

Received: 22 July 2016 Accepted: 01 November 2016 Published: 22 November 2016

OPEN The response of the soil microbial food web to extreme rainfall under different plant systems

Feng Sun^{1,2}, Kaiwen Pan¹, Akash Tariq^{1,2}, Lin Zhang¹, Xiaoming Sun¹, Zilong Li^{1,2}, Sizhong Wang^{1,2}, Qinli Xiong^{1,2}, Dagang Song^{1,2} & Olusanya Abiodun Olatunji^{1,2}

An agroforestry experiment was conducted that involved four planting systems: monoculture of the focal species Zanthoxylum bungeanum and mixed cultures of Z. bungeanum and Capsicum annuum, Z. bungeanum and Medicago sativa and Z. bungeanum and Glycine max. Soil microbial food web (microorganisms and nematodes) was investigated under manipulated extreme rainfall in the four planting systems to assess whether presence of neighbor species alleviated the magnitude of extreme rainfall on nutrient uptake of the focal species by increasing the stability of soil food web. Our results indicate that in the focal species and G. max mixed culture, leaf nitrogen contents of the focal species were higher than in the monoculture and in the other mixed cultures under extreme rainfall. This result was mainly due to the significant increase under extreme rainfall of G. max species root biomass, resulting in enhanced microbial resistance and subsequent net nitrogen mineralization rate and leaf nitrogen uptake for the focal species. Differences in functional traits of neighbors had additive effects and led to a marked divergence of soil food-web resistance and nutrient uptake of the focal species. Climate change can indirectly alleviate focal species via its influence on their neighbors.

Anthropogenic disturbances are continuously causing climatic changes and challenges for ecosystem stability. Under the current scenarios of climate change, extreme precipitation is expected to increase over most regions of the world, particularly in the high-latitude and tropical regions and I. In China, a significant increase in extreme precipitation has occurred in the west, in the mid-lower reaches of the Yangtze River, and in parts of the southwest². The changes in precipitation regimes will be more spatially heterogeneous and less predictable than other climate changes and will leads to more intense flooding occur at local and regional scales³. Scientists have been committed to ecosystem properties to stabilize and increase community stability under climate change^{4,5}.

Ecological stability consists of resistance and resilience⁶. Resistance is the degree of change caused by a disturbance. Resistance as an ecological concept draws the ecologist's attention to how to increase the resistance and stability of a habitat to enable communities and species to overcome the effects of climate change. Various factors are involved in resistance mechanisms and ecological stability, such as plant community composition and soil

As one property in an ecosystem, plant community composition can increase or decrease the community's capacity for stability⁸⁻¹¹. For example, previous studies suggested that neighbor communities had positive, negative or no effects on the performance of focal species under heavy rainfall^{11,12}. Other studies suggested that legumes facilitate or compete with the focal species, depending on the legume species, under heavy rainfall^{8,10}. Thus, the composition of shifting plant species is the main driver for the production of the focal species in response to extreme rainfall. However, scientific evidence for the response of both the ecosystem and the plants to extreme rainfall is still insufficient¹³. The degree to which the response of the focal species is determined by the response of its neighbors is poorly understood 14. Moreover, most previous studies have focused on the relationship between the composition of the plant species and primary productivity. Few studies have examined the effects of the composition of plant species in increasing the uptake of nutrients for the focal species via soil food-web stability¹⁵. In theory, the presence of different plant species (especially the presence of legumes) with different above-and

¹CAS Key Laboratory of Mountain Ecological Restoration and Bioresource Utilization & Ecological Restoration Biodiversity Conservation Key Laboratory of Sichuan Province, Chengdu Institute of Biology, Chinese Academy of Sciences, Chengdu 610041, People's Republic of China. ²University of Chinese Academy of Sciences, Beijing, 100039, People's Republic of China. Correspondence and requests for materials should be addressed to K.P. (email: pankw@ cib.ac.cn) or X.S. (email: sunxm@cib.ac.cn)

belowground functional traits could potentially increase soil food-web complexity and stability^{16,17}. However, stronger competition among different species may also decrease the stability of the soil food web under extreme rainfall, even in the presence of a legume¹⁰. The present study examines these contradictory hypotheses by exploring the stability of soil microbial and nematode community responses to variations in plant community composition and extreme rainfall.

There is a close link between aboveground and belowground sub-systems because plants provide organic matter to decomposing microbes, which are an important part of the soil food web and provide nutrients to the soil by decomposing a plant's dead organic matter¹⁸. Recent investigations showed that subordinate plant diversity increases nematode diversity, abundance and stability¹⁷. However, many researchers have revealed that the species identity has a stronger influence on the nematode community than the species richness^{19,20}. Thus, it is not clear whether the positive biodiversity effect is due to species richness (true biodiversity effect) or to the individual species in a mixture (identical effect).

Extreme rainfall regimes will quickly change soil water content with consequences for habitat, soil food-web stability and resource availability, all of which may cascade onto plant growth^{5,21}. Previous studies suggested that the soil microbial community stability indexes and the nematode stability indexes were not significantly affected by plant diversity against extreme rainfall disturbance, indicating a limited potential from plant diversity to buffer soil food webs under extreme rainfall. However, Mariotte *et al.*²² suggested that subordinate species moderated drought effects on earthworm communities due to the better drought resistance of subordinate species. Taylor and Wolters found that oribatid mite communities in beech litter were less sensitive to summer drought than those in spruce litter due to the presence of drought-tolerant oribatid mite species, which were strongly related to the litter types²³. Thus, the plant-soil system can buffer against drought events. Using soil microbial and nematode communities that also have strong species-specific relationships, we hypothesized that plant community composition might be important in affecting soil food-web stability and the nutrient uptake of the focal species under extreme rainfall.

An agroforestry system is an intercropped growth of forest trees and crop plants. This combination of plants helps to improve soil-available nitrogen, increase carbon sequestration, enhance soil biodiversity, reduce threats and ultimately increase agricultural production. Thus, an agroforestry system is one of the potential tools to mitigate climate change, especially under extreme climate conditions²⁴.

Zanthoxylum is a widely growing genus in southwestern China, India, North America and Australia²⁵. The fruits are used to make Sichuan pepper and have medicinal uses such as for toothache or rheumatism. However, Zanthoxylum species such as Z. bungeanum easily succumb under continuous heavy rainfall, resulting in decreased yield and quality of fruits and increasing mortality rate²⁶. Most previous studies focused on increasing nutrient use in agroforestry systems; however, the effects of increasing soil food-web stability are neglected. These effects relate to the uptake of nutrients indirectly under extreme rainfall conditions.

The objective of this study was to assess whether an agroforestry system could increase the available nitrogen uptake of the focal species by stabilizing the soil microbial food web under extreme rainfall. The present study tested four hypotheses:1) specific species affect soil microbial and nematode communities, 2) extreme rainfall affects the soil food web differently under different plant communities, 3) legume species increase soil microbes and nematode stability, and 4) planting systems facilitate the focal species relay on species-specific neighbors under extreme rainfall.

Results

Soil properties. Two-way ANOVA showed that planting systems significantly (P < 0.05) affected soil NH₄⁺-N, NO₃⁻-N, dissolved organic carbon (DOC) and dissolved organic nitrogen (DON) (Table 1), extreme rainfall significantly (P < 0.01) affected soil water content, NH₄⁺-N, NO₃⁻-N and DOC (Table 1). There was a significant (P < 0.05) interaction between planting system and extreme rainfall on soil NO₃⁻-N (Table 1).

