



# OPEN Preparation of high nutritional value biscuits from germinated chickpea, carrot, and turmeric rhizomes powder

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Legumes, being richer in protein, dietary fiber, vitamins, and minerals, offer a promising ingredient for enhancing the nutritional quality of baked products. This study examined the effect of germinated chickpea powder (GCPP) at different supplements of 10%, 20%, 30%, and 40% combined with turmeric rhizome powder (TRP) and carrot powder (CP), as substitutes for extracted wheat flour in biscuit formulation. The study prepared GCPP, added it to TRP and CP as substitutes for 72% extraction wheat flour (WF) in biscuit formulation, highlighting their nutraceutical properties. The composition of the biscuits was altered according to the specified ratios: Blends of 10%, 20%, 30%, and 40% (w/w) of GCPP/WF, with fixed additions of 5% of CP and 1% of TRP for all samples except the control (100% WF). A farinograph was used to record the dough's rheological characteristics, and biscuits' instrumental quality and nutritional value were recorded with assessment of sensory qualities by nutritionists from the Agricultural Research Center. Farinograph analysis shows significant increases ( $P \leq 0.05$ ) in water absorption, dough stability, dough development time, and softening degree. Based on physicochemical analysis, higher GCPP concentrations in the flour blends led to higher levels of protein, ash, fiber, and crude fiber contents, but with lower carbohydrates and caloric content. In addition, when GCPP, CP, and TRP were added to flour blends and biscuits samples, a significant increase ( $P \leq 0.05$ ) in the essential amino acid profile and biological value was revealed. The biscuits produced had a lower diameter and a lower spread ratio with increased hardness compared to purely wheat flour ones. Moreover, the total scoring of sensory evaluation was decreased. GCPF can be used as a WF substitute in bread goods that contain higher nutritional values and bioactive substances but have lower sensory appeal.

**Keywords** Chickpea, Carrot, Biscuits, Texture, Lysine, Sensory properties

With about 800 genera and over 20,000 species, the Fabaceae family, also referred to as “legumes” or “pulses,” is one of the largest plant groups. It encompasses a broad variety of plants, including grain, pasture, and agroforestry species, and includes dicotyledonous crops, which can be either annual or perennial. In addition to being an important and sustainable source of protein, dietary fiber, and phytochemicals, legumes have a variety of functions in the fields of medicine, economics, biodiversity, and environmental stability<sup>1</sup>.

Legumes are used as nutritional supplements or as partial replacements for wheat flour in baked goods because of their chemical composition<sup>2</sup>. Due to their dietary fiber content, research has shown that pulses have several health benefits, such as a lower glycemic index that is advantageous for diabetic patients, increased feelings of fullness, and possible protection against cancer and cardiovascular diseases<sup>3,4</sup>. Legumes are typically eaten as a cooked meal or as toasted dry seeds for snacking<sup>5</sup>.

Numerous studies have also demonstrated the advantages of replacing meat with plant-based foods high in protein for the environment, animal welfare, and human health, especially in low-income nations. The

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consumption of plant-based proteins from legumes is increasing on a global basis. They are healthier and more sustainable than animal-based proteins<sup>6</sup>.

Chickpea (*Cicer arietinum* L.) is the third most important legume globally, following soybean and common bean, based on total production. It is primarily cultivated in warm climates, particularly in countries such as India, Pakistan, Iran, Ethiopia, and the Mediterranean region<sup>7</sup>. The crop mainly comes in two types: *kabuli* and *desi*. *Kabuli* chickpeas are known for their large, smooth, beige seeds, whereas *desi* chickpeas feature smaller, rough-textured seeds with a darker outer coat<sup>8</sup> (Fig. 1).

Different chickpea cultivars possess unique physicochemical, nutritional, and functional properties. *Brown chickpeas* are ideal for vegan burgers, bread, and meat extenders, while *beige chickpeas* are better suited for other food products due to their enhanced functional properties. Both *desi* and *kabuli* varieties are rich in starch, protein, carbohydrates, lipids, fiber, flavonoids, and essential micronutrients such as zinc, iron, and manganese, and vitamin B12. These nutrients support the potential of chickpeas in managing chronic conditions such as diabetes and hyperlipidemia<sup>10</sup>.

Additionally, processing methods such as germination, cooking, or toasting can further improve protein digestibility<sup>11</sup>. The current trend is towards a diverse array of resilient and tolerant plants that thrive in challenging conditions, so they can offer superior nutritional benefits compared to conventional pulses. Kotsiou et al.<sup>12</sup> demonstrated that incorporating chickpea flour improved the quality of bakery items such as bread. Utilizing the nutritional potential of chickpeas to create nutrient-rich cookies shows promise in promoting improved health and nutrition<sup>13</sup>.

Carrot (*Daucus carota* L.) is the most important crop of the Apiaceae family. It is a root vegetable that can be found all over the world. Originally utilized for medicinal purposes, carrots were later employed as food. Carrots with purple and yellow flesh were the first to be cultivated. Orange carrots became more popular in Central Europe around the 15th and 16th centuries. As more people became aware of orange carrots' high provitamin A concentration, their popularity quickly increased<sup>14</sup>.

