
Comparison of the properties of nanopaper from chitin nanofibers prepared by mechanical and TEMPO-oxidized methods

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1 **Title page**

2

3 **Comparison of the properties of nanopaper from**
4 **chitin nanofibers prepared by mechanical and**
5 **TEMPO-oxidized methods**

6

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23

24 **Abstract**

25 This study compares the structural, optical, and mechanical characteristics of chitin
26 nanopapers fabricated through mechanical fibrillation and TEMPO-mediated
27 oxidation. The TEMPO-oxidized nanopaper exhibited higher optical transparency
28 (approximately 92%) than the mechanically fibrillated sample (around 60%),
29 primarily due to enhanced nanofiber dispersion and smaller fibril diameters. In
30 contrast, the mechanically produced nanopaper showed greater crystallinity (above
31 90%) and stronger hydrogen bonding, resulting in higher tensile strength and
32 Young's modulus compared with the oxidized counterpart. Microscopic analyses
33 confirmed the more homogeneous and well-dispersed network in TEMPO-treated
34 samples, while spectroscopic results indicated the presence of carboxylate groups
35 introduced by oxidation. The research overall highlights the usability of both
36 nanopaper types in different applications and how TEMPO-oxidized nanopaper fits
37 best under transparent and biodegradable packaging and mechanically treated
38 nanopaper in applications requiring more strength.

39

40 **Keywords:**

41 Nanochitin, TEMPO-oxidation, Mechanical properties, Nanopapers, Optical
42 transparency.

51
52
53
54**55 1. Introduction**

56 The global seafood industry generates a substantial amount of waste,
57 particularly from the processing of crustaceans such as shrimp and crabs,
58 leading to significant environmental concerns ^{1,2}. Each year, millions of tons
59 of chitin-containing shellfish waste are discarded, mostly resulting in waste
60 and pollution ^{3,4}. In the sustainability and circular economy, chitin
61 valorization as a green and functional biomaterial is a viable path forward ⁵.
62 A more efficient application of chitin to produce high-sustainability products
63 not only reduces wastage of raw materials, but also adds to economic
64 production whilst maximizing the crude materials, turning waste into high-
65 grade materials ⁶⁻⁸. This method is in line with worldwide initiatives to
66 decrease waste and make greener alternatives a priority, and cements the
67 promise of chitin as a key material in future eco-friendly technologies ^{9,10}.

68 Chitin, the second most abundant biopolymer after cellulose in nature,
69 has drawn much attention for its excellent physicochemical properties such
70 as biocompatibility, biodegradability, and antimicrobial activity ³.

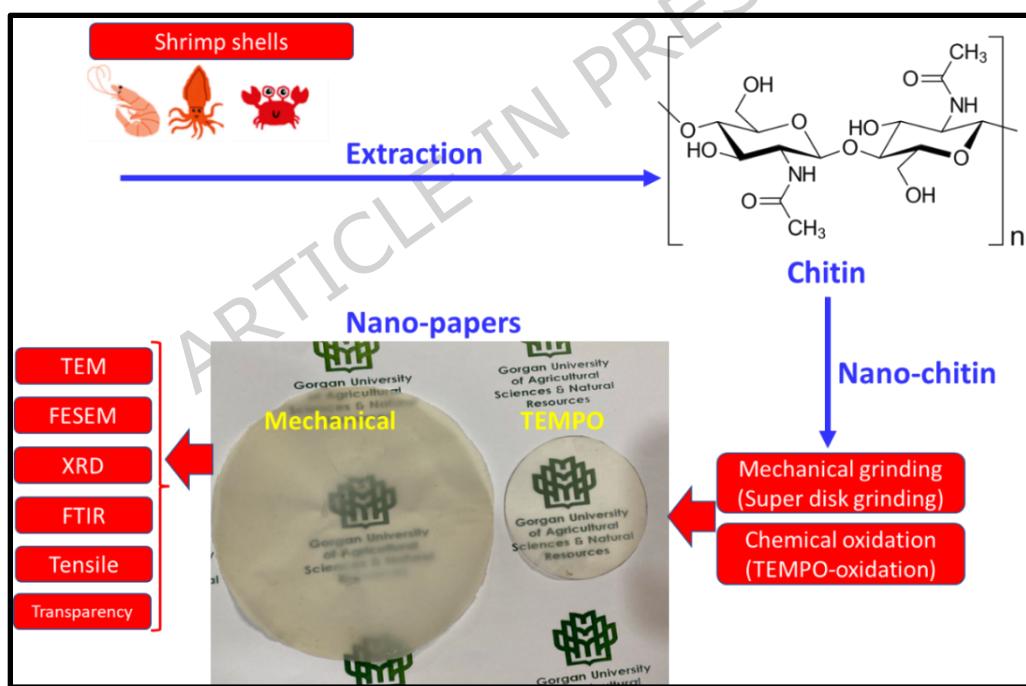
71 Chitin, derived from crustacean exoskeletons and fungal cell walls,
72 possesses a hierarchical structure that can be broken down into nanoscale
73 fibers using various processing strategies. Among these, top-down
74 approaches such as mechanical fibrillation ¹¹ and TEMPO (2,2,6,6-
75 tetramethylpiperidine-1-oxyl)-mediated oxidation ¹² are the most widely
76 used for producing chitin nanofibers. Unlike these methods, electrospinning
77 is primarily a down-top approach technique applied to chitin or its
78 derivatives in solution form and is less common for producing individual
79 nanofibrils from bulk chitin ¹³. Mechanical processing can serve to physically
80 separate the chitin fibrils in the absence of significant chemical
81 transformation ¹¹, whilst TEMPO oxidation leads to the introduction of
82 carboxylated groups at the surface of the nanofibers, which give rise to
83 altered dispersion characteristics, charge properties, and inherent reactivity
84 ^{12,14-16}.