In the normal rainfall treatment, the Z-M mixed culture significantly (P < 0.05) increased soil NH₄⁺-N when compared with the monoculture and other mixed cultures (Table 1). Soil NO₃⁻-N, DOC and DON were greater in the legume mixed culture (P < 0.05) plots than in the monoculture plots and the Z-C mixed culture. Under extreme rainfall, soil NH₄⁺-N, DOC and DON were greater in the legume mixed culture (P < 0.05) plots than in the monoculture plots (Table 1).

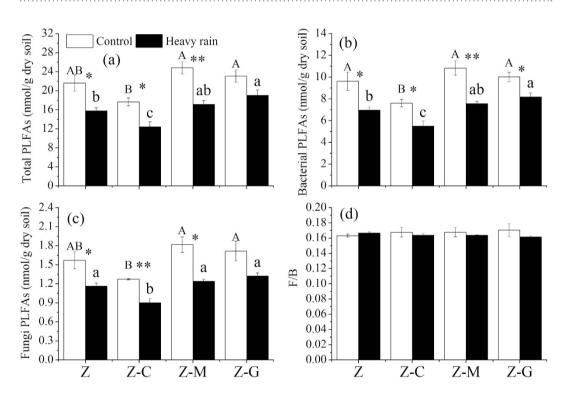
Microbial community. The Z-M and the Z-G mixed cultures significantly (P < 0.05) increased total microbial, bacterial and fungal biomass compared with the Z-C mixed culture in normal rainfall condition (Fig. 1). However, the planting system did not have significant effects on the ratios of fungi to bacteria (Fig. 1d).

Compared with normal rainfall, extreme rainfall significantly (P < 0.05) decreased total microbial and fungal biomass in the Z, the Z-C and the Z-M cultures and significantly (P < 0.05) decreased bacterial biomass in the monoculture and mixed cultures (Fig. 1). Under extreme rainfall, the Z-G mixed culture significantly (P < 0.05) increased total microbial and bacteria biomass compared with the monoculture and the Z-C mixed culture. The Z-C mixed culture had a significantly (P < 0.05) lower fungal biomass than the monoculture and the legume mixed culture (Fig. 1c).

Nematode community. Under normal rainfall, the Z-C mixed culture had significantly (P < 0.05) lower total nematode as well as herbivores and bacterivores densities than the monoculture and the legume mixed culture (Fig. 2). The Z-M mixed culture had significantly (P < 0.05) lower fungivores density than the monoculture and the Z-C and the Z-G mixed culture but had a significantly (P < 0.05) higher omnivore-predators density than the Z-C mixed culture (Fig. 2). The Z-G mixed culture had significantly (P < 0.05) greater bacterivores density than the monoculture and the other mixed cultures (Fig. 2c).

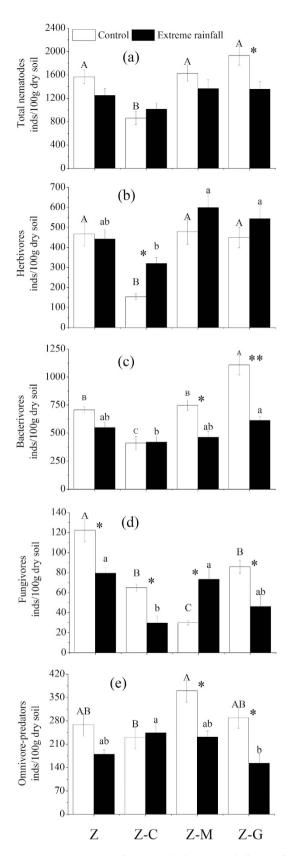
Planting system	Z			Z-C			Z-M			Z-G					System*
Soil properties	Control	Extreme rain	Sig.	Control	Extreme rain	Sig.	Control	Extreme rain	Sig.	Control	Extreme rain	Sig.	System	Rainfall	
Soil water content (%)	18.63 ± 0.80	30.26 ± 0.15	***	18.81 ± 0.24	30.72 ± 0.71	***	19.42 ± 0.67	31.82±0.61	***	19.14 ± 0.54	30.46 ± 0.69	***	ns	***	ns
Soil temperature (°C)	15.59 ± 0.07	15.51 ± 0.10		15.42 ± 0.09	15.39 ± 0.03		15.90 ± 0.18	15.76±0.06		16.10 ± 0.36	15.67 ± 0.07		ns	ns	ns
pН	7.62 ± 0.02	7.57 ± 0.01		7.67 ± 0.04	7.64 ± 0.03		7.59 ± 0.02	7.56 ± 0.03		7.57 ± 0.03	7.56 ± 0.01		ns	ns	ns
NH ₄ ⁺ -N (mg/kg)	4.16 ± 0.41B	5.97 ± 0.66b		$3.70 \pm 0.14B$	7.70 ± 0.61ab	**	$6.07 \pm 0.86 \mathrm{A}$	9.18 ± 0.99a		4.31 ± 0.49B	9.04 ± 0.94a	*	*	***	ns
NO ₃ ⁻ -N (mg/ kg)	17.81 ± 0.45 B	9.62 ± 0.59	***	19.86 ± 1.05B	8.71 ± 0.64	**	24.94 ± 0.71 A	10.52 ± 1.01	***	25.19 ± 1.12 A	11.30 ± 0.79	**	***	***	*
DOC (mg/kg)	$212.88 \pm 1.06B$	189.80 ± 4.43 b	*	207.07 ± 1.70B	193.22 ± 3.63 b	*	237.94 ± 7.07 A	223.27 ± 5.53a		$229.37 \pm 1.98 \mathrm{A}$	216.40 ± 4.11a	*	***	***	ns
DON (mg/kg)	55.25 ± 0.49B	$52.82 \pm 0.80b$		56.14 ± 3.09B	$53.61 \pm 1.14b$		$64.89 \pm 2.25 \mathrm{A}$	$62.04 \pm 2.85a$		$62.83 \pm 0.89 \mathrm{A}$	$61.43 \pm 2.91a$		***	ns	ns

Table 1. Soil physical and chemical properties in study site. Mixed cultures of *Z. bungeanum* and *G. max* (Z-G), *Z. bungeanum* and *M. sativa* (Z-M), and *Z. bungeanum* and *C. annuum* (Z-C), and monoculture *Z. bungeanum* (Z). Different uppercase letters indicate significant differences among control (normal rainfall) treatments; different lowercase letters indicate significant differences among extreme rainfall treatments. ANOVA was used to assess the effects of extreme rainfall on soil properties. *P < 0.05, **P < 0.01, ***P < 0.001.



In the monoculture, extreme rainfall only significantly (P < 0.05) decreased fungivores density (Fig. 2d). In the Z-C mixed culture, extreme rainfall significantly (P < 0.05) increased herbivores density but significantly (P < 0.05) decreased fungivores density (Fig. 2b,d). In the Z-M mixed culture, extreme rainfall significantly (P < 0.05) decreased bacterivores and omnivore-predators density but significantly (P < 0.05) increased fungivores density (Fig. 2c-e). In the Z-G mixed culture, extreme rainfall significantly (P < 0.05) decreased total nematode as well as bacterivores, fungivores, and omnivore-predators density (Fig. 2).