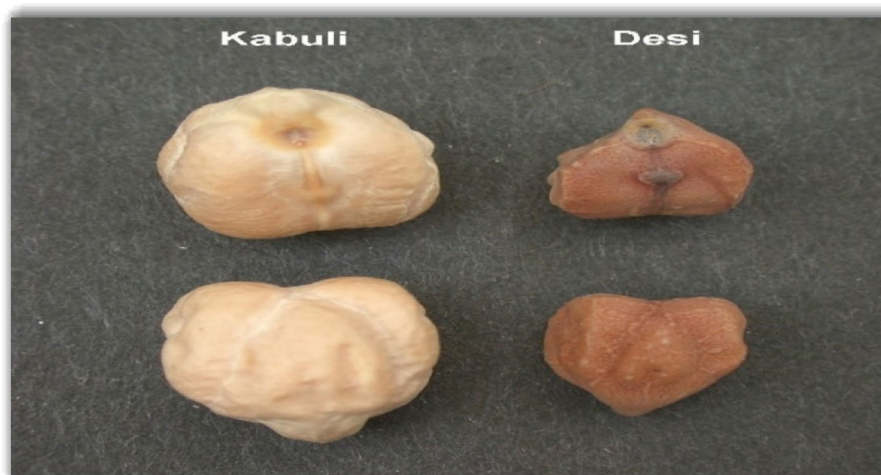
Carrot roots are a valuable source of essential vitamins and bioactive compounds that contribute to their nutritional quality. They are particularly rich in vitamin A ( $\beta$ -carotene), which plays a critical role in vision and increases immune system functions, and vitamin E, which aids cell signaling and membrane stability. Additionally, carrots provide B vitamins like thiamine, riboflavin, cobalamin, and pyridoxine, which are crucial for metabolism and brain health. Among organic acids, ascorbic acid (vitamin C) stands out for its antioxidant properties and roles in blood pressure regulation, iron absorption, and immune support<sup>15,16</sup>.

Carrots also contain benzoic and hydroxycinnamic acids, which possess antibacterial, anti-mutagenic, and anti-inflammatory benefits, and gallic acid, which inhibits mutagenesis<sup>17</sup>. Consequently, carrots can be incorporated as a functional ingredient in various products to enhance their biological and nutritional value<sup>18</sup>.

Turmeric, scientifically known as *Curcuma longa*, this golden spice is a perennial herb with rhizomes that has been employed in traditional medicine to address, prevent, and manage various health conditions, including cancer, diabetes, arthritis, diarrhea, inflammation, psoriasis, hepatobiliary disorders, and gastric and peptic ulcers<sup>19</sup>. In South Asia, it's also used in beauty rituals and as an anti-inflammatory and antibacterial agent<sup>20</sup>.

Turmeric contains polyphenols, alkaloids, diterpenes, sesquiterpenes, triterpenoids, and sterols, according to chemical analysis. The most researched ingredient in turmeric is curcumin, which makes up 2–5% of the plant. Turmeric's antibacterial, antimutagenic, anticancer, insecticidal, larvicidal, and radioprotective qualities have all been shown in laboratory experiments. Additionally, research on animals has demonstrated that turmeric is useful in reducing inflammation, atherosclerosis, cancer, diabetes, depression, and neurological illnesses<sup>21</sup>.

This innovation of using alternatives such as legume flours not only addresses the growing consumer demand for healthier snack options but also contributes to better digestive health, improved satiety, and supports public health initiatives focused on combating nutrient deficiencies.



**Fig. 1.** Chickpeas: Classes, types, and varieties<sup>9</sup>.

So, the aim of the study is To study the effect of replacing refined wheat flour with 10%, 20%, 30%, and 40% germinated chickpea powder combined with turmeric and carrot powders on the functional properties, dough characteristics, quality, sensory attributes, bioactive compounds, and nutritional value of biscuits.

## Materials and methods

### Collection of raw materials

The chickpea was acquired from the Agriculture Research Center in Giza. Wheat flour (72% extraction) was supplied by the North Cairo Flour Mills Company. Other ingredients, including turmeric rhizome powder (*Curcuma longa* L), carrot (*Daucus carota* L.), sugar (*sucrose*), fresh eggs, baking powder, vanilla, skimmed milk, and butter, were purchased from local markets. All ingredients were sourced from Egypt.

### Preparation of germinated chickpea flour

Approximately 2 kg of *kabuli chickpeas* underwent manual inspection to eliminate any impurities. Then to initiate germination, they were cleaned with distilled water, immersed for 12 h, positioned between two sheets of filter paper, and kept in a dark place at 30 °C for 48 h. The sprouted chickpeas were then dried for eight hours at 60 °C in a drying oven. A laboratory cutter mill was then used to grind the dry chickpeas into a fine powder, which was then filtered via a 60 µm mesh screen. The final powder was gathered and kept at 4 °C in an airtight container.

### Preparation of carrot powder

Carrot powder was obtained by the method described by Marvin<sup>22</sup> The carrot fruits were washed in portable water, peeled, sliced into 5–6 mm thickness; the sliced carrots were blanched for 3 min in hot water containing sodium metabisulfite to prevent discoloration. Then they were immediately cooled by exposure to air and dried in a cabinet drier at 50 °C for 12 h. The dried fruit was ground to a fine powder (model HL3294/C Phillips) and sieved with a 0.150 µm sieve and was packaged in a black polythene bag.