85 Nanopaper is a high-density, flexible sheet made up of interwoven
86 nanofibers that has become an attractive, sustainable alternative to regular
87 paper because of its enhanced mechanical strength and gas barrier
88 characteristics, along with adjustable surface chemistry ¹⁷⁻¹⁹. The properties
89 of chitin nanopaper greatly depend on the particular processing route used
90 to extract the nanofibers ^{11,20}. The chitin nanofiber obtained through
91 mechanical disintegration maintains the original chitin structure, with a
92 reduced surface charge and a higher intrinsic crystallinity. Conversely,
93 TEMPO-oxidized chitin nanofiber has more dispersed and higher colloidal
94 stability due to their negatively charged surfaces, which can considerably
95 influence the formation, density, and mechanical integrity of the nanopaper.
96 The extent to which these processing methods influence the final chitin

97 nanopaper properties remains an unanswered question and must be
 98 examined through a comprehensive analysis.

99 Although extensive research has been conducted on chitin nanofibrils
 100 obtained through mechanical fibrillation and TEMPO-mediated oxidation, the
 101 underlying mechanisms linking these processing routes to the resultant
 102 nanopaper properties remain insufficiently understood. The novelty of this
 103 work lies in the systematic, side-by-side comparison of these two major top-
 104 down approaches using identical chitin sources under controlled conditions
 105 to isolate the influence of surface chemistry, fibril morphology, and inter-
 106 fibrillar interactions on the mechanical strength, porosity, and water
 107 resistance of the resulting nanopapers. Unlike previous studies that
 108 examined either mechanical disintegration or chemical oxidation
 109 independently, this study integrates both processes (Fig. 1) within a unified
 110 experimental framework and provides quantitative correlations between
 111 processing parameters and functional performance. Furthermore, by
 112 combining morphological, physicochemical, and barrier analyses, this
 113 research establishes a clearer structure property relationship that advances
 114 the rational design of chitin-based nanopapers for sustainable packaging and
 115 coating applications.

116



117
 118 **Fig. 1.** Schematic of the research steps in this study.
 119

120 **2. Materials and methods**

121 **2.1. Raw materials**

122 Chitin nanofibers with different surface morphologies were produced by
 123 using mechanical grinding and a chemical oxidation (TEMPO-oxidation)
 124 process. The mechanical treatment was performed using bleached dry chitin
 125 (extracted from shrimp shells, supplied by Nano Novin Polymer Co. (Gorgan,

126 Iran), 50 g of which was ground 3 times in a super disk grinding (Masuko
 127 MKCA6-2, Japan) to obtain well-dispersed chitin nanofibers at the speed of
 128 rotation of 1800 rpm. In the chemical process, 0.16 g of TEMPO was
 129 dissolved in 1 L of deionized water and stirred for 24 h; thereupon, 1.0 g of
 130 sodium bromide was added, and the stirring continued for 1 h. Then, 10 g of
 131 bleached dry chitin was added to the solution, and then sodium hypochlorite
 132 was slowly added until the pH was brought to the value of 10. To ensure the
 133 oxidation reaction proceeds continuously, reducing the processing error, the
 134 pH was maintained at 10-11 for 6 h. The obtained gel was washed with
 135 deionized water 3 times (500 mesh polyester filter bags to eliminate residual
 136 chemicals). It was finally passed through the disk grinding to get uniform
 137 TEMPO-oxidized chitin nanofibers.

138

139 **2.2. Nanopaper preparation**

140 For the preparation of 60 g/m² chitin nanopaper samples, an appropriate
 141 amount of each chitin nanofiber gel, including mechanically processed and
 142 TEMPO-oxidized variants, was weighed separately. For uniform dispersion,
 143 each suspension was stirred for 15 min at room temperature with a
 144 magnetic stir plate set at 250 rpm. A prepared suspension was poured onto a
 145 vacuum filtration system equipped with a 500-mesh (12 cm diameter)
 146 polyester filter. Removing water from the suspension with a vacuum
 147 pressure of 0.5 MPa resulted in the generation of a primary wet film. After
 148 dewatering, the as-prepared films were subjected to drying in a vacuum
 149 oven at 70 °C for 15 h between two pieces of glass plates to ensure
 150 sediment formation of nanopaper and provide enough structural stability.

151

152 **2.3. Characterization**

153 **2.3.1. Transmission electron microscope (TEM)**

154 The specimens were vacuum-dried and gold-coated before TEM examination.
 155 The nanopaper specimens were investigated by a TEM (CM 120) with an
 156 accelerating voltage of between 1.5 and 5 kV. Additionally, the average
 157 diameter of 100 fibers was determined utilizing digitizer image software (v.
 158 4.1.1.0).

159

160 **2.3.2. Field emission scanning electron microscopy (FESEM)**

161 FESEM was used to evaluate the sample morphology. The samples were
 162 coated with a layer of platinum <0.2 nm thick, in a vacuum environment
 163 using a sputter coater machine. Finally, the coated samples were
 164 characterized by FESEM (Zeiss Sigma 300-HV, Germany) using an
 165 accelerating voltage of 5.0 kV.

166

167 **2.3.3. X-ray diffraction (XRD)**

168 X-ray Diffraction (XRD) is a widely utilized method for assessing the
 169 crystallinity index ²¹. In this study, an XRD diffractometer (D8-Advance
 170 Bruker Cu K α 1, Germany) was employed to expose the samples to Cu-K α
 171 radiation under operating conditions of 50 kV and 30 mA. The X-ray analysis

172 was conducted with a step size of 0.02° , and the scanning range was set
 173 between 10° and 60° (2θ). The crystallinity index of samples was calculated
 174 using Eq. 1.

175

$$176 \text{ CrI (\%)} = [(I_{200} - I_{\text{am}}) / I_{200}] \times 100$$

177 (1)

178 where I_{200} was the maximum intensity of the [200] lattice diffraction, which is typically in the
 179 range $2\theta = 21^\circ\text{--}23^\circ$ and I_{am} was the intensity diffraction at $2\theta = 18^\circ$

180

181 **2.3.4. Fourier transform infrared (FTIR) spectroscopy**

182 FTIR is an analytical tool used to determine the adventitious or attributable
 183 chemistry of materials by the way the materials absorb IR light at their
 184 distances. Films were investigated for their functional groups and the
 185 chemical change features by FTIR spectrometers (Perkin-Elmer, Spectrum RX
 186 I). Spectra were acquired from 4000 to 500 cm^{-1} (64 scans at 4 cm^{-1}
 187 resolution) ²².

188

189 **2.3.5. Optical transparency**

190 Optical transparency was evaluated through both quantitative and
 191 qualitative methods using a double-beam UV-vis spectrophotometer (U-
 192 2000, Hitachi Ltd., Japan) for precise measurements and a digital camera for
 193 visual assessment.

