (2015).
26. Cao, D. et al. Retinoic acid-related orphan receptor C regulates proliferation, glycolysis, and chemoresistance via the PD-L1/ITGB6/STAT3 signaling axis in bladder cancer. *Cancer Res.* **79**, 2604–2618 (2019).
27. Daniunaite, K. et al. Promoter methylation of prkcb, adamts12, and naalad2 is specific to prostate cancer and predicts biochemical disease recurrence. *Int. J. Mol. Sci.* **22**. <https://doi.org/10.3390/ijms22116091> (2021).
28. You, B. et al. Extracellular vesicles rich in HAX1 promote angiogenesis by modulating ITGB6 translation. *J. Extracell. Vesicles.* **11**. <https://doi.org/10.1002/jev2.12221> (2022).
29. Sayers, E. W. et al. Database resources of the national center for biotechnology information. *Nucleic Acids Res.* **50**, D20–D26 (2022).
30. Wang, S. et al. The angiogenic genes predict prognosis and immune characteristics in esophageal squamous cell carcinoma: evidence from multi-omics and experimental verification. *Front. Oncol.* **12**, 961634 (2022).
31. Chen, Z. et al. Microfibril-associated glycoprotein-2 promoted fracture healing via integrin $\alpha v \beta 3$ /PTK2/AKT Signaling. *Lab. Investig.* **103**. <https://doi.org/10.1016/j.labinv.2023.100121> (2023).
32. Shen, T. L. et al. Conditional knockout of focal adhesion kinase in endothelial cells reveals its role in angiogenesis and vascular development in late embryogenesis. *J. Cell Biol.* **169**, 941–952 (2005).
33. Adams, R. H. & Alitalo, K. Molecular regulation of angiogenesis and lymphangiogenesis. *Nat. Rev. Mol. Cell Biol.* **8**, 464–478 (2007).
34. Khankin, E. V., Royle, C. & Karumanchi, S. A. Placental vasculature in health and disease. *Semin. Thromb. Hemost.* **36**, 309–320 (2010).
35. Russell, M. W. et al. Damaging variants in proangiogenic genes impair growth in fetuses with cardiac defects. *J. Pediatr.* **213**, 103–109 (2019).
36. Harris, A. M. & DeGiorgio, M. A likelihood approach for uncovering selective sweep signatures from haplotype data. *Mol. Biol. Evol.* **37**, 3023–3046 (2020).
37. Kalluri, R. Basement membranes: structure, assembly and role in tumour angiogenesis. *Nat. Rev. Cancer* **3**, 422–433 (2003).
38. Alavi, M. V. et al. Col4a1 mutations cause progressive retinal neovascular defects and retinopathy. *Sci Rep.* **6**, 18602 (2016).
39. Kuo, D. S., Labelle-Dumais, C. & Gould, D. B. Col4a1 and col4a2 mutations and disease: Insights into pathogenic mechanisms and potential therapeutic targets. *Hum. Mol. Genet.* **21**. <https://doi.org/10.1093/hmg/dds346> (2012).
40. Xu, G. et al. The evolution of acquired resistance to BRAFV600E kinase inhibitor is sustained by IGF1-driven tumor vascular remodeling. *J. Invest. Dermatol.* **142**, 445–458 (2022).
41. Baker, J., Liu, J. P., Robertson, E. J. & Efstratiadis, A. Role of insulin-like growth factors in embryonic and postnatal growth. *Cell* **75**, 73–82 (1993).

