



OPEN Multivalent vaccine candidate from conserved immunogenic peptides in entry or exit proteins of *Orthopoxvirus* genus

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Orthopoxvirus (OPXV) genus includes emerging and re-emerging zoonotic viruses that pose threats to global health. Smallpox caused pandemics in the 20th century. Boreapox was responsible for a death in Alaska in 2024. Mpox, declared a Public Health Emergency by the WHO in 2022, with an alert reclassification in 2024. The lack of effective therapies and the limitations of attenuated virus vaccines, especially for immunocompromised individuals, reinforce the urgent need for new strategies to prevent diseases caused by pathogens of the OPXV genus. This study aimed to identify conserved epitopes in proteins essential for the entry and exit of these viruses and, based on this identification, develop a promising multivalent vaccine candidate. Viral protein sequences were extracted from the NCBI Virus database, and 160 sequences were analyzed to identify conserved epitopes using the Immune Epitope Database. After filtering the data, epitopes were concatenated to create a chimeric multi-epitope protein combined with β -defensin and PADRE adjuvants. The resulting protein, with eight conserved epitopes covering all OPXV viruses (including Mpox Clade Ib), was evaluated for antigenicity, allergenicity, and structural stability. It showed strong interaction with the TLR2 receptor, along with good predictions for immune responses after three doses. This proposed multivalent vaccine represents a potential approach against these zoonotic viruses, with promising results for in vitro and in vivo studies.

Keywords *Orthopoxvirus*, Reverse vaccinology, Multi-epitope, Immunoinformatics

Emerging viruses, such as SARS-CoV-2 and re-emerging ones, such as *Monkeypox* (Mpox), pose constant challenges to global health¹. While the former emerges unexpectedly and can cause sudden and devastating epidemics, the latter, although previously documented, resurface significantly, requiring continuous surveillance². Recently, the Mpox virus, belonging to the genus *Orthopoxvirus* (OPXV), has once again become an international concern³.

OPXV, part of the *Poxviridae* family, is one of the most significant viral genera for human health⁴. These viruses, characterized by their double-stranded DNA and complex structure, have a broad host spectrum, including various animal species, classifying them as zoonotic^{2,4}. Although many are named after the animals with which they are associated, it is believed that these animals primarily act as vectors rather than the original sources of the viruses⁴.

Currently, 16 species are recognized under the OPXV genus, including *Abatino*, *Akhmeta*, *Boreapox*, *Buffalopox*, *Camelpox*, *Cowpox*, *Ectromelia*, *Horsepox*, *Monkeypox*, *Rabbitpox*, *Raccoonpox*, *Skunkpox*, *Taterapox*, *Vaccinia*, *Variola*, and *Volepox*^{2,5}. In particular, the *Variola* virus (VARV), the causative agent of smallpox, was responsible for one of the most devastating pandemics of the 20th century, resulting in approximately 300 to 500 million deaths^{4,6}. Smallpox was officially eradicated in 1980, due to a global vaccination program^{2,7}. However, with the decline in immunization, the risk of OPXV transmission has grown, raising concerns about the morbidity of these zoonotic infections⁸.

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In recent years, outbreaks of OPXV infections have been recorded in various animals around the world. Cases of *Cowpox*, *Camelpox*, *Buffalopox*, and *Mpox* have been reported in Europe, the Middle East, India, South America, Africa, and the United States⁸. Moreover, new emerging viruses from this genus continue to be identified globally, such as the *Alaskapox virus* (recently renamed *Borealpox*), which caused the first death of an immunocompromised patient earlier this year⁹.

Among the viruses of the OPXV genus, *Mpox* has stood out due to its recent global impact³. Although it shares symptoms similar to smallpox, *Mpox* is distinguished by characteristic skin lesions¹⁰. Initially restricted to the African continent, *Mpox* was declared a Public Health Emergency of International Concern (PHEIC) by the World Health Organization (WHO) in 2022, following a significant increase in cases, including around 100,000 reported infections worldwide¹¹. In 2024, the WHO reaffirmed this global emergency due to the evolution of Clade Ib, which demonstrated higher transmissibility and the ability to spread to other continents³.

According to the Centers for Disease Control and Prevention (CDC), two vaccines are available for the prevention of infections caused by OPXV: *ACAM2000* and *JYNNEOS*². Both are licensed for use in at-risk individuals who have had contact with *Mpox*-infected persons and are being administered to contain current outbreaks². However, as they use attenuated viruses, these vaccines are contraindicated for immunocompromised individuals and other at-risk groups².

In this context, our research group previously developed an *in silico* multi-epitope vaccine candidate for *Mpox*, focusing on the viral entry and exit proteins in the host cell¹². Additionally, we identified three *Mpox* epitopes that are fully conserved in proteins similar to those of the *Borealpox virus*¹³. These findings led us to suggest that conserved epitopes might be present across the entire OPXV genus¹³.

Based on this hypothesis, the presence of conserved epitopes in the ten key proteins involved in OPXV entry and exit would enable the development of a next-generation vaccine candidate capable of targeting different regions of the viruses. This would potentially trigger a broad and specific immune response against viruses of the OPXV genus, even simultaneously. This approach represents a novelty that has not yet been reported in the literature for this genus.

The aim of this study is to identify conserved epitopes in ten key proteins involved in the viral entry and exit process across all 16 viruses belonging to the OPXV genus, with particular attention to the emerging *Mpox* Clade Ib sub-lineage. If these epitopes are identified, we intend to construct, develop, and validate an immunogenic multi-epitope chimeric vaccine for the OPXV genus, providing a broader and more effective approach to combating these viruses.

Materials and methods

Obtaining protein sequences from viruses of the OPXV genus

The proteins involved in the viral entry mechanisms into the host cell (A17L, A28L, A33R, H2R, L1R), exit (A27L, A35R, A36R, C19L), and in both processes (B5R) of all viruses of the OPXV genus were obtained from the National Center for Biotechnology Information Virus (NCBI Virus) database using the identifier “*taxid:10242*”^{12,14}. The *Mpox* virus Clade Ib sub-lineage sequences were obtained from the *PQ220056.1* sequence, deposited in the NCBI. To ensure the acquisition of proteins for all organisms, the selection followed this order: I—Selection of reference proteins (RefSeq); II—Selection of other proteins and generation of a consensus sequence when RefSeq were not found; III—BLAST for sequence similarity; IV—Proteins from the *PQ220056.1* sequence. If the protein was not found by these methods, it was determined to be absent.