The two-way ANOVA showed that planting systems significantly (P < 0.05) affected the density of *Pratylenchus, Rhabditis, Mesorhabditis, Acrobeloides, Alaimus, Aphelenchus, Ditylenchus,* and *Mesodorylaimus* (Table 2). Extreme rainfall significantly (P < 0.05) affected the density of *Pratylenchus, Rhabditis, Mesorhabditis, Aphelenchus, Ditylenchus,* and *Mesodorylaimus* (Table 2). The interaction effects of planting systems and



Planting system		Z			Z-C			Z-M			Z-G					System*
Genus	Ср	Control	Extreme rain	Sig.	Control	Extreme rain	Sig.	Control	Extreme rain	Sig.	Control	Extreme rain	Sig.	System	Rainfall	Rainfall
Pratylenchus	PP3	52 ± 15	114 ± 10ab	*	34±7	87 ± 5bc	**	45 ± 15	48 ± 9c		56±19	153 ± 29a	*	**	***	ns
Rhabditis	Ba1	$167\pm16\mathrm{B}$	108 ± 3a	*	$115\pm20B$	36±11b	*	$122\pm13B$	75 ± 6a	*	563 ± 98 A	87 ± 17a	**	***	***	***
Mesorhabditis	Ba1	$35\pm12B$	7 ± 4		0B	15±4	*	25 ± 5B	11±6		$172\pm19\mathrm{A}$	7 ± 4	**	***	***	***
Acrobeloides	Ba2	99 ± 8 A	131 ± 25a		51 ± 2B	67 ± 8bc		92 ± 14 A	11 ± 6c	**	95 ± 16 A	89 ± 22ab		**	ns	**
Alaimus	Ba4	57 ± 8B	44±3b		46 ± 2B	78 ± 6ab	**	$130\pm13\mathrm{A}$	59 ± 13b	*	$32\pm3B$	103 ± 15a	**	**	ns	***
Aphelenchus	Fu2	$90\pm12\mathrm{A}$	45±9	*	47 ± 1B	18±6	*	8±4C	18±6		27 ± 6BC	38 ± 11		***	*	**
Ditylenchus	Fu2	$18 \pm 5B$	6 ± 3b		4 ± 2B	0 ± 0b		9 ± 5B	21 ± 4a		40 ± 11 A	4 ± 4a	*	*	*	**
Diphtherophora	Fu3	14±2	10 ± 5ab		9±3	6±3b		9±5	27 ± 9a		5±5	4 ± 4b		ns	ns	ns
Mesodorylaimus	Op5	69±16BC	10 ± 1c	*	26±1C	44 ± 10b		151 ± 26 A	66 ± 5a	*	$123\pm14\text{AB}$	21 ± 5c	**	***	***	**

Table 2. Response of nematode density in different plant patterns under control and extreme rainfall (Individuals per 100 g dry soil) (mean \pm SE). Mixed cultures of *Z. bungeanum* and *G. max* (Z-G), *Z. bungeanum* and *M. sativa* (Z-M), and *Z. bungeanum* and *C. annuum* (Z-C), and monoculture *Z. bungeanum* (Z). Different uppercase letters indicate significant differences among control (normal rainfall) treatments; different lowercase letters indicate significant differences among extreme rainfall treatments. ANOVA was used to assess the effects of extreme rainfall on soil properties. *P < 0.05, **P < 0.01, ***P < 0.001.

Pattern				
Resistance index	z	Z-C	Z-M	Z-G
Total microbes	0.58 ± 0.04 b	0.54 ± 0.05b	0.53 ± 0.02b	0.71 ± 0.003 a
Bacteria	$0.58\pm0.04ab$	$0.57 \pm 0.04 ab$	0.55 ± 0.04 b	$0.69 \pm 0.01a$
Fungi	0.6 ± 0.04	0.55 ± 0.06	0.52 ± 0.03	0.64 ± 0.05
Total nematodes	0.67 ± 0.06	0.69 ± 0.04	0.75 ± 0.13	0.54 ± 0.03
Herbivores	$0.91 \pm 0.06a$	-0.04 ± 0.02 b	0.61 ± 0.16a	0.65 ± 0.16a
Bacterivores	0.63 ± 0.07b	$0.87 \pm 0.01a$	$0.46 \pm 0.07c$	$0.38 \pm 0.02c$
Fungivores	$0.48 \pm 0.02a$	$0.3 \pm 0.08a$	-0.15 ± 0.12 b	0.39 ± 0.13a
Omnivores-predators	0.53 ± 0.09 b	0.87 ± 0.10a	0.46 ± 0.04 b	$0.38 \pm 0.08b$

Table 3. Soil food web resistance index in different patterns under extreme rainfall. Mixed cultures of *Z. bungeanum* and *G. max* (Z-G), *Z. bungeanum* and *M. sativa* (Z-M), and *Z. bungeanum* and *C. annuum* (Z-C), and monoculture *Z. bungeanum* (Z). Different lowercase letters indicate significant differences among plant patterns under extreme rainfall treatments.

extreme rainfall significantly (P < 0.05) affected the density of *Rhabditis*, *Mesorhabditis*, *Acrobeloides*, *Alaimus*, *Aphelenchus*, *Ditylenchus*, and *Mesodorylaimus* (Table 2).

Under normal rainfall, the monoculture significantly (P < 0.05) increased *Aphelenchus* density when compared to all of the mixed cultures. The Z-G mixed culture significantly (P < 0.05) increased the *Rhabditis* and the *Mesorhabditis* densities (Table 2).

Resistance index. Under extreme rainfall stress, the Z-G mixed culture significantly (P < 0.05) increased the total microbe resistance index versus monoculture and the Z-C and the Z-M mixed culture but significantly (P < 0.05) decreased the bacterivores resistance index (Table 3). The Z-C mixed culture significantly (P < 0.05) decreased the herbivores resistance index compared with the monoculture, the Z-M and the Z-G mixed culture but significantly (P < 0.05) increased the omnivores-predators resistance index. The Z-M mixed culture had the lowest bacteria and fungivores resistance index (Table 3).

Net nitrogen mineralization. Under normal rainfall, Z-M and Z-G significantly (P < 0.05) increased the net N mineralization rate when compared to Z and Z-C (Fig. 3). Extreme rainfall significantly (P < 0.05) decreased the net nitrogen mineralization rate in both the monoculture and the mixed culture. Under extreme rainfall, the Z-G mixed culture had a significantly (P < 0.05) higher net N mineralization rate than either the Z or the Z-C mixed cultures (Fig. 3).