42. Liu, J. P., Baker, J., Perkins, A. S., Roberteon, E. J. & Efetratiadie A. Mice carrying null mutations of the genes encoding insulin-like growth factor I (Igf.1) and type 1 IGF receptor (Igfir). *Cell* **75**, 59–72 (1993).
43. Liu, J. L., Yakar, S. & LeRoith, D. Mice deficient in liver production of insulin-like growth factor I display sexual dimorphism in growth hormone-stimulated postnatal growth. *Endocrinology* **141**, 4436–4441 (2000).
44. Woods, K. A., Camacho-Hübner, C., Savage, M. O. & Clark, A. J. Intrauterine growth retardation and postnatal growth failure associated with deletion of the insulin-like growth factor 1 gene. *N. Engl. J. Med.* **335**, 1363–1367 (1996).
45. Li, H., Fredriksson, L., Li, X. & Eriksson, U. PDGF-D is a potent transforming and angiogenic growth factor. *Oncogene* **22**, 1501–1510 (2003).
46. Sitras, V. et al. Differential placental gene expression in severe preeclampsia. *Placenta* **30**, 424–433 (2009).
47. Demydenko, K., Ekhteraei-Tousi, S. & Roderick, H. L. Inositol 1,4,5-trisphosphate receptors in cardiomyocyte physiology and disease. *Philos. Trans. R. Soc. B.: Biol. Sci.* **377**. <https://doi.org/10.1098/rstb.2021.0319> (2022).
48. Lehnart, S. E., Wehrens, X. H. T., Kushnir, A. & Marks, A. R. Cardiac ryanodine receptor function and regulation in heart disease. *Ann. N. Y. Acad. Sci.* **1015**, 144–159 (2004).
49. Daniels, R. E. et al. Cardiac expression of ryanodine receptor subtype 3; a strategic component in the intracellular Ca²⁺ release system of Purkinje fibers in large mammalian heart. *J. Mol. Cell Cardiol.* **104**, 31–42 (2017).
50. Busch, S. A. et al. Global REACH: assessment of brady-arrhythmias in Andeans and Lowlanders during apnea at 4330 m. *Front. Physiol.* **10**, 1603 (2020).
51. Bjertness, E., Wu, T., Stigum, H. & Nafstad, P. Acute mountain sickness, arterial oxygen saturation and heart rate among Tibetan students who reascend to Lhasa after 7 years at low altitude: a prospective cohort study. *BMJ Open*. **7**, e016460 (2017).
52. Gnecci-Ruscone, G. A. et al. Evidence of polygenic adaptation to high altitude from Tibetan and Sherpa genomes. *Genome Biol. Evol.* **10**, 2919–2930 (2018).
53. Smith, S. C. et al. Profiling bladder cancer organ site-specific metastasis identifies LAMC2 as a novel biomarker of hematogenous dissemination. *Am. J. Pathol.* **174**, 371–379 (2009).
54. Pei, Y. F., Liu, J., Cheng, J., Wu, W. D. & Liu, X. Q. Silencing of LAMC2 reverses epithelial-mesenchymal transition and inhibits angiogenesis in cholangiocarcinoma via inactivation of the epidermal growth factor receptor signaling pathway. *Am. J. Pathol.* **189**, 1637–1653 (2019).
55. Garg, M. et al. Laminin-5γ-2 (LAMC2) is highly expressed in anaplastic thyroid carcinoma and is associated with tumor progression, migration, and invasion by modulating signaling of EGFR. *J. Clin. Endocrinol. Metab.* **99**, E62–E72 (2014).
56. Wang, R. et al. Overexpressed LAMC2 promotes trophoblast over-invasion through the PI3K/Akt/MMP2/9 pathway in placenta accreta spectrum. *J. Obstet. Gynaecol. Res.* **49**, 548–559 (2023).
57. Cahill, A. G., Beigi, R., Heine, R. P., Silver, R. M. & Wax, J. R. Placenta accreta spectrum. *Am. J. Obstet. Gynecol.* **219**, B2–B16 (2018).
58. Bartels, H. C., Postle, J. D., Downey, P. & Brennan D. J. Placenta accreta spectrum: a review of pathology, molecular biology, and biomarkers. *Dis. Markers*. **2018**, 1507674 (2018).
59. Goh, W., Yamamoto, S. Y., Thompson, K. S. & Bryant-Greenwood, G. D. Relaxin, its receptor (RXFP1), and insulin-like peptide 4 expression through gestation and in placenta accreta. *Reprod. Sci.* **20**, 968–980 (2013).
60. Hu, H. et al. Evolutionary history of Tibetans inferred from whole-genome sequencing. *PLoS Genet.* **13**, 1–22 (2017).
61. Arciero, E. et al. Demographic history and genetic adaptation in the Himalayan region inferred from genome-wide SNP genotypes of 49 populations. *Mol. Biol. Evol.* **35**, 1916–1933 (2018).
62. Deng, L. et al. Prioritizing natural-selection signals from the deep-sequencing genomic data suggests multi-variant adaptation in Tibetan highlanders. *Natl. Sci. Rev.* **6**, 1201–1222 (2019).
63. Chen, Y. et al. Epigenetically upregulated oncoprotein PLCE1 drives esophageal carcinoma angiogenesis and proliferation via activating the PI-PLC- η -NF- κ B signaling pathway and VEGF-C/ Bcl-2 expression. *Mol. Cancer* **18**. <https://doi.org/10.1186/s12943-018-0930-x> (2019).
64. Scruggs, S. B., Wang, D. & Ping, P. PRKCE gene encoding protein kinase C- ϵ —dual roles at sarcomeres and mitochondria in cardiomyocytes. *Gene* **590**, 90–96 (2016).
65. Ferraretti, G. et al. Archaic introgression contributed to shape the adaptive modulation of angiogenesis and cardiovascular traits in human high-altitude populations from the Himalayas. *Elife*. **12**, RP89815 (2024).
66. Tong, J., Niu, Y., Chen, Z. J. & Zhang, C. Comparison of the transcriptional profile in the decidua of early-onset and late-onset preeclampsia. *J. Obstet. Gynaecol. Res.* **46**, 1055–1066 (2020).
67. Oefner, C. M. et al. Collagen type IV at the fetal-maternal interface. *Placenta* **36**, 59–68 (2015).
68. Okada, H., Tsuzuki, T. & Murata, H. Decidualization of the human endometrium. *Reprod. Med. Biol.* **17**, 220–227 (2018).
69. Plaisier, M. Decidualisation and angiogenesis. *Best Pract. Res. Clin. Obstet. Gynaecol.* **25**, 259–271 (2011).
70. Manier, G., Guenard, H., Castaing, Y., Varene, N. & Vargas, E. Pulmonary gas exchange in Andean natives with excessive polycythemia—effect of hemodilution. *J. Appl. Physiol.* **65**, 2107–2117 (1988).
71. Beall, C. M. et al. Hemoglobin concentration of high-altitude Tibetans and Bolivian Aymara. *Am. J. Phys. Anthropol.* **106**, 385–400 (1998).
72. Villafuerte, F. C., Cárdenas, R. & Monge-C, C. Optimal hemoglobin concentration and high altitude: a theoretical approach for Andean men at rest. *J. Appl Physiol.* **96**, 1581–1588 (2004).
73. Wu, D. et al. How placenta promotes the successful reproduction in high-altitude populations: a transcriptome comparison between adaptation and acclimatization. *Mol. Biol. Evol.* **39**. <https://doi.org/10.1093/molbev/msac120> (2022).
74. Sharma, V., Varshney, R. & Sethy, N. K. Human adaptation to high altitude: a review of convergence between genomic and proteomic signatures. *Hum. Genom.* **16**. <https://doi.org/10.1186/s40246-022-00395-y> (2022).
75. O'Brien, K. A. et al. Genomic selection signals in Andean highlanders reveal adaptive placental metabolic phenotypes that are disrupted in preeclampsia. *Hypertension* <https://doi.org/10.1161/HYPERTENSIONAHA.123.21748> (2023).
76. Chang, C. C. et al. Second-generation PLINK: Rising to the challenge of larger and richer datasets. *Gigascience*. **4**. <https://doi.org/10.1186/s13742-015-0047-8> (2015).
77. Ojeda-Granados, C. et al. Dietary, cultural, and pathogens-related selective pressures shaped differential adaptive evolution among native Mexican populations. *Mol. Biol. Evol.* **39**. <https://doi.org/10.1093/molbev/msab290> (2022).
78. Loh, P. R., Palamara, P. F. & Price, A. L. Fast and accurate long-range phasing in a UK Biobank cohort. *Nat. Genet.* **48**, 811–816 (2016).
79. Das, S. et al. Next-generation genotype imputation service and methods. *Nat. Genet.* **48**, 1284–1287 (2016).
80. Li, H. et al. The sequence alignment/map format and SAMtools. *Bioinformatics* **25**, 2078–2079 (2009).
81. Palmer, C. & Pe'er I. Bias characterization in probabilistic genotype data and improved signal detection with multiple imputation. *PLoS Genet.* **12**. <https://doi.org/10.1371/journal.pgen.1006091> (2016).
82. Patterson, N., Price, A. L. & Reich, D. Population structure and eigenanalysis. *PLoS Genet.* **2**, 2074–2093 (2006).