Determination of consensus sequences and identification of epitopes for BCR, MHC-I, and MHC-II receptors

Consensus sequences for each protein, when necessary, were obtained using the Jalview software¹⁵. The RefSeq and consensus sequences were submitted to the Immune Epitope Database and Analysis Resource (IEDB) server for epitope identification¹⁶. For BCR epitopes, the BepiPred Linear Epitope Prediction 2.0 tool was used with a cutoff of 0.5. For MHC-I epitopes, the NetMHCpan EL 4.1 tool was used, selecting peptide options with sizes ranging from 8 to 14 amino acids and a reference set of 27 alleles, representing 97–99% of the global population. For MHC-II epitopes, the same tool was used, with peptides of 15 amino acids and a reference set of 7 HLA alleles. After obtaining the epitopes, the following exclusion criteria were applied: for MHC-I, a cutoff limit of 1% of epitopes interacting with each allele was established, ensuring an equal number of epitopes for each allele. For MHC-II, a consensus percentile cutoff of ≤ 10.0 was defined. For BCR, all found epitopes were selected. All cutoff thresholds followed the recommendations of the platform¹⁶.

Analysis of antigenic, allergenic, and physicochemical properties

For the antigenicity analysis, the VaxiJen 2.0 server was used, configured to recognize the target organism as a virus with a cutoff threshold of 0.5¹⁷. For allergenicity assessment, the AllerCatPro 2.0 server was used, following the default settings¹⁸. For the analysis of physicochemical properties, the ProtParam tool from the Biopython library was used, with a structural stability parameter set to < 40 .

Alignment, consensus epitope determination, and validation of epitope regions

The epitopes approved in all previous stages were aligned with the sequences of all viruses for each specific protein using the Jalview software, through the ClustalW plugin. Conserved regions for each protein were identified, defined as those with at least eight conserved amino acids in all sequences and containing at least one epitope aligned with the region. Conserved regions located in transmembrane areas and containing N-glycosylation sites were identified using the SOSUI¹⁹ server and NetNGlyc 1.0 server²⁰, respectively, and they were removed from the analysis.

Construction of a multi-epitope chimeric protein with conserved epitopes

Four multi-epitope chimeric proteins were created: I - containing the epitopes with the highest antigenicity value from each protein; II - with all conserved epitopes from the A17L protein; III - with all conserved epitopes from the H2R protein; and IV - with all conserved epitopes from the C19L protein. In the construction of chimeric protein I, the epitopes were arranged in sequence (entry, both, and exit). The other chimeric proteins were designed based on the increasing antigenicity value of the epitopes. To link the epitopes, the AAY linker was used to provide flexibility to the proteins^{12,13}. At the beginning of the proteins, two adjuvants, PADRE and beta-defensin, were added using the EAAAK linker to provide a more rigid characteristic to these adjuvants^{12,13}. At the end of the protein, a HIS-tag (HIS, 6x) was added.

Evaluation of the multi-epitope chimeric protein properties

Allergenicity, toxicity, and physicochemical properties were evaluated using the servers Vaxijen, AllerCatPro 2.0, and ProParam, respectively. The search for potential autoimmune responses was conducted by aligning the sequence of the multi-epitope chimeric protein with human and murine genomes using the BLASTp tool. The solubility analysis of the proteins was performed on the SoluProt 1.0 server, where proteins with values >0.5 are considered soluble²¹. The chimeric protein was submitted to the C-ImmSim server for immune response profile prediction using the following parameters: single injection, random seed = 12,345, injection without LPS, simulation volume of 10, and 300 simulation steps, with doses set at 1, 100, and 200 steps, representing days 0, 30, and 60 respectively, where both humoral and cellular responses were analyzed²².

Modeling, molecular docking, and molecular dynamics

For molecular modeling by homology and molecular docking, the AlphaFold Server was used, powered by the new AlphaFold3 and AlphaFold2 prediction model²³. The amino acid sequences of the multi-epitope chimeric proteins and the TLR2 receptor, available in the PDB under code 2Z7X, were provided to the tool. Additionally, ClusPro 2.0 was used to perform molecular docking between the multi-epitope protein and TLR2 receptor for comparative analysis. Molecular dynamics simulation was performed using GROMACS 2019.3 software with the OPLS-AA/L force field²⁴. The water equilibrium under NVT (Number of particles, Volume, and Temperature constant) and NPT (Number of particles, Pressure, and Temperature constant) conditions was maintained for 100 ps, at a temperature of 300 K and pressure of 1 bar. Subsequently, a 100 ns simulation was conducted, and RMSD values were generated by GROMACS²⁴.

Results

Protein profile of selected OPXV genus viruses

The proteins responsible for entry (A17L, A28L, A33R, H2R, L1R), exit (A27L, A35R, A36R, C19L), and both (B5R) of viruses from the OPXV genus were obtained in FASTA format from the NCBI Virus database. The “taxid:10242”, which covers all organisms of this genus, was selected. A total of 16 virus species were identified, including *Orthopoxvirus Abatino*, *Akhmeta virus*, *Borealpox virus*, *Buffalopox virus*, *Camelpox virus*, *Cowpox virus*, *Ectromelia virus*, *Horsepox virus*, *Monkeypox virus*, *Rabbitpox virus*, *Raccoonpox virus*, *Skunkpox virus*, *Taterapox virus*, *Vaccinia virus*, *Variola virus*, and *Volepox virus*.

Among these, nine have RefSeq for all studied proteins, four have RefSeq for nine of the ten proteins, necessitating the generation of a consensus for one protein. Three viruses do not have reference sequences; thus, the available proteins were collected, and a consensus was generated. It is important to note that *Rabbitpox virus* was the only one with a protein (A27L) found by BLAST, and Mpox virus had two RefSeqs for each protein, corresponding to clades I and II, except for A27L, for which a consensus sequence was generated.

For sequencing of the Mpox clade Ib virus, the PQ220056.1 sequencing was used, with the respective proteins aligned and identified. The A27L protein was not found and thus was marked as absent. All data are detailed in Table 1.

Antigenic, non-allergenic, and stable epitopes identified in conserved regions of eight proteins

The IEDB server was used to find epitopes that can bind to MHC-I, MHC-II, and BCR, with all parameters set by the server. A global consensus sequence was generated for each of the ten proteins from each virus. The epitopes were identified, collected, and subjected to antigenicity analysis on the Vaxijen server, and their stability was assessed using the ProtParam tool from the Biopython library. It was noted that there were more epitopes for MHC-I and MHC-II receptors compared to BCR (Table 2).