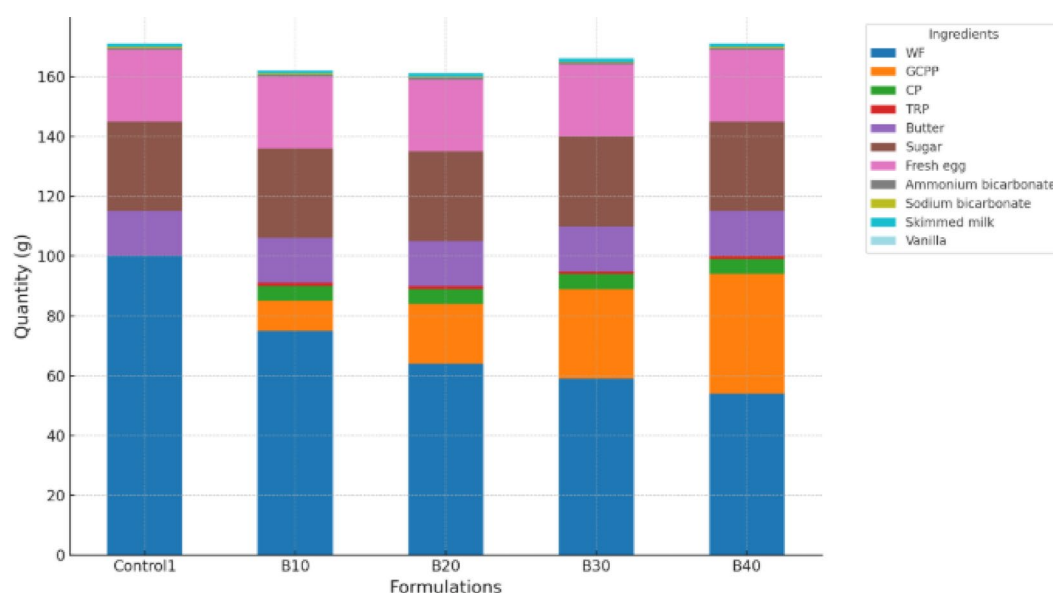
### Determination of chemical composition

The physico-chemical characteristics of both raw and processed legumes were evaluated using AACC<sup>23</sup> techniques. These analyses included measurements of protein concentration, fat content, dietary fiber, available carbohydrates, and mineral composition.

### Preparation of biscuits

Samples of biscuits were made following the standard protocol for semi-hard sweet biscuits utilized by Bisco Misr Company in Cairo, Egypt, as detailed in Fig. 2. Based on the method described by Mohamed et al.<sup>24</sup> this procedure incorporated certain modifications.

The biscuit manufacturing process employed the conventional creaming technique outlined by Chinma et al.<sup>25</sup> A Kenwood mixer (HM 430) was used to combine fat and sugar until the mixture achieved a fluffy consistency. Eggs and milk were then added while continuing to mix. The dough was formed by incorporating baking powder, ground nutmeg, composite flour, carrot powder, turmeric rhizome powder, and salt into the mixture. Once the dough was removed from the bowl, it was kneaded on a flat surface to achieve a consistent texture. It was then rolled out into thin sheets using a rolling pin and cut into the preferred shapes with a cutter.



**Fig. 2.** Ingredient Additions in Biscuit Preparation. WF= Wheat flour, GCPP= Germinated chickpea powder, CP= Carrot powder, TRP= Turmeric rhizomes powder.

The shaped pieces were arranged on a greased baking tray and baked at 180 °C for 17 min (as presented in Fig. 3). A control sample consisting of biscuits made exclusively with wheat flour was also prepared. Blends of 10%, 20%, 30%, and 40% (w/w) of GCPP/WF, with fixed additions of 5% and 1% of CP and TRP respectively, were used for all samples except the control (100% WF).

### Evaluation of biscuit characteristics

#### *Rheological properties of dough*

Farinograph measurements were used to evaluate how adding WF, GCPP, CP, and TRP affected the dough's mixing properties. The rheological characteristics of flour samples were assessed using a Brabender farinograph (mixing bowl 300 g, Brabender OHG, Duisburg, Germany HZ50), based on the AACC<sup>22</sup> procedure 54-21.02. Water absorption, dough development time, dough stability time, softening degree, and arrival time were among the Farinograph parameters that were determined by the analysis.

#### *Proximate composition of biscuits*

The proximate composition of biscuits provides essential information about their nutritional quality, including ashes, protein, fat, fiber, and carbohydrates. Understanding these components is important for assessing the overall quality, stability, and health impact of biscuits<sup>26</sup>.

#### *Amino acid profile of biscuit samples*

Amino acids of all samples were estimated according to the method suggested by AOAC<sup>27</sup>. The calorimetric method previously published by Miller<sup>28</sup> was used to assess the tryptophan content.

Using the formula given by Alsmeyer et al.<sup>29</sup> the computed protein efficiency ratio (C-PER) was calculated as follows:  $-0.4687 + 0.454 (\text{leucine}) - 0.105 (\text{tyrosine})$ .

The biological value (BV) was calculated using the formula  $BV = 49.9 + 10.53 \text{ C-PER}$ , as explained by Farag et al.<sup>30</sup>.