194

195

196

197 **2.3.6. Mechanical properties**

198 The tensile properties were evaluated following the ASTM D882-18 standard
 199 using a SANTAM universal tensile machine (model STM-1, Santam Co.,
 200 Tehran, Iran) equipped with a 1 kN load cell and a cross-head speed of 10
 201 mm/min . Samples were prepared in standard dimensions and secured
 202 between the tensile grips. Before testing, three specimens from each sample
 203 were conditioned at 30°C and 50% relative humidity for 24 h . The reported
 204 tensile parameters included tensile strength and elongation at break.

205

206 **2.4. Statistical analysis**

207 All measurements were performed in triplicate, and the results are reported
 208 as mean \pm standard deviation ($n = 3$). Statistical variability was evaluated
 209 using descriptive statistics, and error bars representing standard deviation
 210 were included in the corresponding figures to ensure data reliability.

211

212 **3. Results and discussion**

213 **3.1. Morphology**

214 The TEM micrographs and the corresponding diameter distributions of
 215 nanochitin obtained from mechanically processed chitin nanofibers and
 216 TEMPO-oxidized processes are presented in Fig. 2. The TEM micrographs are
 217 primarily used to illustrate the overall fibrillar morphology and network
 218 structure rather than to provide high-precision measurements of individual

219 fibril diameters. As shown in Fig. 2a, the mechanically processed nanofibers
220 form an entangled and partially aggregated network, within which individual
221 fibrils are not always clearly distinguishable. Accordingly, the reported
222 average diameter of 26.04 ± 8.57 nm is derived from representative
223 measurable regions, while acknowledging the inherent uncertainty caused
224 by fibril overlapping and limited contrast at this magnification. This behavior
225 can be attributed to the disk grinding process and the absence of sufficient
226 electrostatic repulsion to achieve complete fibril separation, resulting in non-
227 uniformity and aggregation consistent with the previous report²³. In
228 contrast, Fig. 2b demonstrates that TEMPO-oxidized chitin nanofibers exhibit
229 a more homogeneous and finer fibrillar structure, with an average diameter
230 of 7.72 ± 2.23 nm, reflecting the effectiveness of surface oxidation in
231 promoting fibril individualization, while it is emphasized that quantitative
232 diameter values are interpreted cautiously and supported by statistical
233 analysis, and the TEM observations mainly serve to qualitatively confirm the
234 morphological differences and fibrillar network characteristics between the
235 two processing routes in accordance with the resolving capability of the
236 presented images. Although TEMPO-mediated oxidation generally enhances
237 fibril dispersion due to the introduction of negatively charged carboxylate
238 groups, some degree of aggregation can still occur. This is primarily
239 attributed to charge screening and partial re-association of nanofibrils during
240 the drying stage, especially when residual counterions (e.g., Na^+) and
241 hydrogen bonding forces reduce electrostatic repulsion between oxidized
242 fibrils. Moreover, an excessively high degree of oxidation may lead to
243 localized fibril damage, generating shorter fragments that tend to cluster
244 through secondary interactions.

245 FESEM micrographs show that the morphology of the nanostructures used
246 is in the form of fibers, so they are one-dimensional nanostructures; that is,
247 two of their dimensions are in the nanoscale and one of their dimensions
248 (length) is in the non-nanometric scale (more than 5 μm) (Fig. 3). The
249 TEMPO-oxidized nanofiber structure, owing to the generation of carboxylate
250 groups on its surface, is highly dispersed and uniform, offering greater clarity
251 and homogeneity compared to mechanically processed nanofibers;
252 moreover, the reduced diameter of the TEMPO-oxidized nanofibers results in
253 a higher specific surface area, which is particularly significant for
254 applications such as drug delivery and biosensors.

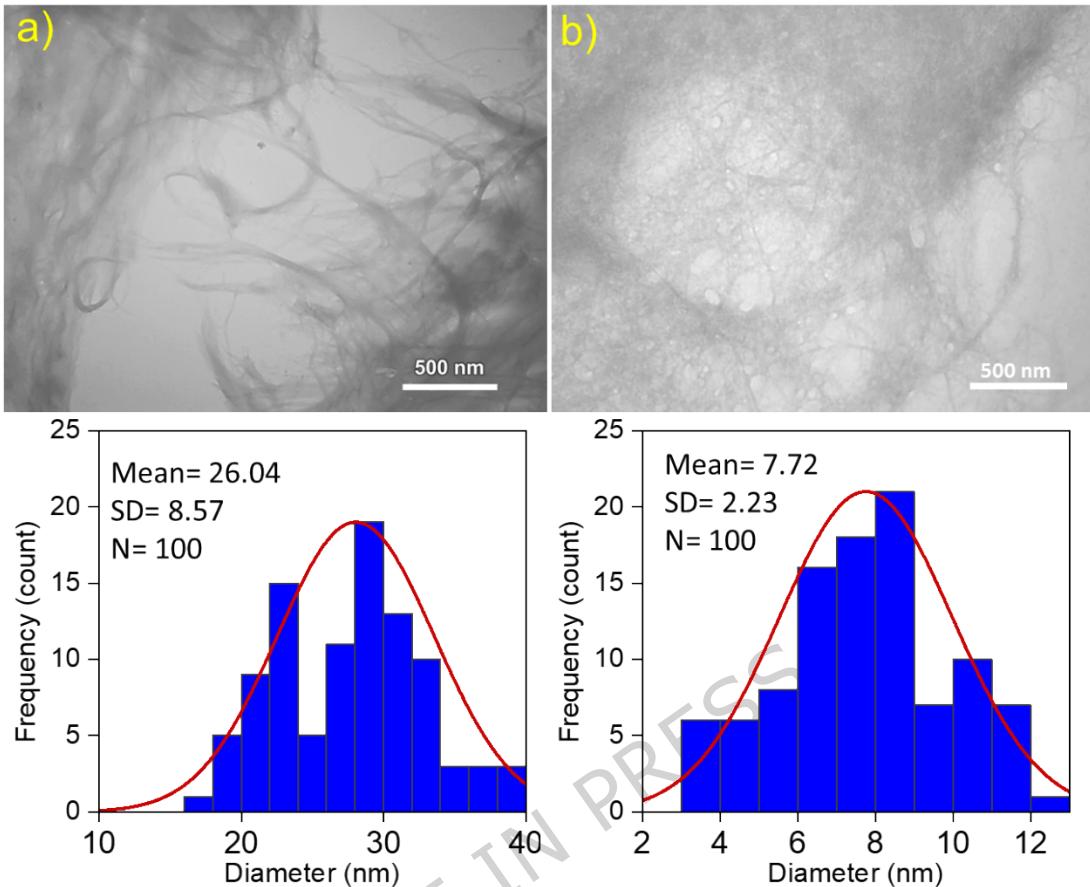


Fig. 2. TEM micrographs and diameter distribution of nanochitin of a) mechanical and b) TEMPO.