The obtained epitopes were aligned using Jalview software and the ClustalW tool with the proteins from the sixteen viruses to identify conserved epitopes across all proteins, i.e., conservation regions that had at least one epitope were selected (Table 2).

Proteins A28L and B5R, although having conserved regions, did not show the presence of epitopes in these regions or, when present, the epitopes did not fully occupy these regions, failing to meet the minimum length of 8 amino acids. A total of 26 epitopes were identified and none of them presented evidence of allergenicity using the AllerCatPro 2.0. The amino acid sequences of each conserved epitope along with their respective antigenicity values are presented in Table 3. It is worth noting that the identified conserved epitopes are also present in the new Mpox variant, clade Ib. Although proteins A17L, A36R, C19L, and L1R have mutations in their protein composition, the identified epitopes remain conserved in this new variant (Fig. 1).

The new Mpox sublineage, declared a global emergency by the WHO due to the evolution of Clade Ib, has demonstrated increased transmissibility and the ability to spread to other continents³. Therefore, it is crucial to verify if conserved epitopes remain preserved in this new variant. Sequencing conducted in 2024 on patients from the Democratic Republic of Congo regarding the new Clade Ib lineage is available on NCBI under code

Virus	Entry					Both	Exit			
	A17L	A28L	A33R	H2R	L1R	B5R	A27L	A35R	A36R	C19L
Abatino	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq
Akhmeta	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq
Borealpox	1	1	1	1	1	1	1	1	1	1
Buffalopox	Cons (4)	Cons (4)	Cons (6)	Cons (2)	1	Cons (5)	Cons (6)	Cons (6)	1	1
Camelpox	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq
Cowpox	Refseq	Cons (52)	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq
Ectromelia	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq
Horsepox	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Cons (3)	Refseq	Refseq	Refseq
Monkeypox*	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Cons (18)	Refseq	Refseq	Refseq
Mpox 1b	1	1	1	1	1	1	X	1	1	1
Rabbitpox	1	1	1	1	1	1	Blast	1	1	1
Raccoonpox	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq
Skunkpox	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq
Taterapox	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq
Vaccinia	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq
Variola	Refseq	Cons (19)	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq
Volepox	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq	Refseq

Table 1. Information on collecting protein sequences involved in the entry and exit mechanisms of viruses belonging to the OPXV genus. *Refseq* Reference sequence available in NCBI Virus, *Cons* Consensus sequence generated from the number of sequences (indicated in parentheses) present in NCBI Virus but not reference sequences, *X* No sequences found, *number* number of sequences available for a given protein. *Two Refseq were obtained for this virus.

Protein	BCR	MHC-I	MHC-II	Total	ProtParam	Vaxijen	Align
A17L	10	305	112	427	271	139	8
A28L	8	509	211	728	365	136	0
A33R	2	107	75	184	142	55	1
H2R	3	118	64	185	150	62	6
L1R	4	106	72	182	118	60	2
B5R	18	509	229	756	401	142	0
A27L	7	1.228	488	1.723	881	316	1
A35R	4	100	52	156	92	41	1
A36R	3	129	52	184	72	20	1
C19L	7	322	194	523	379	169	6

Table 2. Quantities of epitopes obtained at each filtering step.

PQ220056.¹²⁵, and it was used for analysis. The A27L protein was not found, which is consistent with the results in Table 1, where A27L is the only analyzed Mpox protein without a RefSeq. Additionally, among the eight proteins with conserved epitopes, A33R, A35R, and H2R are 100% conserved, ensuring epitope conservation (Fig. 1A). The A17L, A36R, C19L, and L1R proteins show 99.20%, 99.40%, 99.46%, and 99.34% conservation, respectively, compared to Mpox Clade I proteins; however, mutations do not occur in the identified epitopes (Fig. 1B–D).

Characterization and evaluation of the constructed multi-epitope chimeric proteins indicate adequate physicochemical properties, absence of allergenicity, good antigenicity, and positive humoral and cellular immune responses

After identifying the epitopes for each protein, four multi-epitope chimeric proteins were constructed. Two adjuvants were selected and added to the beginning of the chimeric protein: PADRE, a synthetic epitope capable of binding to MHC-II, which enhances the immune response by increasing the activation and proliferation of helper T cells, and β -defensin, which stimulates both the innate and adaptive immune responses¹². It is worth noting that the chimeric proteins related to A17L, C19L, and H2R were created individually as they were the only ones presenting more than one conserved epitope.

The chimeric protein constructs range from 146 to 173 amino acids, molecular weights varying from 16 to 19 kDa, and isoelectric points of approximately 9, indicating a basic nature for all proteins. Furthermore,

	Protein	Conserved region	Epitope	Vaxijen	Protparam
Entry	A17L	GIADIRDKYM	GIADIRDK	2.81	– 15.69
		YYYETSPGEIKPKFCLID	PGEIKPKF	1.97	3.36
		GEPCCSFKFRPG	CSSFKFRP	1.82	38.11
		RCHFIKKDYLLGDSDSA	CHFIKKDY	1.06	25.93
		YKTEHCDDFMTGFC	KTEHCDDF	0.85	38.05
		DPGNPNCLEW	GNPNCLEW	1.50	–3.77
		RPDYFTFGDTAL	FTFGDTAL	1.67	5.39
		GDKYLGPRVCWLHECTDESRDRKWLYNQDVQR	RDRKWLYY	1.27	30.18
	A28L	–	–	–	–
	A33R	SILNTRFLEKTSFYNC	LRFLEKTSF	1.07	30.29
	H2R	MDKTTLSVNACNLEYVREKAI	NLEYVREK	1.11	8.75
		YMRIKNTV	YMRIKNTV	1.04	33.02
		DWKSITDSKTKLES DRG	DWKSITDSK	1.20	30.29
		DFGAYFIAMRLD	DFGAYFIAMR	0.75	– 14.57
		KKAAKVDPQAQQFCQYLIKHKS	KKAAKVDP	1.41	– 20.65
		ITCGNEMLNELGYSYGF	LNELGYSYGF	1.14	– 9.98
	L1R	SLSDILQITQYLDLFLLLLIQSKNKLEA	QYLDLFLLL	1.61	– 10.10
		NKGYLFDFV	NKGYLFDFV	0.90	21.67
Both	B5R	–	–	–	–
Exit	A27L	DRYRNRVLLLTPE	RYRNRVLLL	1.28	– 4.57
	A35R	SLLSMITMSAFLIV	SLLSMITMSAFLIV	0.50	34.61
	A36R	PDTRHLRV	PDTRHLRV	1.04	28.69
	C19L	ETLPENMDFRSD	ETLPENMDFR	1.11	38.10
		NEIITLAKKIYI	NEIITLAKKIYI	1.28	32.83
		LLDERGKRNL	LLDERGKR	0.88	28.28
		SIDIEHLAIVPTTR	LAIVPTTR	2.04	22.21
		IYNSIIEAAINRGVKIRLLV	GVKIRLLV	0.61	– 5.46
		YVHITSANFDGTHYQNHGFVSFNS	YQNHGFVSFN	1.44	– 1.37