Plant leaf nitrogen. Under normal rainfall, the focal species of Z in the Z-G mixed culture had higher leaf nitrogen content than the Z-C mixed culture (P < 0.05) (Fig. 4a). The neighbors of M had significantly higher leaf N contents than C and G (Fig. 4b). Extreme rainfall significantly (P < 0.05) decreased Z leaf N contents in the Z-M mixed culture, extreme rainfall also tended to decrease Z leaf N contents in the monoculture (P = 0.077). However, extreme rainfall significantly (P < 0.05) increased M leaf N contents in the Z-M mixed culture. Under extreme rainfall, the focal species Z in the Z-G mixed culture had significantly (P < 0.05) higher leaf N contents

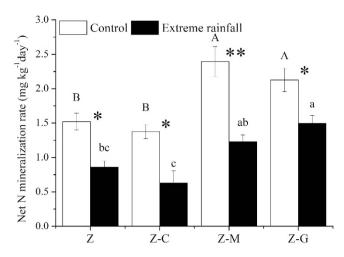


Figure 3. Response of net nitrogen mineralization rate in different planting systems under control and extreme rainfall. Mixed cultures of Z. bungeanum and G. max (Z-G), Z. bungeanum and M. sativa (Z-M), and Z. bungeanum and G. annuum (Z-C), and monoculture Z. bungeanum (Z). Different uppercase letters indicate significant differences among control (normal rainfall) treatments; different lowercase letters indicate significant differences among extreme rainfall treatments. Asterisks indicate significant differences between the bars (*P<0.05, **P<0.01; post hoc ANOVA).

than the monoculture, the Z-C or the Z-M mixed culture (Fig. 4a). The G species had the lowest leaf N contents when compared with C and M (Fig. 4b).

Root biomass. Under normal rainfall, there was no significant difference between the neighboring species root biomass (Fig. 4c). Extreme rainfall significantly decreased the Z-M root biomass but significantly increased the Z-G root biomass. Under extreme rainfall, the Z-G mixed culture had a higher root biomass than the Z-M (Fig. 4c).

Correlation analysis. Under extreme rainfall, the focal species of *Z* leaf N contents had a significant positive correlation with the total microbial resistance index, the bacterial resistance index and the net nitrogen mineralization rate but had a significant negative correlation with the total nematode resistance index, the bacterivores resistance index and the omnivores-predators resistance index (Table 4). However, the neighboring species leaf N contents had a significant negative correlation with the fungivores resistance index only (Table 4).

Discussion

Legume addition significantly increased the soil-available N, DOC and DON, which was apparently attributed to the N-fixing ability of *G. max* and *M. sativa*. Therefore, the input of high-nitrogen residuals (litter and/or root exudates) to soils is the most likely reason for the enhancement of soil carbon and nitrogen content seen in this study. Furthermore, a previous study also recognized that the presence of legume species increased the soil carbon and nitrogen contents²⁷. Extreme rainfall significantly decreased soil NO₃⁻-N and DOC due to greater soil leaching.

Previous studies have found that mixed cultures (especially the presence of a legume) significantly increase microbial biomass because of enhanced litter and rhizodeposition input compared with monocultures 28,29 . However, in the present study, no significant differences were found in terms of total microbial, bacterial, fungal biomass and the ratio of fungi to bacteria in soils between the mixed cultures and the monoculture under normal rainfall (Fig. 1). This finding is possibly due to legume fixation of atmospheric N_2 leading to higher levels of nitrogen in and around the plant (Table 1) and resulting in the rapid growth of bacteria. However, legume-induced changes in soil microorganisms might exert bottom-up control on higher trophic level organisms, such as microbivores and omnivore-predators, via the food web 28 . Our results also suggested that the legume mixed culture had higher bacterivores or omnivore-predator nematodes (Fig. 2). The legume mixed culture had a higher microbial biomass than the *C. annuum* mixed culture, likely because *C. annuum* produces poor-quality litter and root exudates compared to legume species 30 .

Previous studies have revealed that species identity, rather than species richness, has a stronger influence on nematode communities 19,20 . Given their ability to fixed N, legumes are the most common species that consistently influence soil nematodes 27 . Although a recent study argues that neighboring plant diversity increases nematode diversity, abundance and resistance 17 , the role of legumes as a key species in plant communities is well established. Likewise, our study provides evidence that legume addition significantly enhanced bacterivores in the *G. max* mixed culture and tended to increase omnivore-predator nematodes in the *M. sativa* mixed culture (p = 0.064) when compared to the monoculture (Fig. 2c,e). The *G. max* mixed culture significantly increased bacterivores mainly due to the significant increase of *Rhabditis* and *Mesorhabditis*, which both belong to *Ba1* bacterivores (Table. 2). *Ba1* bacterivores have short life spans and are known to rapidly enrich available resources 31 . NO $_3$ -N, DOC and DON contents were higher in the *G. max* mixed culture soil, confirming its nutrient-rich status (Table 1). Therefore, nematode abundance maybe a useful measure to indicate available soil resources.

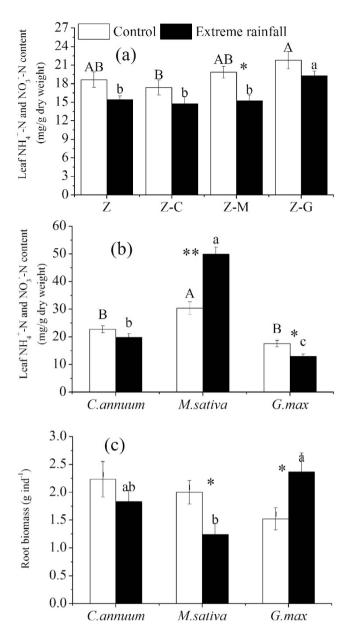


Figure 4. Response of leaf nitrogen contents in focal species and neighbor species and root biomass of neighbor species under control and extreme rainfall. Mixed cultures of Z. bungeanum and G. max (Z-G), Z. bungeanum and M. sativa (Z-M), and Z. bungeanum and G. annuum (Z-G), and monoculture Z. bungeanum (Z). Different uppercase letters indicate significant differences among control (normal rainfall) treatments; different lowercase letters indicate significant differences among extreme rainfall treatments. Asterisks indicate significant differences between the bars (*P<0.05, **P<0.01; post hoc ANOVA).

Consistent with the present study, legumes supported large populations of certain bacterivores, especially *Rhabditis* and *Panagrolaimus*, both of which are *Ba1* nematodes³², and legumes increased bacterivores, most notably *r*-selected *Rhabditida*¹⁹. However, our results showed that another legume, the *M. sativa* mixed culture, remarkably increased *Alaimus* (*Ba4*) and *Mesodorylaimus* (*Op5*) (Table 2). Enhancements of some k-selected nematodes (especially Op5), which occupy high trophic levels in soil food webs, could increase the complexity of the soil food web²⁷. The differences in the effects of the two legumes on soil nematodes might be due to plant chemistry. Indeed, plants supply food resources not only to plant-feeding nematodes but also to microbivores by forming litter above and belowground and by releasing large quantities of organic material from their living roots³³. Different plant species vary greatly in terms of litter quality and root exudates; therefore, their decomposers are varied.

Previous studies had suggested that *C. annuum* could only inhibit root-knot herbivores by releasing different secondary metabolites³⁰. The present study revealed interesting findings that *C. annuum* not only inhibits herbivores but also inhibits bacterivores and fungivores in the *C. annuum* mixed culture (Fig. 2c,d). Bacterial and fungal feeding nematodes play crucial roles in decomposing litter and increasing microbial activities in soil.