#### *Physical characteristics of biscuits*

The diameter (D) and thickness (T) of biscuits were measured for groups of 10 biscuits, following the procedure mentioned by AACC<sup>31</sup>. Then, the spread ratio was calculated as the ratio of diameter (D) to thickness (T). In addition, Texture profile analysis was done by A Brookfield CT3 Texture Analyzer No. M08-372-C0113 (version 2.1, 1000-gram unit). The hardness of samples was auto-recorded using TA-CT-PRO software. Following the method described by AACC<sup>22</sup>, samples underwent two compression cycles at 40% deformation with a trigger load of 5 N and a test speed of 2 mm/s. All procedures were done under ambient conditions.

#### *Sensory evaluation of the end product of biscuits*

The sensory characteristics of biscuits produced were evaluated by a panel of ten staff members from the Bread and Pastry Research Department, Agricultural Research Center, Giza. The evaluation followed the scoring scheme outlined by the AACC<sup>30</sup>, assessing the following attributes: crust color (20 points), taste (20 points), odor (20 points), general appearance (20 points), and texture (20 points), with a total possible score of 100.

### Statistical analysis of the data collected

The SPSS 26.0 program<sup>32</sup> was used to analyze the data from the experiments, which were recorded as means  $\pm$  standard deviations (SD) of triplicate measurements. Analysis of variance (ANOVA) was used to compare the means between groups, and Duncan's multiple range test was then performed to perform pairwise comparisons between the group means. The significance level was determined at a p-value of  $\leq 0.05$ .



**Fig. 3.** Preparation Steps for Biscuits. **A:** flour selection, **B:** dough formation, **C:** rolling, shaping, and tray arrangement.

Components	WF(72%extr) (g/100 g)	GCPP (g/100 g)	CP (g/100 g)	TRP (g/100 g)
Crude protein%	12.00 <sup>b</sup> ± 0.05	26.00 <sup>a</sup> ± 0.40	7.00 <sup>d</sup> ± 0.15	11.80 <sup>c</sup> ± 0.10
Ether extract%	1.85 <sup>c</sup> ± 0.03	7.50 <sup>a</sup> ± 0.01	1.80 <sup>d</sup> ± 0.05	6.70 <sup>b</sup> ± 0.05
Ash%	0.50 <sup>d</sup> ± 0.02	4.00 <sup>c</sup> ± 0.02	6.50 <sup>a</sup> ± 0.03	5.50 <sup>b</sup> ± 0.05
Crude fiber%	0.55 <sup>d</sup> ± 0.01	7.50 <sup>c</sup> ± 0.03	9.50 <sup>b</sup> ± 0.05	10.40 <sup>a</sup> ± 0.07
Available carbohydrates%	85.10 <sup>a</sup> ± 0.05	55.00 <sup>d</sup> ± 0.80	75.20 <sup>b</sup> ± 0.20	65.60 <sup>c</sup> ± 0.70
Caloric value (kcal/100g)	414.95 <sup>a</sup> ± 0.70	400.35 <sup>b</sup> ± 0.50	353.40 <sup>d</sup> ± 0.70	378.31 <sup>c</sup> ± 0.20
Minerals (mg/100 g)				
K	121.30 <sup>d</sup> ± 0.80	560.20 <sup>c</sup> ± 1.00	600.0 <sup>b</sup> ± 1.50	2600.0 <sup>a</sup> ± 2.00
Ca	18.00 <sup>d</sup> ± 0.05	170.45 <sup>c</sup> ± 0.50	690.00 <sup>a</sup> ± 0.90	195.00 <sup>b</sup> ± 0.50
P	140.90 <sup>d</sup> ± 0.20	240.60 <sup>b</sup> ± 0.30	180.00 <sup>c</sup> ± 0.30	270.00 <sup>a</sup> ± 0.90
Na	5.20 <sup>d</sup> ± 0.30	125.50 <sup>b</sup> ± 0.20	550.00 <sup>a</sup> ± 0.50	040.00 <sup>c</sup> ± 0.40
Mg	109 <sup>d</sup> ± 0.40	175.30 <sup>b</sup> ± 0.40	165 <sup>c</sup> ± 0.90	190.00 <sup>a</sup> ± 0.50
Fe	1.90 <sup>d</sup> ± 0.01	8.500 <sup>c</sup> ± 0.04	18.00 <sup>b</sup> ± 0.08	35.60 <sup>a</sup> ± 0.20
Mn	1.00 <sup>d</sup> ± 0.01	2.10 <sup>c</sup> ± 0.01	3.50 <sup>b</sup> ± 0.03	18.90 <sup>a</sup> ± 0.15
Zn	3.95 <sup>d</sup> ± 0.02	4.20 <sup>c</sup> ± 0.02	7.00 <sup>a</sup> ± 0.04	4.00 <sup>b</sup> ± 0.01
Vitamins(mg/100g)				
Vitamin C	Nd	27.05 <sup>b</sup> ± 0.05	45.50 <sup>a</sup> ± 0.40	1.5 <sup>c</sup> ± 0.01
B-carotene	1.30 <sup>d</sup> ± 0.01	9.07 <sup>b</sup> ± 0.04	120 <sup>a</sup> ± 0.03	6.5 <sup>c</sup> ± 0.06

**Table 1.** Chemical composition of Raw materials (On a dry weight basis). Means with different letters in the same row are significantly different at LSD at ( $p \leq 0.05$ ). Each value was an average of twenty determinations  $\pm$  standard deviation. nd = not detected. WF = Wheat flour, GCPP = Germinated chickpea powder, CP = Carrot powder, TRP = Turmeric rhizomes powder. .Potassium (K), phosphorus (P), magnesium (Mg), iron (Fe), and manganese (Mn), calcium (Ca), sodium (Na), and zinc (Zn).