Table 3. Epitopes derived from conserved regions of entry and exit proteins: antigenicity and stability.

all proteins meet the cut-off thresholds for stability, antigenicity, allergenicity, solubility, and autoimmunity, indicating that they are suitable for vaccination (Table 4).

Subsequently, the simulation of the possible immune response to the four vaccine candidates was conducted on the C-ImmSim server. The multi-epitope protein containing epitopes from different proteins exhibited lower IgM + IgG titers compared to the vaccine candidates based on individual proteins (Fig. 2A). Additionally, the population of memory B cells was lower in the vaccine candidate containing epitopes from different proteins during the first 30 days, with an increase observed after the second dose administration. Comparing the cellular response, IFN- γ cytokine production was similar among the four vaccine candidates, while variations were observed in the levels of IL-2, TGF- β , IL-10, and IL-12 (Fig. 2A–D). All four vaccine candidates showed nearly null values for the “Danger” variable (Fig. 2A–D).

Structural stability and interaction with the TLR-2 receptor of the multivalent chimeric protein: modeling and molecular dynamics reveal good stability and fluctuation patterns

To determine the three-dimensional structure of the vaccine candidates and assess their interaction capability with the TLR-2 receptor, the AlphaFold 3 server was used. The predicted template modeling (pTM) score, interface predicted template modeling (ipTM) score, and per-residue measure of local confidence (pLDDT) were evaluated for each vaccine candidate. The modeling showed similarities, with TLR-2 exhibiting high pLDDT values, while the vaccine candidates showing low values. It was observed that the adjuvant region is responsible for anchoring to TLR-2 (Fig. 3A–D). Additionally, the pTM values ranged between 0.70 and 0.73, suggesting that the overall folds are close to the true structure, as values above 0.5 indicate good correspondence. However, ipTM values ranged from 0.20 to 0.22, indicating a potential failure in predicting the interaction. Therefore, validation of the structures through molecular dynamics was necessary.

Molecular dynamics was performed exclusively with the chimeric protein containing epitopes from different proteins due to its potential representativeness and comprehensiveness for immunization against the OPXV genus. Structural stability over time was assessed using the Root Mean Square Deviation (RMSD) graph, and the protein's residual fluctuation was analyzed with the Root Mean Square Fluctuation (RMSF) graph. The RMSD graph showed that the protein reached stability after 20 ns and remained stable until the end of the simulation (100 ns), with a structural variation from 0.4 nm to 0.5 nm (Fig. 3E). The RMSF graph indicated low variation in the initial part of the chimeric protein, with an increase in fluctuation occurring in the last amino acids of

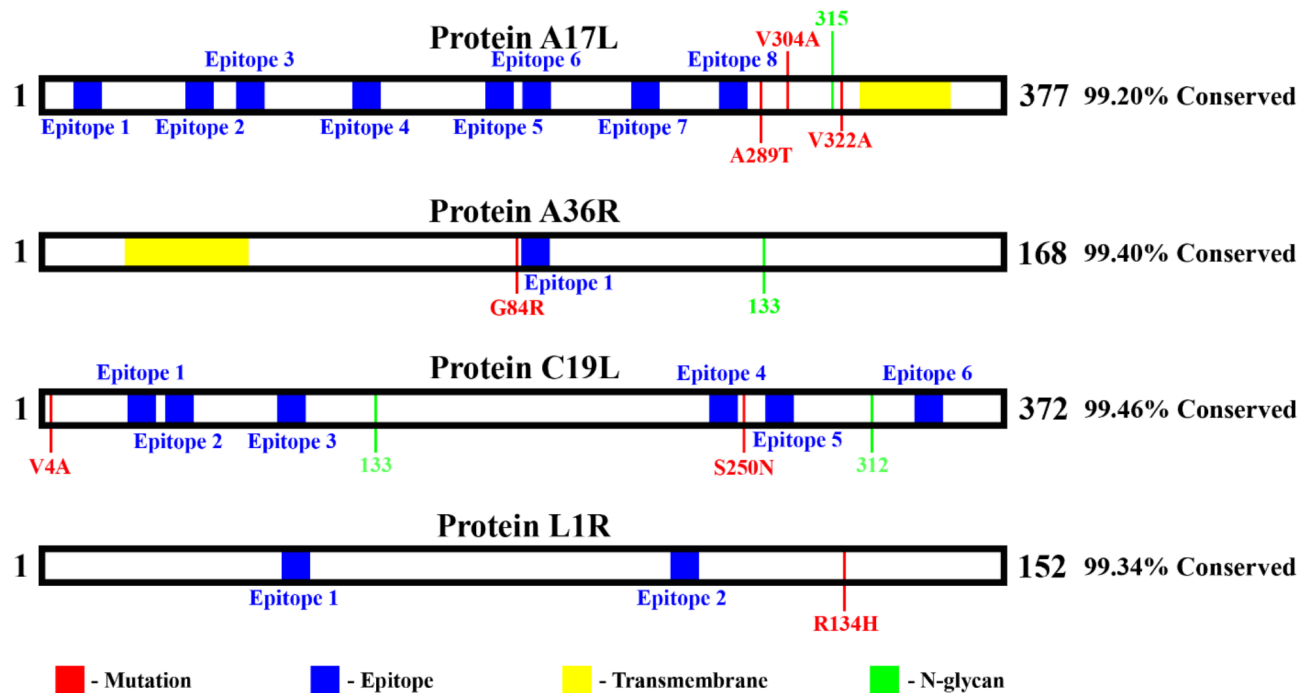


Fig. 1. Analysis of the A17L, A36R, C19L, and L1R proteins from the new Mpox virus lineage (Clade Ib), sequenced in 2024. The conservation percentages of these proteins compared to Clade I are indicated to the right of each protein. Conserved epitopes are highlighted in blue, mutations in red, transmembrane regions in yellow, and N-glycosylation sites in green. (A) Protein A17L with 99.20% conservation, displaying multiple conserved epitopes but with mutations outside these regions. (B) Protein A36R with 99.40% conservation, showing one mutation and one conserved epitope. (C) Protein C19L with 99.46% conservation, revealing several mutations and conserved epitopes. (D) Protein L1R with 99.34% conservation, having two conserved epitopes and one mutation outside these regions. The identified mutations do not affect the conserved epitopes, suggesting that they remain preserved in this new virus lineage.