	Total microbe Resistance index	Bacteria Resistance index	Fungi Resistance index	Total nematodes Resistance index	Herbivores Resistance index	Bacterivores Resistance index	Fungivores Resistance index	Omnivore- predators Resistance index	Net Nitrogen mineralization rate
Z. bungeanum leaf N contents	0.786**	0.706 [*]	0.404	-0.823**	0.329	-0.632*	0.245	-0.714**	0.667*
Neighboring species leaf N contents	-0.625	-0.565	-0.477	0.505	0.240	-0.211	-0.779^{*}	-0.167	0.054

Table 4. Correlation analysis of soil biota resistance index and leaf NH₄⁺-N and NO₃⁻-N contents. $^*P < 0.05, ^{**}P < 0.01.$

Inhibitory effects on nematode density in the *C. annuum* mixed culture resulted in a significant decrease in the net nitrogen mineralization rate in comparison with the legume mixed culture (Fig. 3).

The *Z. bungeanum* monoculture had higher fungivores than the mixed cultures, mainly due to *Z. bungeanum's* waxy leaf, which is difficult to decompose. Slow decomposition of poor-quality litter (low C/N) favors fungi and fungivores, while rapid decomposition of high-quality litter (high C/N) favors bacteria and bacterivores.

Taken together, our results suggest that the identities of a single plant species and plant functional groups are the most important factors influencing microbial and nematode communities. The above findings support our first hypothesis that specific species affect soil microbial and nematode communities.

Extreme rainfall regimes will increase soil water content rapidly with consequences for both soil food web and habitat⁵. However, very few studies have evaluated the effect of extreme rainfall on soil microbial communities^{4,21}. The above two results suggest that extreme soil moisture significantly decreases microbial biomass and alters microbial communities because of low levels of oxygen and excessive amounts of water in the soil. These findings are consistent with our results that extreme rainfall significantly reduces total microbial, bacterial and fungal biomass (Fig. 1). First, an excessive amount of water in the soil causes the microbial cell membrane to rupture due to hypoxic condition³⁴. Second, this marked decline in microbial biomass corresponds with a significant increase in nutrient loss in the soil (Table 1). This pattern suggests that extreme rainfall-induced leaching is likely to be linked to the decrease in microbial biomass. However, the F/B ratio did not show any significant change under extreme rainfall because of a simultaneous reduction in bacterial and fungal biomass (Fig. 1d). Therefore, this finding indicated that the fungi also did not have sufficient capacity to resist extreme soil water content stress^{21,35}, although fungi have strong cell walls and intrinsic resistance potential against moisture stress events. This condition mainly exists because most fungi are located in/on the outer part of soil aggregates and are susceptible to extreme rainfall. Landesman and Dighton³⁶ suggested no significant change in microbial biomass and community composition in response to extreme rainfall. This discrepancy may be due to different saturation levels and textures of tested soils. Landesman and Dighton³⁶ investigated sandy soil with low water-holding capacity that cannot induce large fluctuations in soil moisture content, while in the present study, soil water content increased by up to 30%, and the soil had a higher capacity to hold water than sandy soil.

Nematodes respond rapidly to water stress because they depend upon water for movement and migration toward their prey in soil. Previous studies showed that increased soil moisture markedly decreased bacterivore abundance and increased herbivore density^{37,38}. However, a recent investigation suggested that extreme rainfall stress increased total nematode abundance depending on the diversity of the plant species⁴. Our findings suggest that extreme rainfall had a significant effect on the density of bacterivores and herbivores depending on the composition of the plant species (Fig. 2b,c). Extreme rainfall significantly increased herbivore abundance in the C. annuum mixed culture (Fig. 2), possibly because high soil moisture content decreased the level of secondary metabolites released by C. annum to the soil. A higher abundance of herbivores in the soil could have serious negative effects on both soil and plants by feeding the roots of C. annum (Fig. 4c). Extreme rainfall significantly decreased bacterivores in the G. max mixed culture and M. sativa mixed culture (Fig. 2c). In the G. max mixed culture, a decrease in bacterivore abundance was mainly due to the decreased density of Rhabditis and Mesorhabditis, both of which belong to Ba1 nematodes (Table 2). However, leaching induced by extreme rainfall had a limit effects on nematode dissemination³⁹. The reason for this might be that extreme rainfall significantly increased G. max's root biomass (Fig. 4c) and perhaps changed the root exudates, which negatively influenced Ba1 nematodes in response to extreme rainfall^{28,29}. Another reason might be that extreme rainfall disrupted the aggregate structure of soil where the habitable nematodes occur. In contrast to our study, a recent study suggested that Ba1 nematodes increase after extreme rainfall because of their high acclimatization potential due to their high reproduction rate and impermeable cuticle⁴. However, that study showed nematode densities increasing strongly with increasing plant diversity after the extreme rainfall. Different plants have different strategies for withstanding flooding disturbances, e.g., by aerating the soil and thereby maintaining activity⁴⁰. Thus, in the referenced study, a plant species' existing tolerance to extreme rainfall in a community with high plant diversity was another reason for the increase in Ba1 nematodes. In the M. sativa mixed culture in the present study, extreme rainfall mainly decreased the genera Rhabditis, Acrobeloides and Alaimus (Table 2). Nematodes responded differently in G. max and M. sativa mixed cultures under almost the same soil water content. Thus, the resistance of populations to soil organisms may be partly dependent on the traits of the respective plant species.

Extreme rainfall significantly decreased fungivores in monoculture as well as the *C. annuum* and *G. max* mixed cultures (Fig. 2). This pattern primarily occurred because extreme rainfall significantly decreased the genera *Aphelenchus* and *Ditylenchus*, which are cp-2 nematodes (Table 2). However, extreme rainfall significantly increased fungivores in the *M. sativa* mixed culture (Fig. 2). This result may be due to the significant reduction

in omnivore-predator abundance. Thus, the food source or feeding relationship is also a dominant aspect for structuring the soil food web.

In the present study, extreme rainfall significantly decreased omnivore-predator abundance in G. max and M. sativa mixed cultures and tended to decrease them in the monoculture (p = 0.074). The density of the genus Mesodorylaimus (cp-5) decreased significantly in these cultures (Table 2). Cp-5 nematode species are the type most susceptible to environmental stress because of their long life cycle, permeable cuticle and high sensitivity. However, recent investigations revealed that Cp-5 nematode species have a strong resistance to different environmental stresses because of their varied and unidentified feeding strategies and their complex interactions with higher trophic levels⁴¹. All of the above results confirm our second hypothesis that extreme rainfall affected the soil food web differently under different planting systems.

Although a recent study argued that greater plant diversity is responsible for decreasing ecosystem stability under flooding stress⁵, ecosystem stability is directly related to plant functional diversity, e.g., nitrogen-fixing legumes⁴². Thus, no consistent conclusion exists about how plant community composition affects ecological stability, especially soil food-web resistance.

In the present study, total microbial resistance was higher in the G. max mixed culture than in the monoculture, the C. annuum mixed culture and the M. sativa mixed culture under extreme rainfall stress (Table 3). This finding was mainly due to the significantly increased root biomass of G. max under extreme rainfall, which greatly stimulated the growth of adventitious roots (F. Sun, personal observation) and increased tolerance to hypoxia, both of which safeguarded the root activity and functionality⁴³. These factors also resulted in higher microbial resistance than was found in the monoculture or in the C. annuum and M. sativa mixed cultures. Another reason for this finding was that the reduction of bacterivores grazing on bacteria resulted in a higher microbial biomass than was found in the either the monoculture or the C. annuum mixed culture. Although the M. sativa mixed culture contained a legume species, the culture did not show an increase in microbial resistance (Table 3) because extreme rainfall significantly decreased the root biomass of M. sativa and reduced the input of root exudates to soil microorganisms. Therefore, microbial resistance depends on the composition and traits of the species. Bacterivores' resistance was lower in the legume mixed culture than it was in the monoculture (Table 3) due to the stress extreme rainfall had on bacterivores and their abundance (Fig. 2). The composition of the species, even in the presence of the legume, could not enhance soil microbial and nematode resistance simultaneously. This apparent trade-off in the soil food web is used to constrain the composition and diversity of the entire soil community. Thus, these results only partly support our third hypothesis that legume species could increase soil microbes and nematode stability.