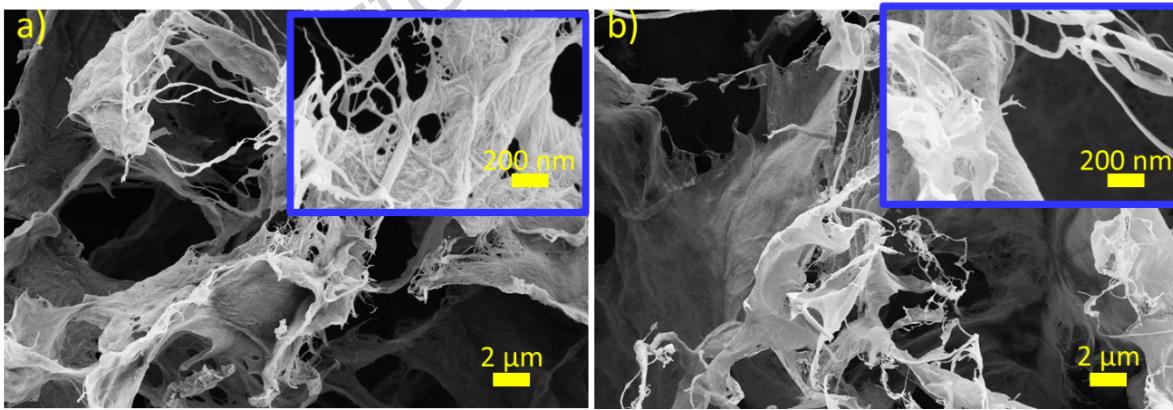


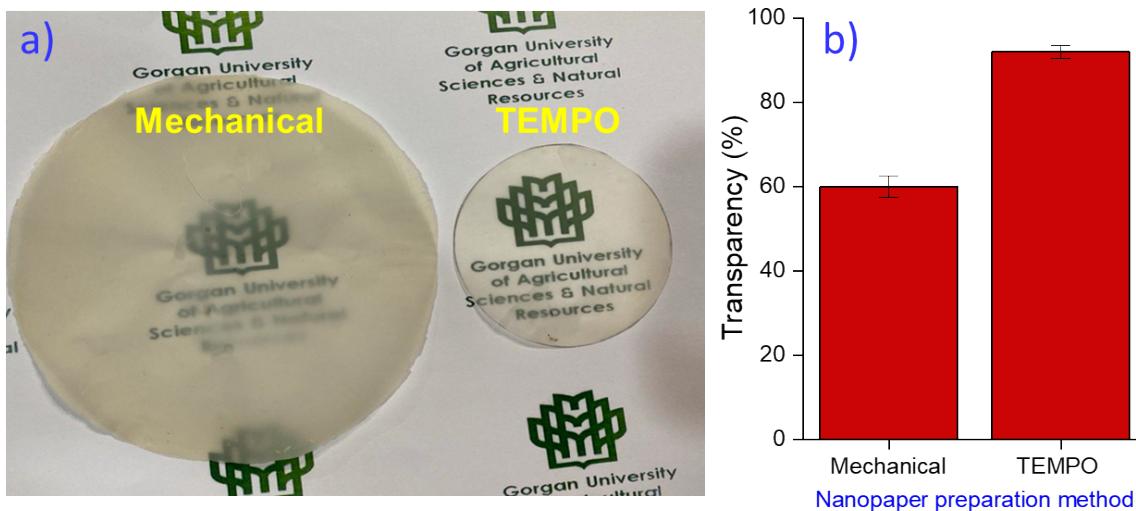
Fig. 3. FESEM micrographs of nanochitin of a) mechanical and b) TEMPO-oxidized.

3.2. Transparency

Figure 4 shows that the apparent transparency of TEMPO-oxidized nanochitin nanopapers is significantly higher than that of mechanical nanopapers (Fig. 4a). The lower transparency in the mechanical nanochitin sample could be due to the non-uniform distribution of fibers and the presence of voids within the structure. These voids increase light scattering and lead to lower transparency. Fukuzumi, et al.²⁴ demonstrated in their

268 study that increased porosity and reduced fiber alignment lead to greater
 269 light scattering and lower transparency. In contrast, the high transparency of
 270 TEMPO-oxidized nanochitin is attributed to its more uniform structure and
 271 decreased fiber size. The TEMPO oxidation process produces nanopaper with
 272 lower porosity and better fiber alignment. Isogai, et al.²⁵ found that TEMPO
 273 oxidation enhances the transparency of nanopaper by generating thinner
 274 fibers and reducing internal porosity.

275 The transparency of mechanically processed and TEMPO-oxidized
 276 nanochitin was evaluated using a UV-Vis spectrophotometer. At a
 277 wavelength of 600 nm, TEMPO-oxidized nanochitin exhibited approximately
 278 92% transparency, whereas mechanically processed nanochitin showed only
 279 60% (Fig. 4b). The higher transparency of the TEMPO-oxidized nanochitin is
 280 attributed to its homogeneous architecture, smaller nanofiber diameters, and
 281 decreased scattering of light owing to its refined microstructural feature²⁶.
 282 Moreover, light transmission is much smoother in the visible region. This
 283 characteristic is a consequence of the chemically modified structure
 284 (carboxylate groups introduction) and the increased surface negative charge,
 285 which induces electrostatic repulsion between the fibers. This repulsion
 286 leads to improved fiber orientation and better matrix formation²⁷. In
 287 contrast, the lower transparency of the mechanically processed nanochitin is
 288 attributed to surface roughness, structural heterogeneity, and enhanced
 289 light scattering²⁵. This heterogeneity results from the mechanical processing
 290 method, which partially damages the internal hydrogen bonds and crystalline
 291 structure of the fibers without changing their chemical structure²⁷. Due to
 292 the high transparency, TEMPO-oxidized nanochitin shows great potential in
 293 biodegradable transparent packaging, optical display panels, and protective
 294 light coatings. On the other hand, mechanically processed nanochitin with
 295 lesser transparency fits towards the requirements of opaque packaging or
 296 shading against the plant light. TEMPO-oxidized nanochitin can be prepared
 297 into films that could replace common plastic films like polyethylene or PET as
 298 biodegradable thin films in the conventional plastic film sector.