Software	Info	Multi-epitope different proteins	Multi-epitope A17L	Multi-epitope C19L	Multi-epitope H2R
Protparam	Length (aa)	173	164	150	146
	Molecular weight (Da)	19217.64	18541.33	16829.60	16358.91
	Theoretical pI	9.99	9.44	9.81	9.70
	Instability index	21.21	23.87	25.11	27.71
	GRAVY	-0.052	-0.493	-0.299	-0.475
Vaxijen	Antigenicity	0.73	0.78	0.66	0.71
AllerCarPro	Allergenicity	No evidence	No evidence	No evidence	No evidence
SoluProt	Solubility	0.831	0.932	0.906	0.850
BLASTp	Autoimmunity	No evidence	No evidence	No evidence	No evidence

Table 4. Multi-epitope proteins properties.

the protein (Fig. 3F). In contrast, the TLR-2 receptor exhibited a characteristic fluctuation pattern, with two fluctuation peaks occurring between amino acids 200 and 300 (Fig. 3G).

Additionally, analyses of hydrogen bonds (H-bonds) and the radius of gyration (Rg) were conducted, as presented in Supplementary Fig. S1. The data show that the interaction complex between the TLR-2 receptor and the multi-epitope protein, composed of epitopes from different proteins, maintained between 450 and 550 hydrogen bonds throughout the simulation (Supplementary Fig. S1A). The radius of gyration ranged from 3.00 to 3.14 nm (Supplementary Fig. S1B), corroborating the RMSD values presented in Fig. 3E. These results indicate the preservation of structural integrity and stability of amino acid interactions during the 100 ns simulation, demonstrating the structural viability of the complex.

Discussion

The OPXV genus, with its 16 documented species and new emerging threats, continues to pose an increasing concern for global health^{2,3,5}. Vaccination is a crucial strategy to prevent these potentially fatal diseases, but current

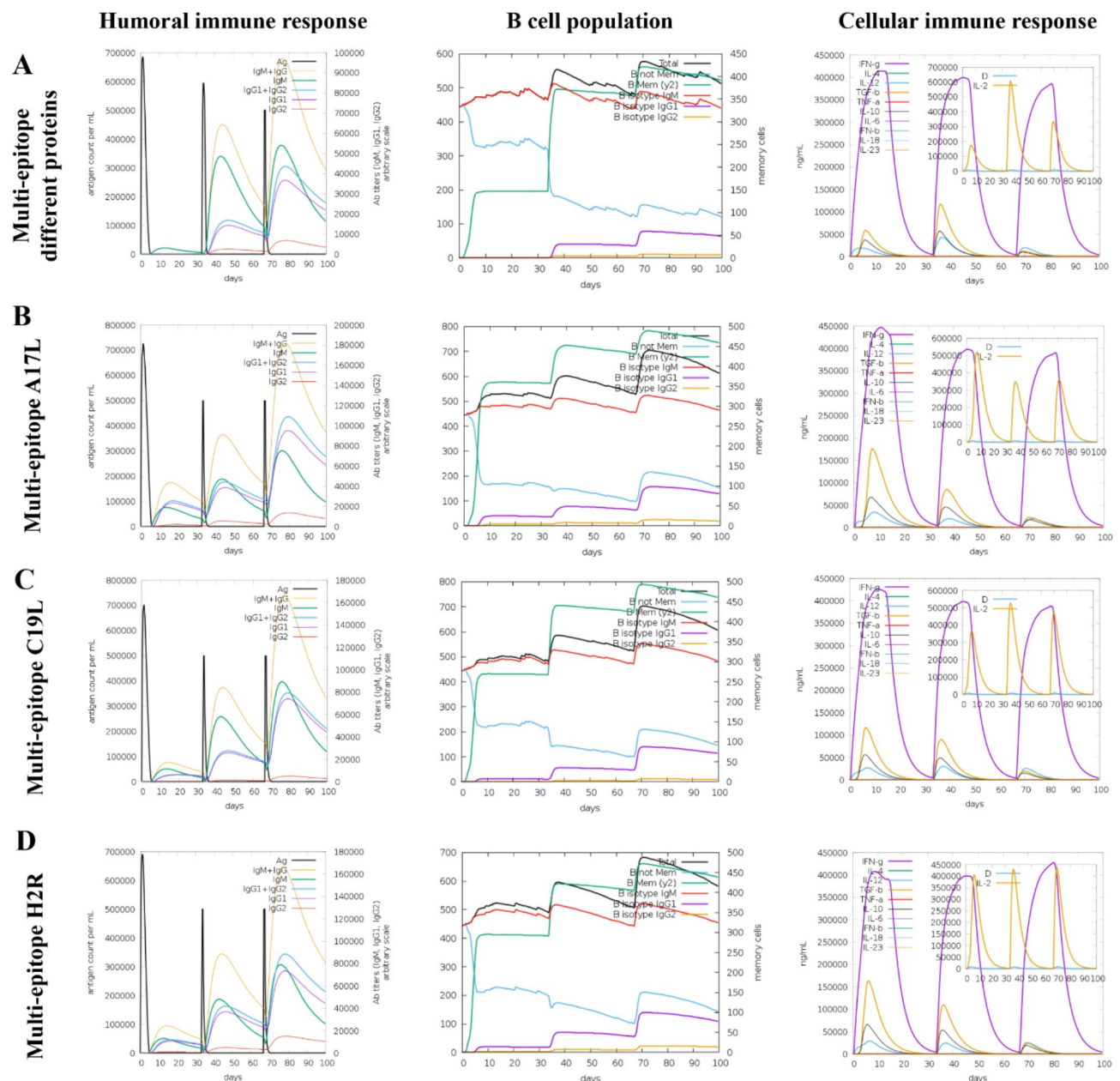


Fig. 2. The constructed multi-epitope chimeric proteins show satisfactory predictions for cellular and humoral responses after three doses. Cytokine and interleukin levels after 3 doses are displayed, with “D” in the graph indicating a potential danger signal. (A) Results for the chimeric protein containing epitopes from different proteins. (B) Protein with epitopes from A17L. (C) Protein with epitopes from C19L. (D) Protein with epitopes from H2R.

vaccines like *ACAM2000* and *JYNNEOS*, which use attenuated viruses, are unsuitable for immunocompromised patients, as reported by the CDC². Therefore, subunit vaccines, designed with computational techniques, offer a promising alternative to overcome these limitations and address genetic and antigenic variations¹².