Neighbor plants have direct positive effects on whole plant communities by altering and modifying physical and/or biotic conditions⁴⁴. Although the facilitation process is important for the structuring and functioning of an ecosystem, the mechanism behind this process is still poorly understood, especially in the belowground soil food web. Some studies suggest that leguminous plant presence enhances the N uptake of the focal species in ambient weather⁴⁵ and in extreme rainfall⁸. However, our results were inconsistent with previous findings. Specifically, under extreme rainfall, only the G. max mixed culture had higher leaf nitrogen contents in the focal species when compared to the monoculture, the *C. annuum* mixed culture and the *M. sativa* mixed culture (Fig. 4a). The reason for this was that extreme rainfall significantly increased the *G. max* species root biomass, thereby enhancing microbial resistance and the net nitrogen mineralization rate. Thus, this process increased soil available N and enhanced nutrient uptake of the focal species. Our data also suggested that the focal leaf N contents had a significant positive correlation with total microbial resistance, bacterial resistance and the net nitrogen mineralization rate but a negative correlation with total nematode resistance and the bacterivore resistance index (Table 4). Thus, maintaining lower trophic level stability (microbial resistance) is more important than maintaining higher trophic level stability (nematode resistance) under extreme rainfall stress. In the M. sativa mixed culture, although a legume species was also present, the focal species had lower leaf nitrogen contents than the G. max mixed culture (Fig. 4a). The main reason was the strong interspecific competition for nitrogen between M. sativa and the focal species under extreme rainfall (Fig. 4b). The leaf nitrogen contents of M. sativa were significantly higher (+64%) than they were under normal rainfall. The deterioration effects for the focal species under extreme rainfall occurred because of increased competition for nutrients. These findings confirmed the study's fourth hypothesis, that planting systems could facilitate the focal species' relay on species-specific neighbor species under extreme rainfall. Legume species could facilitate the focal species, but the effect is highly species specific, related to climate change disturbance, and mostly due to belowground community processes. Thus, different mixed cultures would result in various soil food web structures under extreme rainfall and subsequently different biogeochemical cycling, soil function and nutrient availability. This finding is important in agricultural management that will continue to change to alleviate extreme climate conditions under future climate change. Belowground communities must be integrated into land management and climate change mitigation strategies.

Materials and Methods

Study site. The experiment was carried out in Maoxian, Sichuan Province, southwestern China (103°53′E, 31°41′N), at an altitude of 1686 m above the sea level. The mean annual temperature, precipitation and evaporation were 8.9 °C, 920 mm and 796 mm, respectively, according to meteorological monitoring from the Maoxian Ecological Station of Chinese Academy of Science. Most rainfall occurred in August, and most drought occurred in February. In this area, a significant increase in extreme precipitation was found in a previous study². The soil was classified as Udic Luvisols⁴⁶.

Experimental design. Farmland soil with different organic fertilizers that is normally used to grow vegetables was use to establish the present experiment. The randomized complete block design (RCBD) was used to establish the experiment in April 2013. Four planting systems containing the monoculture *Zanthoxylum*

bungeanum (Z), and the mixed cultures Z. bungeanum and Capsicum annuum L. (Z-C), Z. bungeanum and Medicago sativa L. (Z-M) and Z. bungeanum and Glycine max (L.) Merr. (Z-G) were intercropped in an agroforestry experiment. Two-year-old uniform seedlings of Z. bungeanum were collected from a local nursery based on plant stem base diameter and height. Z. bungeanum belongs to the family Rutaceae and is a perennial plant reproductively, while M. sativa, G. max, and C. annuum are annual plants. M. sativa and G. max are leguminous plant species, while C. annuum is an herbaceous plant. The focal species Z. bungeanum was grown once, in the center of each plot. The neighboring species G. max, M. sativa and C. annuum were planted at the same density in all plots. Each plot area was $2.6 \times 2.6 \,\mathrm{m}^2$, and the distance between each plot was 1 m. Each pattern was replicated six times randomly, for a total of 24 plots. In Z monoculture, all the understory plants were completely removed. All the planting systems had no fertilization from the establishment of experiment, and the understory grasses were removed by hand each week; herbicide was not used to avoid potentially negative effects on soil organisms²⁹. In August 2015, we conducted precipitation treatments randomly. Control (normal rainfall) and extreme rainfall each had three replications. Control and extreme rainfall plots were maintained under rainout shelters during the experiment period. The shelters were constructed using steel frames and were covered with transparent plastic⁴⁷ to protect from precipitation from August 1 to 30. To minimize greenhouse effects, the rainout shelters were 1.8 m aboveground. Tap water was used to mimic extreme precipitation events, and a watering pot was used to compensate for rain. Rain regimes were designated according to Ng et al.48 in which the average rainfall in the area during August was 3.0 mm/day (based on the average rainfall data during 1983-2013 from the Maoxian Ecological Station of Chinese Academy of Science), and was designated as the control rain regime, while extreme rainfall was designated according to the abnormally high rainfall in August (9.5 mm/day). All the plots were watered (7-9 am and 18-20 pm) from August 1 to 30. Around all plots, thick PVC panels were inserted at the top of the 0.5 m soil layer to prevent lateral water movement between plots and to prevent interaction between roots from neighboring plots.

Field sampling. Soil sampling was conducted on 31 August 2015. A total of five soil cores (5 cm diameter) were collected using the five-spot method from a 10 cm soil depth and combined to form one composite sample for each plot. *Z. bungeanum* and neighbor species leaves were sampled on the same day, kept in a liquid nitrogen container, and taken back to the laboratory for determining the concentrations of leaf NH₄⁺-N and NO₃⁻-N. In total, three individual neighbor species were collected from each plot.

Soil properties analysis. Estimation of soil moisture content was performed gravimetrically by oven drying (105 °C for 24 h) 20 g of the field soil sample. A button thermometer (QT-ST010, Beijing Channel Science Equipment Co., LTD, China) was used for soil temperature measurement, and data were recorded from each plot at 1-h time intervals. Soil pH was estimated in a 1:2.5 soil/CaCl₂ (0.01 mol/L) suspension. Soil NH₄+-N and NO₃-N were determined with the help of a flow injection auto analyzer (AA3, Bran+Luebbe, Germany). Dissolved organic carbon (DOC) and nitrogen (DON) were estimated using a TOC/TN analyzer (Multi N/C[®] 2100(S), Analytik Jena AG, Germany). Net N mineralization rate was calculated according to the changes in NH₄+-N and NO₃--N before and after the 7-d incubation.