83. Mantel, N. *Cancer Research*. **27**. http://aacrjournals.org/cancerres/article-pdf/27/2_Part_1/209/2382183/cr0272p10209.pdf (1967).
84. Patterson, N. et al. Ancient admixture in human history. *Genetics* **192**, 1065–1093 (2012).
85. Lawson, D. J., Hellenthal, G., Myers, S. & Falush D. Inference of population structure using dense haplotype data. *PLoS Genet.* **8**. <https://doi.org/10.1371/journal.pgen.1002453> (2012).
86. Alexander, D. H., Novembre, J. & Lange, K. Fast model-based estimation of ancestry in unrelated individuals. *Genome Res.* **19**, 1655–1664 (2009).
87. Garud, N. R., Messer, P. W., Buzbas, E. O. & Petrov, D. A. Recent selective sweeps in North American *Drosophila melanogaster* show signatures of soft sweeps. *PLoS Genet.* **11**, 1–32 (2015).
88. Ferrer-Admetlla, A., Liang, M., Korneliussen, T. & Nielsen, R. On detecting incomplete soft or hard selective sweeps using haplotype structure. *Mol. Biol. Evol.* **31**, 1275–1291 (2014).
89. Szpiech, Z. A. & Hernandez, R. D. Selscan: An efficient multithreaded program to perform EHH-based scans for positive selection. *Mol. Biol. Evol.* **31**, 2824–2827 (2014).
90. Gilbert-Kawai, E. T., Milledge, J. S., Grocott, M. P. W. & Martin, D. S. King of the mountains: Tibetan and sherpa physiological adaptations for life at high altitude. *Physiology* **29**, 388–402 (2014).