300 **Fig. 4. a)** Digital photograph of nanopapers on the Gorgan University of Agricultural
301 Sciences and Natural Resources logo, and b) transparency value of nanopapers.

302
303

304 **3.3. X-ray diffraction (XRD)**

305 Figure 5 illustrates the XRD patterns for the mechanocycled and TEMPO-
306 oxidized nanofibers of chitin, and since XRD is a strong tool for probing the
307 crystalline structure of the material, the nanofibers of chitin were confirmed
308 to be crystalline with the diffraction peaks characteristic of the chitin at 2 θ
309 values of approximately 9°, 19°, 20°, and 21° for the crystallographic planes
310 of chitin. Mechanical and TEMPO-oxidized chitin nanofiber crystallinity
311 indexes were 91.23% and 91.19%, respectively. The increase in chitin
312 crystallinity after the formation of nanofibers has also been reported in the
313 study of Ye, et al. ²⁸ and confirms that at the nanoscale, the crystalline
314 structure of chitin is more regular in shape and compact, biasing the
315 increasing hardness and strength of chitin.

316 Notably, the TEMPO-oxidized nanofibers show a weaker and slightly
317 broader peak at around 9° compared to the mechanically processed
318 nanofibers; in the XRD pattern of the TEMPO-oxidized sample, the diffraction
319 peaks may appear relatively shorter or broader due to the reduction in
320 crystallinity induced by oxidation, while additionally exhibiting stronger
321 peaks at around 14°, 20°, and 21°. The presence of extra peaks or shifts in
322 peak positions may indicate potential alterations in the crystal lattice
323 structure resulting from introducing carboxylate groups ²⁹. Nonetheless, no
324 dramatic differences in the peaks of the two samples in their XRD patterns
325 were noted, implying that the crystalline structure of the chitin is mostly
326 preserved throughout the TEMPO-mediated oxidation procedure ³⁰. It is
327 important to note that variations in preparation methods, the degree of
328 oxidation, and the presence of impurities can affect the XRD patterns of both
329 mechanically processed and TEMPO-oxidized chitin nanofibers.

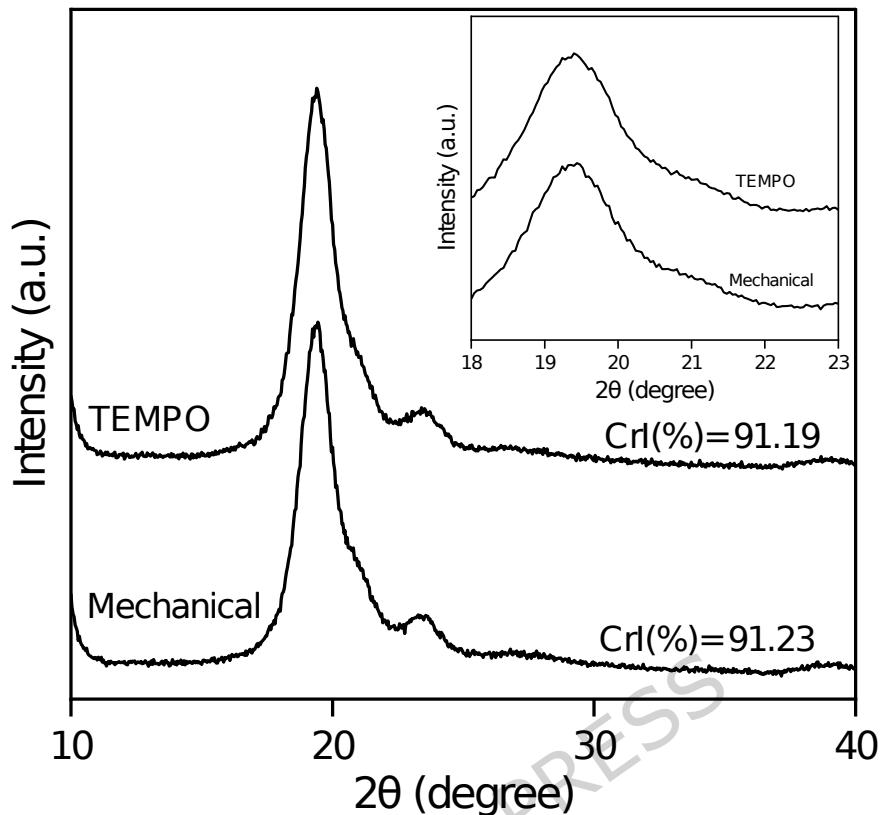
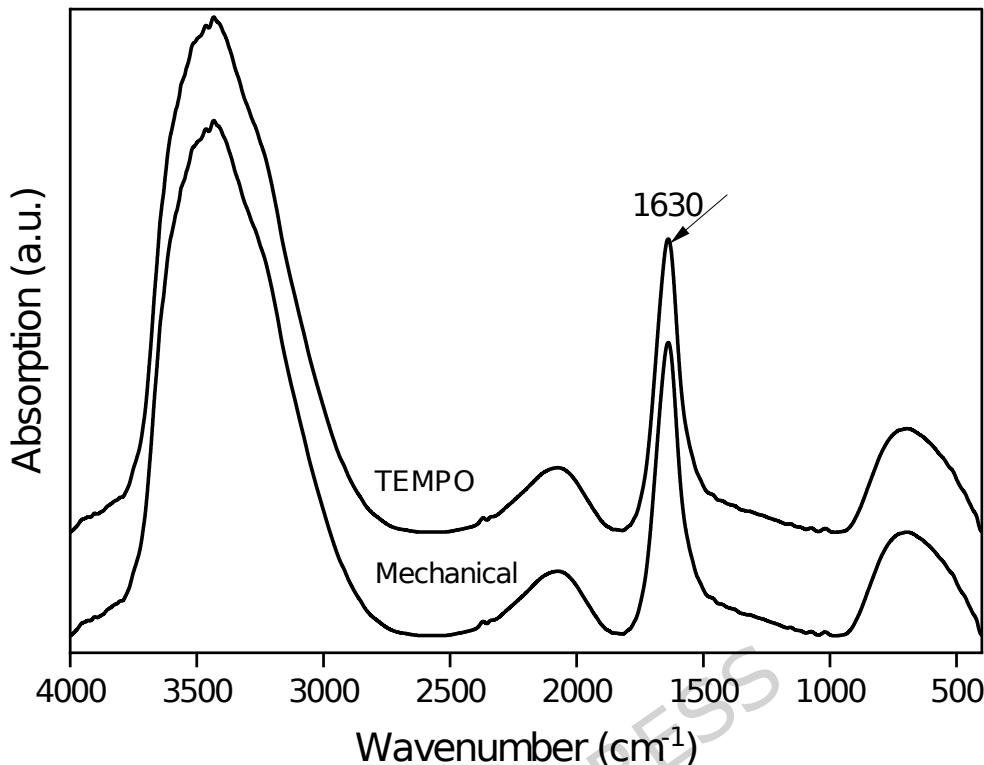