Phospholipid fatty acid (PLFA) analysis. Community structure and soil microbial biomass were determined using PLFA⁴⁹. Eight grams of dry-weight-equivalent fresh soil was used for the extraction of lipids from a 23 mL extraction mixture containing chloroform:methanol:phosphate buffer (1:2:0.8 v/v/v). The concentration of each PLFA was calculated based on the 19:0 internal standard concentrations, and the abundance of the individual fatty acids was measured as nmol lipid per gram of dry soil. Total microbial biomass was assessed using total extracted PLFAs, with the sum of i15:0, a15:0, 15:0, i16:0, $16:1\omega 7$, i17:0, a17:0, 17:0, cy17:0, $18:1\omega 7c$ and cy19:0 taken as an indicator of bacterial biomass, while $18:2\omega 6$, 9c PLFAs were considered for fungi identification^{28,50}. The above estimated biomass was used for the fungi to bacteria ratio.

Nematode community analysis. The modified cotton-wool filter method was followed to determine soil (50 g) nematodes⁵¹. From each sample, the total number was identified, and then 100 nematode specimens were randomly selected and identified with the help of an inverted compound microscope. Nematode populations were expressed as the number of nematodes per 100 g dry soil. Based on their feeding habits, nematodes were categorized into four groups: (1) bacterivores, (2) fungivores, (3) omnivores-predators and (4) herbivores⁵².

Resistance index analysis. Microbial and nematode resistance indexes were calculated according to Orwin and Wardle⁶, RS $(t_0) = 1 - (2|D_0|)/(c_0 + |D_0|)$, where c_0 represents the control value after disturbance, while $|D_0|$ represents the difference between control and disturbed soil.

Neighboring species root biomass analysis. All harvested neighboring root biomass was dried at 75 °C to constant weight and weighed.

Plant leaf nitrogen analysis. Plants readily take up and directly use two main forms of soil available nitrogen (NH $_4$ ⁺-N and NO $_3$ ⁻-N) for growth. For leaf NH $_4$ ⁺-N and NO $_3$ ⁻-N measurement, 0.2-g leaf samples were homogenized in 2 mL of 10% HCL solution and 5 mL of deionized water, respectively. The resultant supernatants were analyzed using the quantitative colorimetric method⁵³.

Statistical analyses. Statistical analysis was performed using SPSS v. 17.0 (SPSS Inc., Chicago, IL). Two-way ANOVA was used to test the effects of planting systems and extreme rainfall on the soil properties and genera

of nematode. One-way ANOVA was used to test the effects of planting systems on the soil properties, microbial biomass, nematode abundance, net nitrogen mineralization, microbial and nematode resistance index, neighbor species root biomass and leaf nitrogen content under control and extreme rainfall. One-way ANOVA was used to assess the effect of extreme rainfall on soil properties, microbial biomass, nematode abundance, net nitrogen mineralization, microbial and nematode resistance index, neighbor species root biomass and leaf nitrogen content under each planting system. Correlation analysis was used to assess soil biota resistance index and leaf $\mathrm{NH_4}^{+-}\mathrm{N}$ and $\mathrm{NO_3}^{--}\mathrm{N}$ contents. Least significant difference (LSD) was used to test differences among treatment means. Statistical significance was determined at P < 0.05.

References

- IPCC et al. Managing the risks of extreme events and disasters to advance climate change adaptation. In: Field, C. (Ed.), A Special Report of Working Groups I and II of the Intergovernmental Panel on Climate Change. Cambridge University Press, Cambridge, UK (2012).
- 2. Zhai, P. M., Zhang, X. B., Wan, H. & Pan, X. H. Trends in total precipitation and frequency of dailiy precipitation extremes over China. *J. Climate* 18, 1096–1108 (2005).
- 3. Sun, G. et al. Biotic and abiotic controls in determining exceedingly variable responses of ecosystem functions to extreme seasonal precipitation in a mesophytic alpine grassland. Agr. Forest. Meteorol. 228, 180–190 (2016).
- 4. Wagner, D., Eisenhauer, N. & Cesarz, S. Plant species richness does not attenuate responses of soil microbial and nematode communities to a flood event. *Soil Biol. Biochem.* 89, 135–149 (2015).
- 5. Wright, A. J. et al. Flooding disturbances increase resource availability and productivity but reduce stability in diverse plant communities. Nat. Commun. doi: 10.1038/ncomms7092 (2015).
- Orwin, K. H. & Wardle, D. A. New indices for quantifying the resistance and resilience of soil biota to exogenous disturbances. Soil Biol. Biochem. 36, 1907–1912 (2004).
- 7. Griffiths, B. S. & Philippot, L. Insights into the resistance and resilience of the soil microbial community. FEMS Microbiol Rev 37, 112–129 (2013).
- 8. Khan, M. A. S. A. *et al.* Climatic extremes lead to species-specific legume facilitation in an experimental temperate grassland. *Plant Soil* 379, 161–175 (2014).
- 9. Gellesch, E. *et al.* Plant community composition is a crucial factor for heath performance under precipitation extremes. *J. Veg. Sci.* **26.** 975–984 (2015).
- Grant, K., Kreyling, J., Heilmeier, H., Beierkuhnlein, C. & Jentsch, A. Extreme weather events and plant–plant interactions: shifts between competition and facilitation among grassland species in the face of drought and heavy rainfall. *Ecol. Res.* 29, 991–1001 (2014).
- Eskelinen, A. & Harrison, S. Biotic context and soil properties modulate native plant responses to enhanced rainfall. Ann. Bot. 116, 963–973 (2015).
- 12. Liancourt, P. et al. Plant response to climate change varies with topography, interactions with neighbors, and ecotype. Ecology 94, 444–453 (2013).
- 13. Niu, S. et al. Plant growth and mortality under climatic extremes: An overview. Environ. Exp. Bot. 98, 13-19 (2014).
- 14. Levine, J. M., McEachern, A. K. & Cowan, C. Do competitors modulate rare plant response to precipitation change? *Ecology* 91, 130–140 (2010).
- 15. Rivest, D., Lorente, M., Olivier, A. & Messier, C. Soil biochemical properties and microbial resilience in agroforestry systems: effects on wheat growth under controlled drought and flooding conditions. *Sci. Total. Environ.* **463**, 51–60 (2013).
- Zhao, J., Wang, X., Wang, X. & Fu, S. Legume-soil interactions: legume addition enhances the complexity of the soil food web. Plant Soil 385, 273–286 (2014).
- 17. Shao, Y. *et al.* Subordinate plants sustain the complexity and stability of soil micro-food webs in natural bamboo forest ecosystems. *J. Appl. Ecol.* **53**, 130–139 (2016).
- 18. Wardle, D. A. et al. Ecological linkages between aboveground and belowground biota. Science 304, 1629-1633 (2004).
- 19. Viketoft, M. *et al.* Long-term effects of plant diversity and composition on soil nematode communities in model grasslands. *Ecology* **90**, 90–99 (2009).
- Cesarz, S. et al. Tree species diversity versus tree species identity: Driving forces in structuring forest food webs as indicated by soil nematodes. Soil Biol. Biochem. 62, 36–45 (2013).
- 21. Unger, I. M., Kennedy, A. C. & Muzika, R. M. Flooding effects on soil microbial communities. Appl. Soil Ecol. 42, 1-8 (2009).
- 22. Mariotte, P., Le Bayon, R. C., Eisenhauer, N., Guenat, C. & Buttler, A. Subordinate plant species moderate drought effects on earthworm communities in grasslands. *Soil Biol. Biochem.* **96**, 119–127 (2016).
- 23. Taylor, A. R. & Wolters, V. W. Responses of oribatid mite communities to summer drought: The influence of litter type and quality. *Soil Biol. Biochem.* 37, 2117–2130 (2005).
- 24. Schoeneberger, M. et al. Branching out: Agroforestry as a climate change mitigation and adaptation tool for agriculture. J. Soil Water Conserv. 67, 128–136 (2012).
- 25. Li, H. Y., Pan, K. W., Liu, Q. & Wang, J. C. Effect of enhanced ultraviolet-B on allelopathic potential of *Zanthoxylum bungeanum*. Sci. Hortic. 119, 310–314 (2009).
- 26. Wang, J. Y. et al. Coupling effects of water and fertilizer on diurnal variation of photosynthesis of Zanthoxylum bungeanum Maxim 'hanyuan' seedling leaf. Acta Ecologica Sinica. 36, 1321–1330 (2016).
- 27. Zhao, J., Wang, X. L., Wang, X. L. & Fu, S. L. Legume-soil interactions: legume addition enhances the complexity of the soil food web. *Plant Soil* 385, 273–286.
- Zhao, J., Zeng, Z. X., He, X. Y., Chen, H. S. & Wang, K. L. Effects of monoculture and mixed culture of grass and legume forage species on soil microbial community structure under different levels of nitrogen fertilization. Eur J Soil Biol 68, 61–68 (2015).
- 29. Zhao, J., Wang, X., Shao, Y., Xu, G. & Fu, S. Effects of vegetation removal on soil properties and decomposer organisms. *Soil Biol. Biochem.* 43, 954–960 (2011).
- 30. Buena, A. P. et al. Use of pepper crop residues for the control of root-knot nematodes. Bioresource Technol. 98, 2846-2851 (2007).
- 31. Ferris, H., Bongers, T. & De Goede, R. A framework for soil food web diagnostics: extension of the nematode faunal analysis concept. *Appl. Soil Ecol.* **18**, 13–29 (2001).
- 32. Viketoft, M., Palmborg, C., Sohlenius, B., Huss-Danell, K. & Bengtsson, J. Plant species effects on soil nematode communities in experimental grasslands. *Appl Soil Ecol.* **30**, 90–103 (2005).
- 33. Papatheodorou, E. M. *et al.* Differential responses of structural and functional aspects of soil microbes and nematodes to abiotic and biotic modifications of the soil environment. *Appl. Soil Ecol.* **61**, 26–33 (2012).
- 34. Schimel, J., Balser, T. C. & Wallenstein, M. Microbial stress-response physiology and its implications for ecosystem function. *Ecology* **88**, 1386–1394 (2007).
- 35. Drenovsky, R. E., Vo, D., Graham, K. J. & Scow, K. M. Soil water content and organic carbon availability are major determinants of soil microbial community composition. *Microb. Ecol.* 48, 424–430 (2004).