Acknowledgements

We acknowledge support from the Fondazione Cassa di Risparmio in Bologna through the project ‘Genetic adaptation and acclimatisation to high altitude as experimental models to investigate the biological mechanisms that regulate physiological responses to hypoxia’, which was granted to M.S. (n. 2019.0552). We also would like to thank John Lindo for having kindly shared Aymara WGS data without which this work would have not been possible.

Author contributions

Data curation, formal analysis, investigation, writing original draft, G.F., A.I., S.S.; software, formal analysis, P.A., K.S., M.A., C.O.-G.; data curation, review and editing original draft, S.D.F., M.I., A.D., P.T.S., P.C., M.T., M.D.M., G.A.G.-R., L.N., A.C., G.M., D.P.; resources, P.G., D.L., D.P.; conceptualisation, resources, supervision, funding acquisition, methodology, writing, review and editing original draft, M.S.

Competing interests

The authors declare no competing interests.

Ethics

A written informed consent for data treatment was signed by each participant as attested in previous works^{15,22,23} and all ethical regulations

relevant to human research participants were followed. Moreover, on 09/12/2019 the Ethics Committee of the University of Bologna released approval (prot. 205142) for the present study within the framework of the project ‘Genetic adaptation and acclimatisation to high altitude as experimental models to investigate the biological mechanisms that regulate physiological responses to hypoxia’ funded by Fondazione Cassa di Risparmio in Bologna (grant n. 2019.0552).

Additional information

Supplementary information The online version contains supplementary material available at <https://doi.org/10.1038/s42003-025-07813-6>.

Correspondence and requests for materials should be addressed to Marco Sazzini.

Peer review information *Communications Biology* thanks Wanjun Gu and the other, anonymous, reviewer(s) for their contribution to the peer review of this work. Primary Handling Editor: Ophelia Bu. A peer review file is available.

Reprints and permissions information is available at <http://www.nature.com/reprints>

Publisher’s note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Open Access This article is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License, which permits any non-commercial use, sharing, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if you modified the licensed material. You do not have permission under this licence to share adapted material derived from this article or parts of it. The images or other third party material in this article are included in the article’s Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article’s Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by-nc-nd/4.0/>.

© The Author(s) 2025