

## RIBOSOMAL DNA CONTENT AND BOBBED PHENOTYPE IN DROSOPHILA HYDEI

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### SUMMARY

The number of ribosomal RNA genes in different *Drosophila hydei* stocks has been determined by filter saturation hybridisation experiments. It has been shown that there is no marked correlation between the average rRNA gene number per cell in the whole animal and the bobbed phenotype when Y chromosomal nucleolar organisers are present.

### 1. INTRODUCTION

THE ribosomal RNA cistrons (rDNA) are clustered at the nucleolus organiser region (NO) (Ritossa and Spiegelman, 1965). In most of the *Drosophila* species, the NO's are localised on the sex chromosomes. In *D. melanogaster* it was shown that the NO is identical with the bobbed (*bb*) locus since a

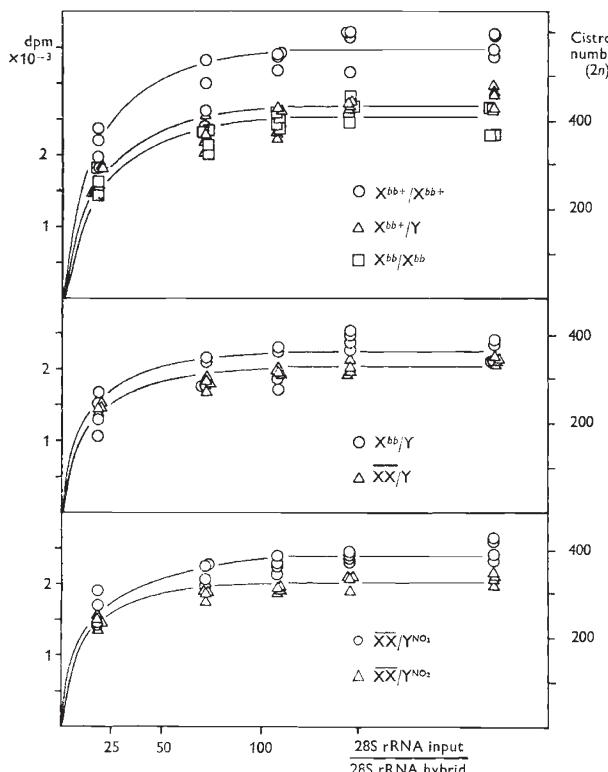


FIG. 1.—Saturation curves of DNA of different *D. hydei* stocks hybridised with 28S rRNA.

marked deficiency in rRNA genes is correlated with the *bb* phenotype (short, thin bristles) (Ritossa, Atwood and Spiegelman, 1966). A phenotypically similar *bb* mutation has been described in *D. hydei* (Clausen, 1923). In this species, the X chromosome has one NO while the Y has two separate NO's, one on the short arm ( $NO_1$ ), the second near the tip of the long arm ( $NO_2$ ) (Hennig, Link and Leoncini, 1975; Schäfer and Kunz, 1975). Since the attached-X ( $\overline{XX}$ ) chromosome used in our experiments lacks the NO containing heterochromatic arms, it is free of rDNA (Schäfer and Kunz, 1975; Kunz and Schäfer, 1976). A comparison of the total rDNA content of females and males shows that the ribosomal cistron number of the Y chromosome is smaller than that of the X (Hennig, 1968; Kunz and Schäfer, 1976; see also fig. 1). Therefore, genetic constitutions containing the  $\overline{XX}$  chromosome should have only small amounts of rDNA and are suitable for the study of the relation between the rDNA gene number of the *bb* phenotype.

## 2. MATERIALS AND METHODS

### (i) *D. hydei* stocks

Seven different genotypes were investigated.

(a) Females ( $X^{bb+}/X^{bb+}$ ) and males ( $X^{bb+}/Y$ ) from our laboratory wild type inbred stock.

(b) Females ( $X^{bb}/X^{bb}$ ) and males ( $X^{bb}/Y$ ) carrying a spontaneous *bb* mutation.