Fig. 5. X-ray diffraction (XRD) pattern of nanochitin nanopapers.

3.4. FTIR

FTIR analysis of the nanopapers reveals distinct absorption peaks that elucidate the chemical structure and modifications resulting from the different processing methods (Fig. 6). The reduced intensity of amine-related bands is in agreement with TEMPO-mediated surface modification, which replaces surface amino functionalities with carboxylate groups, as similarly reported by Salem, et al. ²². The spectra of both mechanically processed and TEMPO-oxidized nanopapers exhibit characteristic bands around 3400 cm^{-1} , corresponding to O-H stretching vibrations that indicate the presence of hydroxyl groups and extensive hydrogen bonding within the chitin matrix ³¹. The peaks near 1630 cm^{-1} and 1560 cm^{-1} are attributed to the amide I ($\text{C}=\text{O}$ stretching) and amide II ($\text{N}-\text{H}$ bending) vibrations ¹², respectively, which confirm the preservation of the chitin backbone. Other bands, such as those near 1310 cm^{-1} and 1150 cm^{-1} , further reflect CH bending and C-O stretching vibrations, underscoring the chemical complexity of the nanostructured films. FTIR findings demonstrate that while the fundamental chitin structure is maintained in both samples, the TEMPO oxidation process induces specific chemical modifications that enhance the functional properties of the resulting nanopapers.



352
353 **Fig. 6.** FTIR spectra of nanochitin nanopapers: a) mechanical and b) TEMPO.
354