- 36. Landesman, W. J. & Dighton, J. Response of soil microbial communities and the production of plant-available nitrogen to a two-year rainfall manipulation in the New Jersey Pinelands. Soil Biol. Biochem. 42, 1751–1758 (2010).
- 37. Sylvain, Z. A. et al. Soil animal responses to moisture availability are largely scale, not ecosystem dependent: insight from a cross-site study. Glob. Change Biol. 20, 2631–2643 (2014).
- 38. Nielsen, U. N. *et al.* Global-scale patterns of assemblage structure of soil nematodes in relation to climate and ecosystem properties. *Glob. Ecol. Biogeogr.* **23**, 968–978 (2014).
- 39. Chabrier, C. et al. Nematode dissemination by water leached in soil: Case study of Radopholus similis (Cobb) Thorne on nitisol under simulated rainfall. Appl. Soil Ecol. 40, 299–308 (2008).
- 40. Voesenek, L. & Bailey-Serres, J. Flooding tolerance: O₂ sensing and survival strategies. Curr Opin Plant Biol 16, 647-653 (2013).
- 41. Cesarz, S. et al. Nematode functional guilds, not trophic groups, reflect shifts in soil food webs and processes in response to interacting global change factors. Pedobiologia 58, 23–32 (2015).
- 42. Mccann, K. S. The diversity-stability debate. Nature 405, 228-233 (2000).
- 43. Kawase, M. Aerenchyma development in waterlogged plants. Am J Bot 67, 18-22 (1980).
- 44. Zhao, J. et al. Dicranopteris-dominated understory as major driver of intensive forest ecosystem in humid subtropical and tropical region. Soil Biol. Biochem. 49, 78–87 (2012).
- 45. Pirhofer-Walzl, K. *et al.* Nitrogen transfer from forage legumes to nine neighbouring plants in a multi-species grassland. *Plant Soil* **350,** 71–84 (2012).
- 46. IUSS Working Group WRB. World reference base forsoil resources 2006, first update 2007. World soilresources reports no. 103. FAO, Rome (2007).
- 47. Borken, W., Savage, K., Davidson, E. A. & Trumbore, S. E. Effects of experimental drought on soil respiration and radiocarbon efflux from a temperate forest soil. *Glob. Change Biol.* 12, 177–193 (2006).
- 48. Ng, E. L. *et al.* Do organic inputs alter resistance and resilience of soil microbial community to drying? *Soil Biol. Biochem.* **81**, 58–66 (2015)
- Bossio, D. & Scow, K. Impacts of carbon and flooding on soil microbial communities: phospholipid fatty acid profiles and substrate utilization patterns. *Microb. Ecol.* 35, 265–278 (1998).
- Frostegård, A., Tunlid, A. & Bååth, E. Changes in microbial community structure during long-term incubation in two soils experimentally contaminated with metals. Soil Biol. Biochem. 28, 55–63 (1996).
- 51. Townshend, J. A Modification and Evaluation of the Apparatus for the Oostenbrink Direct Cottonwool Filter Extraction Method. *Nematologica* 9, 106–110 (1963).
- 52. Yeates, G., Bongers, T., De Goede, R., Freckman, D. & Georgieva, S. Feeding habits in soil nematode families and genera—an outline for soil ecologists. *J. nematol.* 25, 315 (1993).
- 53. Tang, Z. C. The Experimental Guide of Modern Plant Physiology. Science Press, Shanghai, pp. 138-139, 154-157 (1999).

Acknowledgements

This study was sponsored by the National Natural Science Foundation of China (Grant Nos 31370632 and 31500517) and by the Ministry of Sciences and Technology of China (Grant No. 2015BAD07B050304). We thank associate prof. Yao Minjie for her valuable comments on this manuscript.

Author Contributions

K.P. and F.S. designed the experiments. F. S. performed the experiments. F.S. performed the instrumental analysis. F.S. analyzed the data, drew the figures and wrote the manuscript. K.P., A.T., Z.L., S.W., Q.X., D.S. and O.O. revised the manuscript. All authors reviewed the manuscript.

Additional Information

Competing financial interests: The authors declare no competing financial interests.

How to cite this article: Sun, F. et al. The response of the soil microbial food web to extreme rainfall under different plant systems. Sci. Rep. 6, 37662; doi: 10.1038/srep37662 (2016).

Publisher's note: Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit http://creativecommons.org/licenses/by/4.0/

© The Author(s) 2016