(c)  $\overline{XX}$ -females having either the whole Y chromosome ( $\overline{XX}/Y$ ) of one of two reciprocal Y fragments ( $\overline{XX}/Y^{NO_1}$  and  $\overline{XX}/Y^{NO_2}$ , respectively); for details of these stocks see Kunz and Schäfer (1976).

### (ii) Bristle index

To quantify the *bb* phenotype, we used the bristle index (B.I.) of Beck (1972) with a slight modification. B.I. is defined as the sum of the lengths of the two posterior scutellar setae divided by the distance between the two anterior scutellar setae. Twenty-five flies of each genotype were scored.

### (iii) DNA-RNA hybridisations

DNA extraction,  $^3\text{H}$  28S rRNA purification, and filter hybridisations were carried out as previously described (Kunz and Schäfer, 1976) except for some alterations in the hybridisation conditions: 4 hours incubation at 60°C in 50 per cent formamide and 6  $\times$  SSC. One DNA preparation was made for each genotype using whole adult flies, collected 1 to 3 days after emergence. Four different hybridisation experiments were carried out. In each of these experiments seven filters each loaded with the DNA of one genotype and two blank filters were incubated in varying arrangements within the same vial. The specific activity of the  $^3\text{H}$  28S rRNA was  $2.5 \times 10^5$  dpm/ $\mu\text{g}$ . For the calculation of the cistron numbers see Kunz and Schäfer (1976).

## 3. RESULTS

The wild type female ( $X^{bb+}/X^{bb+}$ ) has a B.I. of  $2.70 \pm 0.09$ , our  $X^{bb}/X^{bb}$  female a B.I. of  $1.97 \pm 0.09$  (fig. 2). DNA-RNA filter saturation hybridisa-

tions have shown that the  $bb+$  female has  $565 \pm 32$  rRNA cistrons whereas the homozygous  $bb$  female has a significantly lower rRNA gene number ( $411 \pm 27$ ). Thus, according to expectation, the  $bb$  phenotype in *D. hydei*, like that in *D. melanogaster*, is correlated with a lower rDNA content.

The wild type male ( $X^{bb+}/Y$ ) shows a bristle index which is clearly within the range of the wild phenotype ( $2.72 \pm 0.05$ ). Nevertheless, its rRNA gene number ( $434 \pm 35$ ) is as small as that of the  $bb$  female.

The  $X^{bb}/Y$  male has an rDNA content ( $365 \pm 39$  cistrons) which is similar to that of the  $X^{bb}/X^{bb}$  female ( $411 \pm 27$ ), but in spite of this low rRNA gene number its bristle index is significantly higher ( $2.44 \pm 0.11$ ). The index is somewhat smaller than the wild type B.I. (fig. 2).

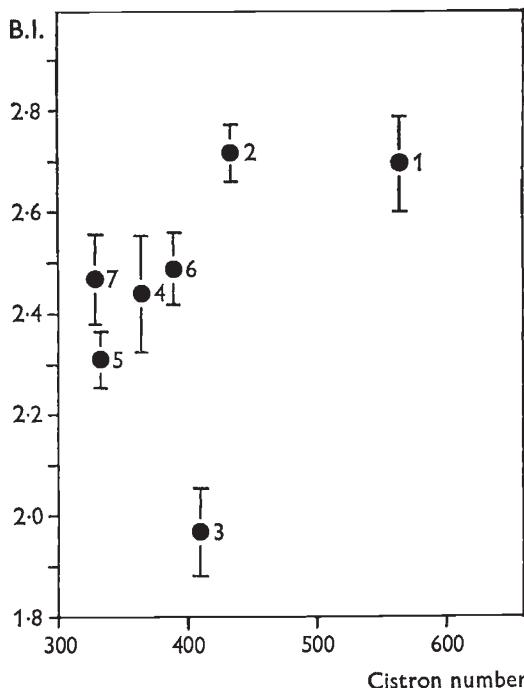


FIG. 2.—Plot of rRNA gene numbers relative to the bristle index. Genotypes:

- (1)  $X^{bb+}/X^{bb+}$ ; (2)  $X^{bb+}/Y$ ; (3)  $X^{bb}/X^{bb}$ ; (4)  $X^{bb}/Y$ ; (5)  $\overline{XX}/Y$ ; (6)  $\overline{XX}/Y^{NO_1}$ ;
- (7)  $\overline{XX}/Y^{NO_2}$ .