Sample	Water absorption %	Arrival time (min)	Dough development Time (min)	Dough stability time (min)	Degree of softening (BU)
Wheat flour	59.50	1.5	1.5	9.0	60
B10%	61.45	2.0	2.5	10.0	65
B20%	65.50	3.0	3.5	11.5	70
B30%	67.30	4.5	4.5	12.5	80
B40%	76.90	5.0	5.5	13.0	90

**Table 2.** Farinograph parameters of dough.

Results and discussion  
Chemical composition of raw materials (on a dry weight basis)

Table 1 shows the results of the proximate analysis of WF, GCPP, CP, and TRP. Crude protein and ether extract contents were highest in GCPP (26.0% and 7.5%, respectively). Ash content was highest in CP (6.50%), while crude fiber content was highest in TRP (10.40%). Moreover, the available carbohydrate content and caloric value were highest in WF, at 85.10% and 414.95 kcal/100 g, respectively. These findings are in line with Saeed et al.<sup>33</sup>.

As regard mineral and vitamin composition of WF, GCPP, CP, and TRP, it was found that Potassium (K), phosphorus (P), magnesium (Mg), iron (Fe), and manganese (Mn) were highest in TRP, whereas calcium (Ca), sodium (Na), and zinc (Zn) were most abundant in CP. Additionally, CP exhibited the highest levels of ascorbic acid and  $\beta$ -carotene, at 45.5 mg and 120  $\mu$ g, respectively.

Rheological characteristics of dough

The farinograph parameters of WF and its blends with GCPP, CP and TRP are presented in Table 2.

It was observed that the water absorption of WF gradually increased as the level of substitution with GCPP increased. The higher fiber levels of GCPP, CP, and TRP than WF may be the cause of the dough's increased water absorption. These results are consistent with Abd El-Moniem and Yaseen<sup>34</sup> who revealed that increasing the amount of fiber sources added to WF resulted in the produced dough absorbing more water. This may be due to fiber's high water hydration capacity<sup>35</sup>.

The *dough development time* is the interval between adding water and when the dough reaches its maximum torque. The water hydrates the components of the flour during this mixing period. The current farinograph results revealed that adding GCPP, CP, and TRP lengthened the time needed for dough to develop; this could be because the plant sources delayed the hydration and development of gluten (because fibers and starches from plants “steal” water, so gluten needs more time to develop properly).



In addition, it was observed that the *stability time* of the control dough was 9.0 min, and that the addition of GCPP, CP, and TRP dough increased this time to approximately 10, 11.5, 12.5, and 13.0 min for B10%, B20%, B30%, and B40%, respectively. This stability time is a crucial indicator of dough strength based on the quantity and quality of dough gluten.

Proximate composition of biscuits

Table 3 shows that the replacement of WF with GCPP, CP, and TRP, with different supplemented ratios of B10%, B20%, B30%, and B40%, significantly improved the nutritional composition of the biscuits. *Protein content* increased from 9.97% in the control to a range from 10.21 to 13.77%, while *fat content* rose from 12.26% to 13.45–14.69%. Similarly, *ash content* increased significantly from 0.55% to 1.16–2.39%, and *crude fiber* content improved from 0.36% to 1.29–3.19%. In contrast, *carbohydrate content* decreased from 76.68% in the control to 65.96% in B40%. Although *energy content* was significantly higher in the control (467.52 kcal), it slightly decreased to a range of 467.21–460.57 kcal with the incorporation of GCPP, CP, and TRP.

These findings agree with Saeed et al.<sup>32</sup> who revealed that the increased contents of protein, crude fiber, ash, and fat enhance the nutritional profile of food products. Although there are numerous health benefits associated with consuming fiber, such as improved metabolic parameters, microbiota composition, and the synthesis of beneficial metabolites, Western countries continue to have low fiber intake. According to Kumar et al.<sup>36</sup> and Sreerama et al.<sup>37</sup> suggested that the food industry should focus on enhancing food products by increasing their fiber content, which presents an opportunity for food reformulation.

Amino acid profile of biscuits

The amino acid analysis presented in Table 4 revealed that biscuits supplemented with GCPP, CP, and TRP (B10%, B20%, B30%, B40%) had significantly ( $p \leq 0.05$ ) higher *essential amino acid* content compared to the control (WF-only biscuits), while the control sample contained higher levels of *non-essential amino acids*. The table also shows that the *C-PER* and *BV* were higher in biscuits prepared from GCPP, CP, and TRP-WF biscuits at different supplemented ratios than in the control sample. Our results are coordinate with Šramková et al.<sup>38</sup> who reported that wheat protein is notably low in tryptophan, threonine, and lysine, which limits its overall nutritional value.