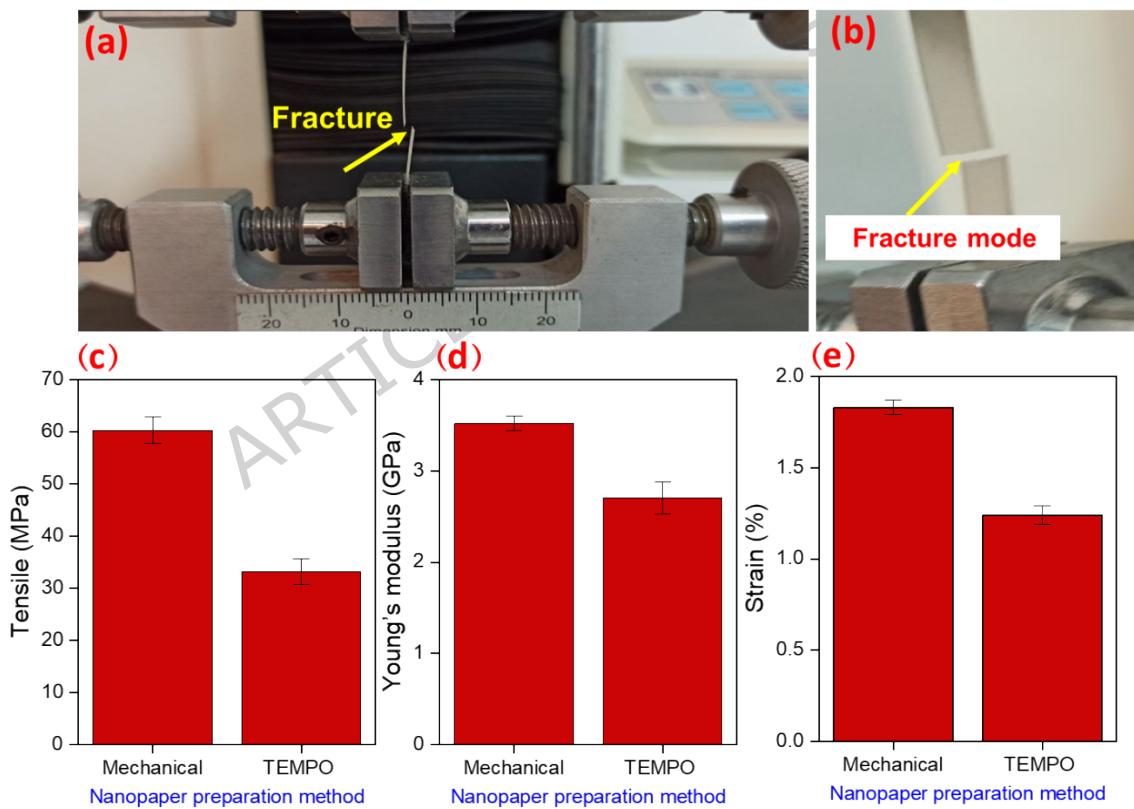
355 **3.5. Mechanical properties**

356 The tensile strength, Young's modulus, and strain of nanopapers made with
357 mechanically processed nanochitin and TEMPO-oxidized nanochitin are
358 shown in Fig. 7. The value of tensile strength for the mechanically processed
359 nanochitin (60.3 MPa) is almost two times higher than that for TEMPO-
360 oxidized nanochitin (33.2 MPa), which may be related to more strong
361 hydrogen bonding of the mechanically processed nanochitin, leading to
362 greater resistance to tensile stresses ³². In the TEMPO-oxidation process, the
363 introduction of carboxyl groups into the nanochitin structure impairs these
364 hydrogen bonds, which may lead to lower tensile strength ²⁵. The Young's
365 modulus of the mechanical nanochitin (3.52 GPa) is higher than that of the
366 TEMPO-oxidized nanochitin (2.70 GPa). The higher Young's modulus of
367 mechanical nanochitin indicates higher stiffness and greater resistance to
368 small deformations. This property is very suitable for preparing films that
369 require high structural strength (such as robust packaging or protective
370 layers) ¹⁷. Moreover, this difference also stems from the more ordered
371 crystalline structure of the mechanically processed nanochitin, as during the
372 mechanical milling process, molecular chains are broken and reconstructed
373 into a denser packing, thus increasing the Young's modulus ¹⁸. Whereas
374 TEMPO oxidation disrupts some hydrogen bonds between the chains and
375 reduces the density of the crystalline structure resulting in a lower Young's
376 modulus ²⁷. The mechanically processed nanopaper exhibits slightly higher
377 crystallinity, which likely contributes to its superior tensile performance. This

378 is because increased crystallinity enhances inter-fibril hydrogen bonding and
 379 load transfer efficiency. These observations are in line with recent findings
 380 on the role of nanoscale ordering in polysaccharide materials ³³.

381 The strain of mechanical nanochitin (1.83%) was higher than that of
 382 TEMPO-oxidized nanochitin (1.24%). In general, the greater flexibility of the
 383 nanochitin structure results in films with thinner thickness and higher specific
 384 surface area, which distributes stress better ³⁴. Although the TEMPO-oxidized
 385 chitin exhibited a more uniform fibril distribution, the mechanical properties
 386 were lower than those of the mechanically processed nanopaper because the
 387 TEMPO treatment introduced carboxylate groups that reduced hydrogen
 388 bonding and decreased crystallinity, thereby limiting stress transfer between
 389 adjacent nanofibrils. The TEMPO-oxidation process creates defects in the
 390 chitin chains that limit the flexibility of the material ²⁷. TEMPO-oxidized
 391 nanochitin, although less flexible, can be well integrated with other materials
 392 or used in environments that require chemical interaction due to its specific
 393 chemical surface (the presence of carboxyl groups) ¹².

394



395

396 **Fig. 7.** a) Digital photograph of nanopaper specimen in the tensile test, b) fracture modes,
 397 c) tensile test, d) Young's modulus, and e) strain.

398

399 **4. Conclusions**

400 The study revealed that both nanopapers prepared from chitin nanofibers
 401 produced through chemical (TEMPO-oxidized) and mechanical (super disk
 402 grinding) processes exhibited high crystallinity values exceeding 90%, with

403 slightly higher crystallinity in the mechanically processed samples. This
404 structural feature contributed to their greater tensile strength and Young's
405 modulus, which are related to stronger hydrogen bonding and a more
406 compact fibrillar network. In contrast, the TEMPO-oxidized nanopapers
407 showed higher optical transmittance (~92%) and better transparency,
408 attributed to finer fibril diameters and more uniform dispersion. These results
409 indicate that mechanical processing favors the formation of stronger and
410 denser nanopapers, while TEMPO oxidation enhances optical uniformity and
411 light transmittance. Overall, the findings demonstrate that the choice of
412 processing method has a direct influence on the structure and properties of
413 chitin nanopapers, allowing their characteristics to be adjusted according to
414 specific performance requirements.

415 **Declarations**

416 **Ethical Approval**

417 Humans or animals are not the subjects of this study. No animals or humans
418 have been studied in this study.

419

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423 **Competing interests**

424 The authors declare that they have no known competing financial interests
425 or personal relationships that could have appeared to influence the work
426 reported in this paper.

427

428

429 **Authors' contributions**

430 **Alireza Mohammadlou:** Data curation, Software, Resources, Formal
431 analysis, Writing-original draft, supervision, and visualization.

432 **Mohammadreza Dehghani Firouzabadi:** Conceptualization, Methodology,
433 Validation, Investigation, Writing-original draft. **Hossein Yousefi:** Project
434 administration and Methodology. All authors read and approved the final
435 manuscript.

436

437 **Data availability**

438 The data provided in this study are available to the corresponding author
439 and can be presented on considerable request.

440

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447 **References**

448 1 Venugopal, V. Green processing of seafood waste biomass towards blue
449 economy. *Current Research in Environmental Sustainability* **4**, 100164
450 (2022).

451 2 Lal, J. *et al.* Diverse uses of valuable seafood processing industry waste for
452 sustainability: a review. *Environmental Science and Pollution Research*, 1-15
453 (2023).

454 3 Iber, B. T., Kasan, N. A., Torsabo, D. & Omuwa, J. W. A review of various
455 sources of chitin and chitosan in nature. *Journal of Renewable Materials* **10**,
456 1097 (2022).

457 4 Chakravarty, J. & Edwards, T. A. Innovation from waste with biomass-derived
458 chitin and chitosan as green and sustainable polymer: a review. *Energy
459 Nexus* **8**, 100149 (2022).

460 5 Tan, H. W., Lim, Z. Y. J., Muhamad, N. A. & Liew, F. F. Potential economic
461 value of chitin and its derivatives as major biomaterials of seafood waste,
462 with particular reference to southeast asia. *Journal of Renewable Materials*
463 **10**, 909 (2022).

464 6 Maraksa, K., Suyotha, W. & Cheirsilp, B. Production of alpha-and beta-chitin,
465 chitosan and protein hydrolysate from seafood processing wastes using an
466 integration of lactic acid and digestive protease from fish viscera as
467 alternative green extraction. *Biocatalysis and Agricultural Biotechnology*,
468 103496 (2025).

469 7 Ezekiel Mushi, N., Butchosa, N., Zhou, Q. & Berglund, L. A. Nanopaper
470 membranes from chitin-protein composite nanofibers—structure and
471 mechanical properties. *Journal of applied polymer science* **131** (2014).