In 2024, our research group reported the presence of immunogenic epitopes in the Mpox virus, developing an *in silico* multi-epitope vaccine candidate targeting viral entry and exit processes proteins¹². We also observed the conservation of these epitopes in *Boreapox virus* proteins, suggesting that conserved epitopes may exist throughout the OPXV genus¹³. In this study, we identified 26 conserved epitopes in eight proteins related to viral entry and release processes across the 16 OPXV species. These conserved epitopes can act as targets for neutralizing antibodies or induce a T-cell mediated cellular response^{12,13}.

OPXV viruses share similarities in viral entry and exit processes, making their proteins ideal targets for vaccine development²⁶. The A17L, a myristoylated protein, and A27L, an inclusion body type A protein, facilitate viral entry, while A28L, the membrane protein L1R, and the fusion complex H2R are essential for virus penetration into the cytoplasm²⁷. Being exposed during viral entry, these proteins are more susceptible to

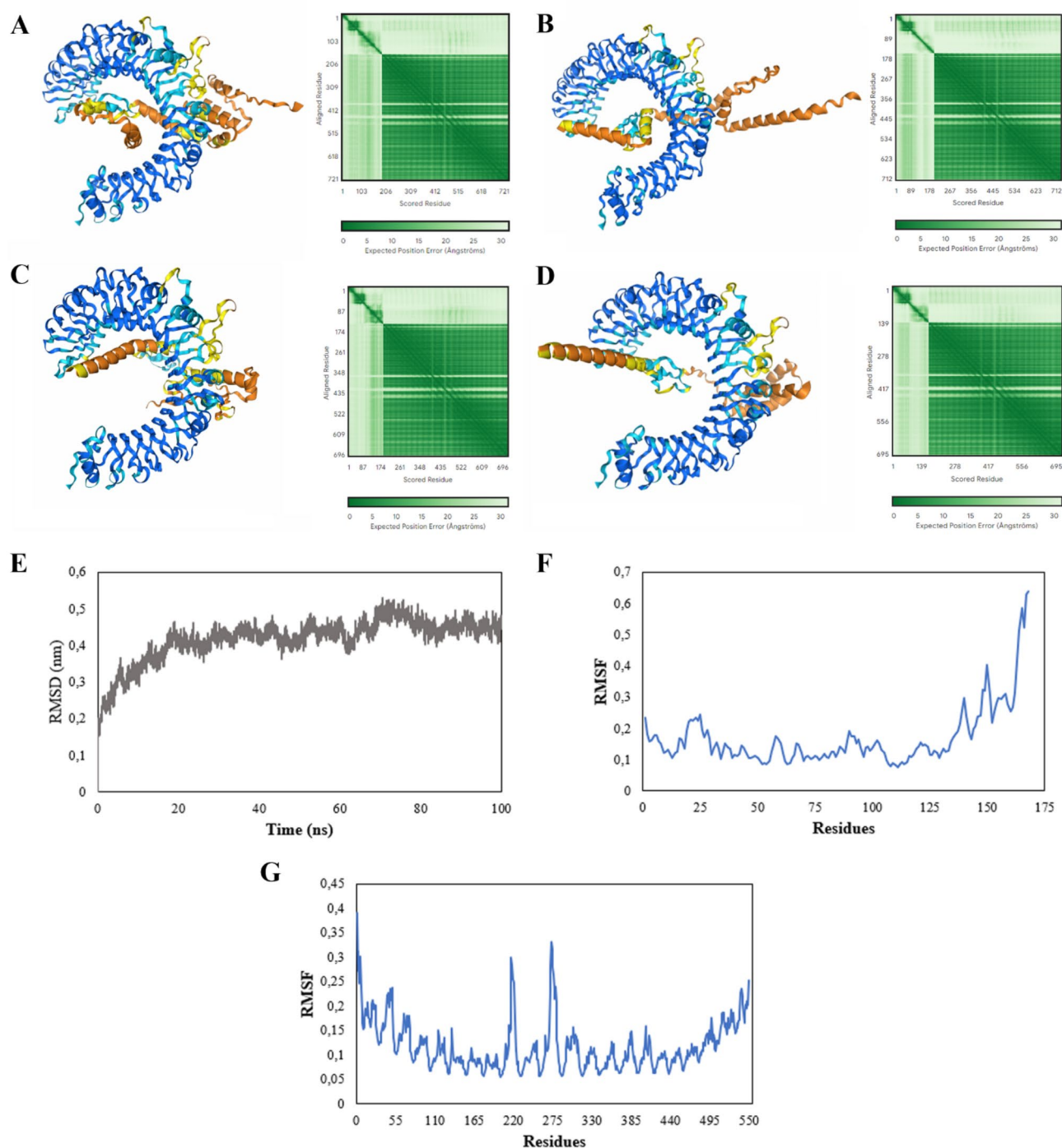


Fig. 3. The constructed chimeric proteins interact with the Toll-like receptor 2 (TLR-2), with modeling indicating good results for the receptor. However, the multiepitope proteins show low reliability. The multiepitope chimeric protein from different proteins demonstrates stability after 20 nanoseconds, maintaining a good interaction with TLR-2. **(A)** Modeling and binding of the multiepitope protein with TLR-2. **(B)** Binding of the A17L epitope protein with TLR-2. **(C)** Multiepitope from C19L and TLR-2. **(D)** Multiepitope from H2R and TLR-2. **(E)** RMSD obtained after 100 ns of molecular dynamics of the TLR-2 complex with the multiepitope protein. **(F)** RMSF of the multiepitope protein. **(G)** RMSF of TLR-2.

immune system recognition²⁶. Similarly, A33R, A35R, A36R, B5R, and C19L proteins, involved in the formation and maturation of extracellular viral particles (EEV), play a crucial role in virus dissemination²⁷ (Fig. 4).