Physical characteristics of biscuits

The dimensional characteristics of prepared biscuits with different levels of GCPP, CP, and TRP are represented in Table 5. The values of diameter and spread ratio decreased with the increased concentration of GCPP, CP, and TRP in biscuit samples.

The diameter of biscuits prepared with GCPP, CP, and TRP ranged from 58.50 to 53.33 mm, while their spread ratio varied from 6.16 to 4.19. In comparison, the control biscuits had a larger diameter of 60.50 mm and a higher spread ratio of 7.12. This finding might be due to the increased hydrophilic sites and water-soluble protein in GCPP, which compete for the limited free water in the dough, thereby increasing its viscosity<sup>39</sup> Moreover, the germination process leads to partial enzymatic degradation of starch, resulting in higher dextrin content in GCPP. This enhances the water absorption capacity of the dough and further reduces biscuit spread and diameter<sup>40</sup>. Similar results were also observed in biscuits made with roasted and germinated black gram flour<sup>41</sup> and germinated lupine flour<sup>42</sup>.

The table also displays the texture of biscuit samples to measure the hardness level, i.e., breakability, which was increased with the level of GCPP, CP, and TRP replacement. The hardness value of B10%, B 20%, B 30%, and B 40% was observed as 18.90, 20.30, 22.19, and 25.33, respectively, with GCPP 40% showing the highest value of hardness (25.33). This can be explained by the lower level of gluten content due to fat replacement with CP and GCPP, causing a weak gluten network, which affected the hardness of biscuits<sup>43</sup> and due to the higher fiber contents of GCPP, CP, and TRP<sup>44</sup>. Similar findings were reported by Saeed et al.<sup>45</sup> when black gram flour was used as a fat replacer in biscuits.

Blends	Protein %	Fat %	Ash %	Fiber %	Available carbohydrates%	Energy (kcal/100g)
Control	09.97 <sup>e</sup> ± 0.02	12.26 ± 0.05	0.55 <sup>c</sup> ± 0.01	0.36 <sup>c</sup> ± 0.01	76.86 <sup>a</sup> ± 0.05	467.57 <sup>a</sup> ± 0.05
B10%	10.21 <sup>d</sup> ± 0.05	13.45 ± 0.07	1.16 <sup>d</sup> ± 0.02	1.29 <sup>d</sup> ± 0.04	73.89 <sup>b</sup> ± 0.06	467.21 <sup>b</sup> ± 0.05
B20%	11.07 <sup>c</sup> ± 0.06	13.81 ± 0.04	1.55 <sup>c</sup> ± 0.03	1.89 <sup>c</sup> ± 0.01	71.68 <sup>c</sup> ± 0.06	464.95 <sup>c</sup> ± 0.07
B30%	12.20 <sup>b</sup> ± 0.09	14.25 ± 0.07	1.97 <sup>b</sup> ± 0.04	2.50 <sup>b</sup> ± 0.06	69.08 <sup>d</sup> ± 0.07	462.92 <sup>d</sup> ± 0.08
B40%	13.77 <sup>a</sup> ± 0.10	14.69 ± 0.03	2.39 <sup>a</sup> ± 0.05	3.19 <sup>a</sup> ± 0.05	65.96 <sup>e</sup> ± 0.05	460.57 <sup>e</sup> ± 0.09

**Table 3.** Proximate composition of Biscuits. Means with different letters in the same row are significantly different at LSD at ( $p \leq 0.05$ ). Each value was an average of three determinations ± standard deviation.

Amino acids	Control	B10%	B20%	B30%	B40%	FAO/WHO/UNU (1985) pattern
Lysine	2.82	3.31	3.80	4.29	4.77	5.80
Isoleucine	4.37	4.34	4.32	4.29	4.26	2.80
Leucine	6.73	6.80	6.86	6.93	7.00	6.60
Phenylalanine	4.90	4.99	5.08	5.17	5.26	6.30
Tyrosine	2.80	2.87	2.94	3.01	3.08	
Histidine	2.10	2.28	2.46	2.64	2.82	1.90
Valine	4.50	4.41	4.32	4.23	4.14	3.5
Threonine	2.80	2.87	2.94	3.01	3.12	3.40
Methionine	1.50	1.5	1.5	1.5	1.5	2.20
Tryptophan	1.30	1.3	1.3	1.3	1.3	1.00
Cysteine	2.00	1.93	1.86	1.79	1.72	
Total (EAA)	<b>35.82</b>	<b>36.6</b>	<b>37.38</b>	<b>38.16</b>	<b>38.97</b>	
Aspartic acid	5.46	6.14	6.83	7.51	8.20	
Glutamic acid	28.00	26.88	25.76	24.64	23.52	
Serine	4.70	4.68	4.66	4.64	4.62	
Proline	10.00	9.5	9	8.5	8	
Glycine	6.19	5.94	5.69	5.44	5.19	
Alanine	3.50	3.55	3.60	3.65	3.70	
Arginine	4.79	5.33	5.87	6.41	6.95	
Total (NEAA)	<b>62.64</b>	<b>62.02</b>	<b>61.41</b>	<b>60.71</b>	<b>60.18</b>	
C-PER	2.56	2.59	2.61	2.65	2.68	
BV	76.83	77.16	77.43	77.77	78.09	

**Table 4.** Amino acid profile of biscuits. EAA = (Essential Amino Acids) – NEAA = (Non-Essential Amino Acids). C-PER = Computed protein efficiency ratio. BV = Biological value.