472 8 Zanchetta, E. *et al.* Purification of Cellulose and Chitin Polymers and Other
473 Value-Added Products from the Microalga *Chlorella vulgaris* Using a Green
474 Biorefinery Process. *Fermentation* **11**, 120 (2025).

475 9 Mohan, K. *et al.* Green and eco-friendly approaches for the extraction of chitin
476 and chitosan: A review. *Carbohydrate Polymers* **287**, 119349 (2022).

477 10 Aoun, R. B., Trabelsi, N., Abdallah, M., Mourtzinos, I. & Mhamdi, R. Towards a
478 greener future: Exploring the challenges of extraction of chitin and chitosan
479 as bioactive polysaccharides. *Materials Today Communications*, 108761
480 (2024).

481 11 Ifuku, S. & Saimoto, H. Chitin nanofibers: preparations, modifications, and
482 applications. *Nanoscale* **4**, 3308-3318 (2012).

483 12 Liu, L. *et al.* TEMPO-oxidized nanochitin based hydrogels and inter-structure
484 tunable cryogels prepared by sequential chemical and physical crosslinking.
485 *Carbohydrate Polymers* **272**, 118495 (2021).

486 13 Min, B.-M. *et al.* Chitin and chitosan nanofibers: electrospinning of chitin and
487 deacetylation of chitin nanofibers. *Polymer* **45**, 7137-7142 (2004).

488 14 Ye, W., Liu, L., Wang, Z., Yu, J. & Fan, Y. Investigation of pretreatment
489 methods for improving TEMPO-mediated oxidation and nanofibrillation
490 efficiency of α -chitin. *ACS Sustainable Chemistry & Engineering* **7**, 19463-
491 19473 (2019).

492 15 Ye, W., Yokota, S., Fan, Y. & Kondo, T. A combination of aqueous counter
493 collision and TEMPO-mediated oxidation for doubled carboxyl contents of α -
494 chitin nanofibers. *Cellulose* **28**, 2167-2181 (2021).

495 16 Liu, D., Huang, S., Wu, H. & Zhang, J. Using TEMPO oxidation to tailor
496 deacetylation of carboxyl β -chitin nanofibers from squid pen. *Cellulose* **29**,
497 8539-8549 (2022).

498 17 Yousefi, H. *et al.* Comparative study of paper and nanopaper properties
499 prepared from bacterial cellulose nanofibers and fibers/ground cellulose
500 nanofibers of canola straw. *Industrial Crops and Products* **43**, 732-737
501 (2013).

502 18 Yousefi, H., Faezipour, M., Nishino, T., Shakeri, A. & Ebrahimi, G. All-cellulose
503 composite and nanocomposite made from partially dissolved micro-and
504 nanofibers of canola straw. *Polymer Journal* **43**, 559-564 (2011).

505 19 González, I. *et al.* From paper to nanopaper: evolution of mechanical and
506 physical properties. *Cellulose* **21**, 2599-2609 (2014).

507 20 Fan, Y., Saito, T. & Isogai, A. Preparation of chitin nanofibers from squid pen
508 β -chitin by simple mechanical treatment under acid conditions.
Biomacromolecules **9**, 1919-1923 (2008).

509 21 French, A. D. Idealized powder diffraction patterns for cellulose polymorphs.
Cellulose **21**, 885-896 (2014).

512 22 Salem, K. S., Starkey, H. R., Pal, L., Lucia, L. & Jameel, H. The topochemistry
513 of cellulose nanofibrils as a function of mechanical generation energy. *ACS
514 Sustainable Chemistry & Engineering* **8**, 1471-1478 (2019).

515 23 Chen, R., Huang, W.-C., Wang, W. & Mao, X. Characterization of TEMPO-
516 oxidized chitin nanofibers with various oxidation times and its application as
517 an enzyme immobilization support. *Marine Life Science & Technology* **3**, 85-
518 93 (2021).

519 24 Fukuzumi, H., Saito, T., Iwata, T., Kumamoto, Y. & Isogai, A. Transparent and
520 high gas barrier films of cellulose nanofibers prepared by TEMPO-mediated
521 oxidation. *Biomacromolecules* **10**, 162-165 (2009).

522 25 Isogai, A., Saito, T. & Fukuzumi, H. TEMPO-oxidized cellulose nanofibers.
Nanoscale **3**, 71-85 (2011).

524 26 Fang, Z. *et al.* Novel nanostructured paper with ultrahigh transparency and
525 ultrahigh haze for solar cells. *Nano letters* **14**, 765-773 (2014).

526 27 Saito, T., Kimura, S., Nishiyama, Y. & Isogai, A. Cellulose nanofibers prepared
527 by TEMPO-mediated oxidation of native cellulose. *Biomacromolecules* **8**,
528 2485-2491 (2007).

529 28 Ye, W. *et al.* Comparison of cast films and hydrogels based on chitin
530 nanofibers prepared using TEMPO/NaBr/NaClO and TEMPO/NaClO/NaClO₂
531 systems. *Carbohydrate Polymers* **237**, 116125 (2020).

532 29 Sajomsang, W. & Gonil, P. Preparation and characterization of α -chitin from
533 cicada sloughs. *Materials Science and Engineering: C* **30**, 357-363 (2010).

534 30 Mushi, N. E., Utsel, S. & Berglund, L. A. Nanostructured biocomposite films of
535 high toughness based on native chitin nanofibers and chitosan. *Frontiers in
536 Chemistry* **2**, 99 (2014).

537 31 Hossin, M. A., Al Shaqsi, N. H. K., Al Touby, S. S. J. & Al Sibani, M. A. A review
538 of polymeric chitin extraction, characterization, and applications. *Arabian
539 Journal of Geosciences* **14**, 1870 (2021).

540 32 Sehaqui, H., Salajková, M., Zhou, Q. & Berglund, L. A. Mechanical
541 performance tailoring of tough ultra-high porosity foams prepared from
542 cellulose I nanofiber suspensions. *Soft Matter* **6**, 1824-1832 (2010).

543 33 Salem, K. S. *et al.* Comparison and assessment of methods for cellulose
544 crystallinity determination. *Chemical Society Reviews* **52**, 6417-6446 (2023).

545 34 Habibi, Y., Lucia, L. A. & Rojas, O. J. Cellulose nanocrystals: chemistry, self-
546 assembly, and applications. *Chemical reviews* **110**, 3479-3500 (2010).

547