The use of conserved epitopes, as well as their validation in *in vitro* and *in vivo* tests for other viruses like SARS-CoV-2 and hepatitis C, has been reported in the literature^{28–30}.

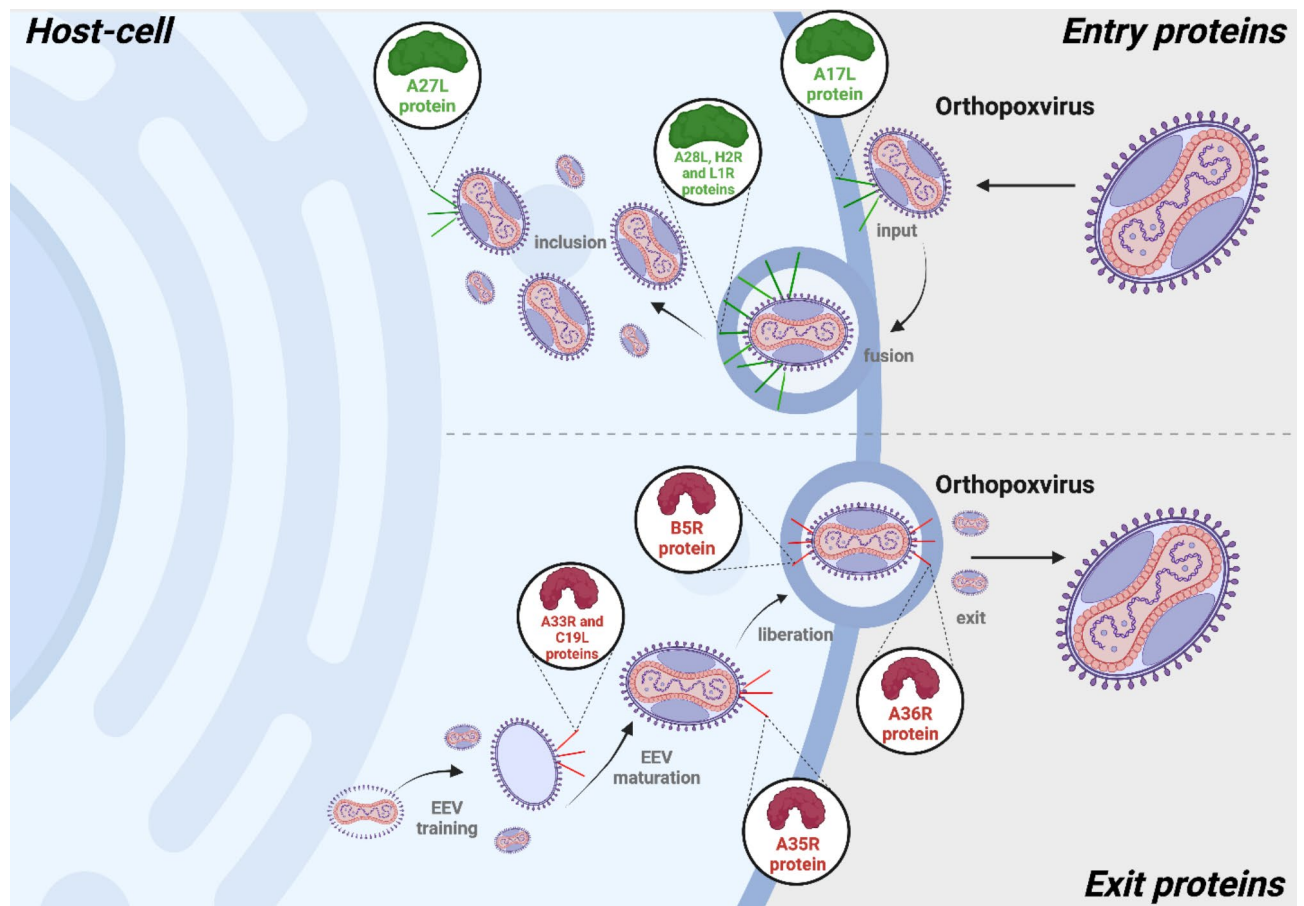


Fig. 4. Entry and exit proteins of *Orthopoxvirus* in the host cell. In the upper part of the figure, the A17L protein is involved in the virus's entry into the host cell; subsequently, the viral membrane proteins A28L, H2R, and L1R facilitate the fusion of the virus with the host cell, with A28L also enabling the formation of viral inclusions, assisting in the virus entry. In the lower part of the figure, the A33R and C19L proteins contribute to the formation of extracellular viral particles. The A35R protein is involved in the formation and maturation of extracellular enveloped virions (EEV); the B5R protein is essential for the formation of EEV and for the release of the virus; and finally, the A36R protein facilitates the virus's exit and the infection of neighboring cells. A17L Myristylate protein for entry, A27L A-type inclusion protein, A28L Truncated P4c or P4c/ATI inclusion factor, A33R CPXV166 protein, A35R EEV membrane phosphoglycoprotein, A36R IEV and EEV membrane glycoprotein, B5R Ankyrin-containing protein, C19L Palmytilated EEV membrane protein, H2R Entry-fusion complex essential component, L1R IMV membrane protein J1.

The OPXV conserved epitopes were combined with β -defensins and PADRE adjuvants into four vaccine candidates: one containing epitopes from multiple proteins, and others specific to A17L, C19L, and H2R. The use of the pipeline presented in this study reinforces the accuracy and reliability of the proposed approach, as it was also compared with a traditional workflow involving molecular modeling, molecular dynamics simulation, and molecular docking using previous-generation tools, such as AlphaFold2 and ClusPro 2.0¹². As shown in Supplementary Fig. 2, the AlphaFold3-based strategy not only reduces the number of steps required for multi-epitope vaccine development but also provides superior structural modeling quality when compared to AlphaFold2. Furthermore, AlphaFold3 accurately predicted the interaction between the vaccine construct and the TLR2 receptor, clearly illustrating the interaction of adjuvants with the receptor.

These chimeric proteins aim to induce local immune responses, including the production of inflammatory cytokines by macrophages^{12,13}. Captured by dendritic cells or macrophages, these antigens are presented on the surface by MHC molecules, allowing recognition by T-cell receptors and resulting in the generation of memory T-cells and the development of adaptive immunity^{7,8}. Additionally, the presence of B-cell specific epitopes is essential for antibody production, and a multi-epitope vaccine that interacts with both T-cell and B-cell receptors can effectively stimulate both humoral and cellular responses^{7,8,11}.