Parameters	Control	B10%	B20%	B30%	B40%
Diameter(mm)	60.50 <sup>a</sup> ± 0.02	58.50 <sup>b</sup> ± 0.04	57.10 <sup>c</sup> ± 0.07	55.20 <sup>d</sup> ± 0.09	53.33 <sup>e</sup> ± 0.04
Thickness(mm)	8.50 <sup>c</sup> ± 0.05	9.50 <sup>d</sup> ± 0.09	10.50 <sup>e</sup> ± 0.07	11.90 <sup>b</sup> ± 0.08	12.80 <sup>a</sup> ± 0.07
Spread ratio	7.12 <sup>a</sup> ± 0.03	6.16 <sup>b</sup> ± 0.01	5.43 <sup>c</sup> ± 0.09	4.65 <sup>d</sup> ± 0.04	4.19 <sup>e</sup> ± 0.02
Hardness(N)	16.50 <sup>c</sup> ± 0.10	18.90 <sup>d</sup> ± 0.18	20.30 <sup>c</sup> ± 0.20	22.19 <sup>b</sup> ± 0.10	25.33 <sup>a</sup> ± 0.20

**Table 5.** Physical attributes of biscuits. Means with different letters in the same row are significantly different at LSD at ( $p \leq 0.05$ ). Each value was an average of three determinations ± standard deviation.

Samples No	Color 20	Taste 20	Odor 20	Appearance 20	Texture 20	Total Score 100
Control	19.50 <sup>a</sup> ± 0.20	19.10 <sup>a</sup> ± 0.25	19.20 <sup>a</sup> ± 0.15	19.30 <sup>a</sup> ± 0.15	19.40 <sup>a</sup> ± 0.15	96.50 <sup>a</sup> ± 0.20
B10%	19.30 <sup>a</sup> ± 0.30	19.00 <sup>a</sup> ± 0.30	18.90 <sup>b</sup> ± 0.20	19.15 <sup>a</sup> ± 0.20	18.70 <sup>b</sup> ± 0.10	95.05 <sup>b</sup> ± 0.15
B20%	17.20 <sup>b</sup> ± 0.25	18.40 <sup>b</sup> ± 0.15	18.10 <sup>c</sup> ± 0.10	18.20 <sup>b</sup> ± 0.15	17.20 <sup>c</sup> ± 0.10	89.10 <sup>c</sup> ± 0.40
B30%	16.15 <sup>c</sup> ± 0.30	17.50 <sup>c</sup> ± 0.30	17.50 <sup>d</sup> ± 0.15	17.15 <sup>c</sup> ± 0.40	16.13 <sup>d</sup> ± 0.35	84.43 <sup>d</sup> ± 0.10
B40%	15.15 <sup>d</sup> ± 0.40	16.33 <sup>d</sup> ± 0.35	17.20 <sup>c</sup> ± 0.10	16.33 <sup>d</sup> ± 0.20	15.80 <sup>e</sup> ± 0.30	80.81 <sup>e</sup> ± 0.20

**Table 6.** Sensory analysis of biscuits. Means with different letters in the same row are significantly different at LSD at ( $p \leq 0.05$ ). Each value was an average of twenty determinations ± standard deviation. Score 90–100 = very good, 80–90 = good, 70–80 = satisfactory, less than 70 = Questionable.

### Sensory analysis of biscuits

Table 6 depicts the effect of the incorporation of GCPP, CP, and TRP substitutes on the *sensory properties* of biscuit samples. The observations revealed that the biscuits prepared by substituting GCPP 10% such as control biscuits concerning color, taste, appearance, texture, and overall acceptability, while increasing the concentration of GCPP above 10% resulted in a significant decrease ( $P < 0.05$ ) in those sensory scores.

Color is a critical quality attribute in baked products, as it not only influences consumer appeal but also serves as an indicator of the formulation and processing conditions. In the current study, higher concentrations of GCPP 40% resulted in the darkest color of biscuit samples, which might be possible due to the increased rate of Millard reaction and caramelization process during baking, because of the high sugar and protein content. Color was illustrated in Fig. 4 measured by colorimeter that works by measuring the light reflected or transmitted by the biscuit at different wavelengths<sup>46</sup>.