The  $\overline{XX}/Y$  female should possess only the rRNA gene number of a single wild type Y chromosome (152) because it has no X chromosomal NO. Due to the so-called gene compensation mechanism (Tartof, 1973; Kunz and Schäfer, 1976), however, its gene number is higher ( $332 \pm 14$ ). Nevertheless, this is distinctly lower than the rDNA content of the  $bb$  female. Yet, the bristle index ( $2.31 \pm 0.06$ ), as in the  $X^{bb}/Y$  male, falls between the wild type and the  $bb$  phenotype indices.

The rRNA gene numbers and bristle indices were also measured in two stocks where the  $\overline{XX}$  chromosome was combined with one half of a Y. Therefore, each of these two genotypes contained only one of the two Y

chromosomal NO's. Due to gene compensation, the rDNA contents of these stocks also are increased and were found to be  $390 \pm 24$  and  $329 \pm 16$  cistrons, respectively. These values, again, are not higher than the rRNA gene number of the *bb* female but their bristle indices are significantly higher ( $2.49 \pm 0.07$  and  $2.47 \pm 0.09$ ).

#### 4. DISCUSSION

Our data show that in the investigated *D. hydei* genotypes with Y chromosomal NO's there was no marked correlation between the bristle index and the rRNA gene number (fig. 2). This indicates either that the NO's of the Y chromosome react differently from that of the X or that there exist other factors besides the amount of rDNA influencing the *bb* expression. The latter conclusion is supported by the existence of an autosomal mutation which modifies the degree of bristle reduction in *bb* stocks of *D. hydei* (Beck, 1975).

Similar to our results in *D. hydei*, in *D. melanogaster* recently a genetic constitution has been described which had wild type bristle length but a very low rRNA gene number (Shermoen and Kiefer, 1975). The authors presented data which showed a direct relationship between the rate of rRNA accumulation and bristle length. This is in accord with the previous observations of Weinmann (1972). Therefore, the factor which determines the *bb* expression is possibly not the rRNA gene number but the rate of the rRNA synthesis. Another explanation could be that a specific degradation of the 28S rRNA causes the *bb* phenotype in spite of a high rRNA gene number (Marrakechi, 1974).

Also, females of *D. melanogaster* which had increased their rDNA by gene magnification were still bobbed (Tartof, 1973). In this case, it has been suggested that the additional rDNA is not functional. In *D. hydei* the situation seems to be different since our three  $\overline{XX}$  genotypes with a high percentage of compensated rDNA did not show the *bb* phenotype (fig. 2).

Our stocks where a low rDNA content was coupled with a relatively high bristle index, all contained intact NO's, whereas the *bb* female having a similar rDNA content accompanied by a low bristle index has undergone a deletion in the NO region. This observation supports the hypothesis (Marrakechi and Prud'homme, 1971; Shermoen and Kiefer, 1975) that the rate of rRNA synthesis is controlled by special segments in the rDNA which may be inactivated due to deletion.

On the other hand, it has to be considered that our gene number determinations were carried out on the DNA extracted from whole animals whereas the bristle development is cell autonomous. Since it has been shown that different tissues may vary in the number of their ribosomal RNA genes (Hennig and Meer, 1971; Spear and Gall, 1973; Renkawitz and Kunz, 1975) a correlation between the bristle length and the rDNA content may still exist even in genotypes with Y chromosomal NO's but it may be restricted to the bristle forming cells.

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