Immunological simulation revealed a decrease in IgM + IgG titration for the candidate with multiple protein epitopes compared to the multi-epitope protein developed for Mpox and individual protein candidates¹². This variation in humoral response may be attributed to differences in protein immunogenicity, as the focus of this study is on conserved epitopes rather than the most immunogenic of a protein. However, the cellular immune response, mediated by cytokines such as IFN- γ , TGF- β , IL-2, IL-10, and IL-12, remained similar, indicating a

consistent Th1 and regulatory profile. Furthermore, the robust cellular response suggests that both vaccines are effective in activating T-cells.

The results, introduce a new approach to vaccine development against OPXV genus viruses. The identification of conserved epitopes across different viruses demonstrates potential to induce robust humoral and cellular immune responses, even against the genetic diversity of these pathogens. We report the presence of conserved epitopes in eight OPXV genus proteins related to viral entry and exit processes in host cell.

As this study is based exclusively on in silico analyses, additional in vitro and in vivo studies are required to confirm the structural stability, immunogenic potential, and safety of the proposed multi-epitope vaccine candidate. Nevertheless, the structural, dynamic, and immunological data obtained strongly support the biological plausibility of the construct and provide a solid foundation for its future application not only as a prophylactic vaccine but also as a potential immunodiagnostic tool for all 16 *Orthopoxvirus* species.

Conclusion

It is evident that emerging and re-emerging viruses of the OPXV genus pose a persistent concern. Currently, vaccination is the most effective method against infections caused by this viral genus. In this study, we reported 26 conserved epitopes in key proteins and developed, as well as validated in silico, four multiepitope vaccine candidates. Analyses of antigenicity, allergenicity, structural stability, prediction of humoral and cellular immune responses, as well as the three-dimensional evaluation of interaction with the innate immune receptor TLR-2, indicated that the vaccine candidate with conserved epitopes from eight different proteins showed promising results.

This approach, combined with in vitro and in vivo testing, will be crucial for validating and refining the proposed vaccine candidates, strengthening the development of effective vaccines against the OPXV genus.

Data availability

The data supporting the findings of this study are available in the NCBI database under Taxid 10242 (https://www.ncbi.nlm.nih.gov/labs/virus/vssi/#/virus?SeqType_s=Nucleotide&VirusLineage_ss=Orthopoxvirus,%20taxid:10242) and in the PDB database under the code 2Z7X (<https://www.rcsb.org/structure/2Z7X>). Additionally, the following protein sequences were utilized in this study: Mpox virus Clade Ib sub-lineage: PQ220056.1.Protein A17L: YP_010085594.1, QED21147.1, NP_570524.1, NP_619932.1, NP_671638.1, YP_010509346.1, YP_010377125.1, NP_536554.1, YP_010085803.1, AAS49838.1, YP_009143444.1, YP_009282829.1, YP_717445.1, YP_233018.1, NP_042164.1, YP_009281883.1.Protein A27L: YP_010085604.1, QED21098.1, NP_570534.1, NP_619942.1, NP_671647.1, YP_010085812.1, YP_009282838.1, YP_717456.1, YP_233030.1, NP_042176.1, YP_009281892.1.Protein A28L: YP_010085605.1, QED21129.1, NP_570535.1, AUO16291.1, YP_010509355.1, YP_010377134.1, NP_536565.1, YP_010085813.1, AAS49847.1, YP_009143454.1, YP_009282839.1, YP_717457.1, YP_233031.1, YP_009281893.1.Protein A33R: YP_010085610.1, QED21259.1, NP_570540.1, NP_619950.1, NP_671652.1, YP_010509361.1, YP_010377140.1, NP_536570.1, YP_010085820.1, AAS49852.1, YP_009143460.1, YP_009282845.1, YP_717462.1, YP_233036.1, NP_042182.1, YP_009281899.1.Protein A35R: YP_010085610.1, QED21259.1, NP_570540.1, NP_619950.1, NP_671652.1, YP_010509361.1, YP_010377140.1, NP_536570.1, YP_010085820.1, AAS49852.1, YP_009143460.1, YP_009282845.1, YP_717462.1, YP_233036.1, NP_042182.1, YP_009281899.1.Protein A36R: YP_010085613.1, QED21233.1, AVO21161.1, NP_570543.1, NP_619953.1, NP_671655.1, YP_010509364.1, YP_010377143.1, NP_536573.1, YP_010085823.1, AAS49855.1, YP_009143463.1, YP_009282848.1, YP_717465.1, YP_233039.1, NP_042185.1, YP_009281902.1.Protein B5R: YP_010085641.1, QED21126.1, NP_570570.1, NP_619979.1, NP_671673.1, YP_010509386.1, YP_010377161.1, NP_536593.1, YP_010085850.1, AAS49879.1, YP_009143489.1, YP_009282875.1, YP_717494.1, YP_233068.1, NP_042218.1, YP_009281929.1.Protein C19L: YP_010085505.1, QED21148.1, NP_570438.1, NP_619848.1, NP_671554.1, YP_010509261.1, YP_010377040.1, NP_536472.1, YP_010085720.1, AAS49754.1, YP_009143359.1, YP_009282743.1, YP_717360.1, YP_232934.1, NP_042081.1, YP_009281797.1.Protein H2R: YP_010085556.1, QED21222.1, AVO21113.1, NP_570488.1, NP_619896.1, NP_671602.1, YP_010509310.1, YP_010377089.1, NP_536519.1, YP_010085768.1, AAS49802.1, YP_009143408.1, YP_009282793.1, YP_717409.1, YP_232982.1, NP_042129.1, YP_009281847.1.Protein L1R: YP_010085549.1, QED21242.1, NP_570481.1, NP_619889.1, NP_671595.1, YP_010509303.1, YP_010377082.1, NP_536512.1, YP_010085761.1, AAS49795.1, YP_009143401.1, YP_009282786.1, YP_717403.1, YP_232975.1, NP_042122.1, YP_009281840.1.

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Author contributions

L.P.D.A. conceptualization, data curation, formal analysis, investigation, methodology, validation and writing—original draft. E.N.S., S.M.D.A. and P.P.C. data curation, formal analysis, investigation and methodology. L.A.D.A. conceptualization, data curation, formal analysis, investigation, resources, supervision, validation, visualization and writing—original draft.

Declarations

Competing interests

The authors declare no competing interests.

Additional information

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