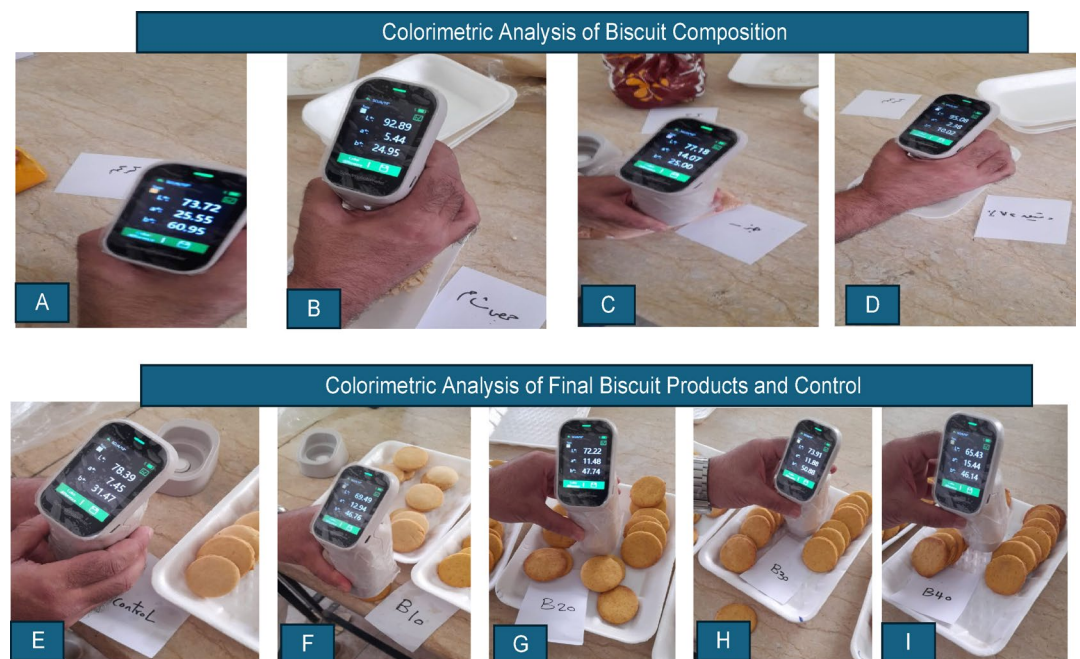
Biscuit samples prepared from GCPP 40% showed the lowest texture score of 15.80; the texture was affected by the level of fat and increased concentration of starch and non-starch compounds, i.e., fibers and proteins, and their resulting interactions with each other, which developed firm texture<sup>47,48</sup>. The color and appearance of the final product of different biscuits were illustrated in Fig. 5.

## Conclusion

This study's findings revealed that biscuits prepared from different proportions of germinated chickpea powder combined with carrot powder and turmeric rhizome powder, compared with biscuits made from wheat flour only, had a dough with higher water absorption, longer development, and stability time. The produced biscuits had higher crude protein, fat content, and crude fiber, while having a lower carbohydrate and caloric content. These biscuits were also rich in essential amino acids, with a higher computed protein efficiency ratio and biological value. But the overall sensory characteristics of these biscuits had lower scores comparable to those made with wheat flour.

## Recommendations

Considering the current findings, it is possible to produce high-quality baked goods using ingredients such as germinated chickpea powder, turmeric rhizome powder, and carrot powder. While the caloric difference between the control biscuit and the produced enriched biscuits is minimal, the enhanced nutritional and bioactive properties make these products beneficial as nutrient-dense snacks, especially for individuals at risk of malnutrition. Moreover, these biscuits may also serve as a healthy snack option for patients with obesity when consumed in a reasonable quantity. However, incorporating these functional ingredients may impact sensory



**Fig. 4.** Colorimetric Analysis of Biscuit Composition Ingredients and Final Products. Each measurement was taken using a portable colorimeter to evaluate L (lightness), a (red/green axis), and b (yellow/blue axis) values, providing quantitative assessment of ingredient and product color. **A:** Turmeric rhizomes powder, showing high b value indicating intense yellow pigmentation. **B:** Germinated chickpea powder, with the highest L value indicating lightness. **C:** Carrot powder, with moderate a (redness) and b (yellowness) values, contributing to the orange tone. **D:** Wheat flour, showing low a and b values and high lightness (L), as the base flour. **E:** Control biscuit (100% wheat flour), exhibiting a pale color profile with high L and low and high b values. **F:** Biscuit sample B10 (10% composite flour), with slightly reduced L and increased a and b value, indicating yellow color development. **G:** Biscuit sample B20 (20% composite flour), showing further reduction in L and increased a and b values, indicating enhanced color intensity. **H:** Biscuit sample B30 (30% composite flour), displaying reduced L and elevated a and more elevated b values, contributing to a deeper orange-brown tone. **I:** Biscuit sample B40 (40% composite flour), showing the lowest L, indicating the darkest and most intense coloration among all formulations.





**Fig. 5.** Images of the Final Biscuit Product vs. the control. Control biscuit:100% wheat flour. B10: 10% GCPP, 84% wheat flour, 5% carrot powder (CP), and 1% turmeric powder (TRP), B20 : 20% GCPP, 74% wheat flour, 5% CP, and 1% TRP, B30: 30% GCPP, 64% wheat flour, 5% CP, and 1% TRP, B40: 40% GCPP, 54% wheat flour, 5% CP, and 1% TRP.

attributes. Therefore, to improve overall acceptability, it is recommended to optimize their levels, including moisture-enhancing and flavor-masking agents, and adjust baking conditions to achieve desirable texture and color.

## Data availability

Data is provided within the manuscript.

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# Author contributions

MAA, GSE, AMH, and RAS": contributed equally to this article as follows conception, design of the work, analysis, method and statistical analysis, interpretation of data, have drafted the work and substantively revised it. All authors reviewed the manuscript.

## Declarations

### Competing interests

The authors declare no competing interests

### Ethics approval and consent to participate

This study did not involve the recruitment of human or animal subjects.

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