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Phototrophic bacteria as potential probiotics for corals



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Coral-associated microorganisms provide crucial nutritional, protective, and developmental benefits, yet many functional traits remain unexplored. Phototrophic bacteria may enhance coral nutrition and reduce oxidative stress during bleaching via photosynthesis and antioxidant production. Despite this potential, their role in the holobiont's energy budget and heat stress resilience is understudied. This review explores the functional traits and potential of phototrophic bacteria to enhance coral health and resilience under environmental stress.

The key role of the coral microbiome

Coral bleaching represents a critical threat to coral reefs worldwide, largely driven by the increasing frequency and extended duration of marine heatwaves^{1,2}. This bleaching process occurs due to the breakdown of the symbiotic relationship between the cnidarian host and its photosynthetic algae, *Symbiodiniaceae*^{3,4}. This symbiosis is vital because it provides the coral hosts with photosynthetically fixed carbon, which fulfills up to 90% of their nutritional requirements, in exchange for inorganic waste, which is essential for algal photosynthesis⁵.

During a bleaching event, likely triggered by competition for nutrients^{3,6}, a post-heat stress disorder takes place⁷, and the holobiont needs to deal with a cascade of chain or parallel impacts. One of them is the excessive production of reactive oxygen species (ROS) which overwhelms the antioxidant system and is correlated with the expulsion of the *Symbiodiniaceae*^{4,8,9}. When corals experience severe bleaching, their chances of survival may depend on several key factors. These factors are complementary and, when combined, can enhance coral survival, including: (i) the host's ability to acquire food from heterotrophic sources or use food reserves (e.g., lipids) present in the host tissue, if these reserves are sufficient to sustain the host's metabolic needs during the stress period¹⁰, (ii) the successful recolonization of the algal symbionts in the host tissue once the environmental conditions return to optimal levels, in the cases where corals are still alive when that happens, and (iii) the ability to retain a microbiome that can mitigate some of the damages and offer additional nutritional support, which will be addressed in detail below.

In addition to their symbiotic relationship with algae, corals also form associations with diverse microbial communities that play crucial roles in coral biology (reviewed in refs. 11,12). The coral microbiome composition seems to vary according to the coral host^{13,14}, compartment (i.e., mucus,

skeleton, phycosphere, and tissue)^{15–17}, and geographic location^{18,19}, and provide critical functions for the holobiont's biology, such as providing and recycling essential nutrients, mitigating toxic compounds, controlling pathogens, and releasing other important metabolites to maintain the holobiont homeostasis^{20–22}. The diversity and composition of coral microbiomes are complex^{23,24}, as microbiomes might be flexible and quickly respond to environmental changes^{25,26}, or—comparatively—stable, while still favoring taxa that support the holobiont in withstanding stressors²⁷. Consequently, microbiomes associated with corals and other marine hosts/environments have been heavily impacted by anthropogenic activities, which is correlated with biodiversity loss^{28,29}, turning marine microbiome stewardship (i.e., the use of different microbial therapies that can promote the beneficial restoration and rehabilitation of damaged microbiomes^{12,28}) into an urgent topic^{29,30}.

As an important background for the microbiome stewardship approach²⁸, the coral probiotic hypothesis highlighted the ecological importance and functional roles of the coral microbiome several decades ago³¹. Early data also suggested the potential to manipulate the coral microbiome at different life stages^{32,33}. This potential was later proposed to be harnessed and actively applied to enhance coral resilience to different impacts²². The definition of specific functional beneficial targets and a framework²² as a microbial therapy to promote microbiome stewardship (i.e., the targeted management of the microbiome)²⁸ was followed by the proof of concept that the active rehabilitation of the microbiome of adult corals is possible and shown to mitigate the effects of pathogen infection and thermal bleaching³⁴. A surge of other reports further expanded our knowledge of the potential power of coral probiotics to mitigate an array of impacts, such as oil spills, disease, and thermal stress [e.g.,^{35–42}], and even prevent coral mortality⁷, while demonstrating the feasibility for field applications⁴³.

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However, our understanding of microbial dominance and the potential of the microbiome to be beneficial for the coral host is currently limited to a few taxa, such as the specific probiotic bacteria studied in the papers mentioned above (e.g., *Pseudoalteromonas* spp., *Halomonas* spp., *Cobetia* spp., *Bacillus* sp.), some of them found within the coral tissue⁴², or the prevalence of dominant and coral-tissue associated^{44,45} such as *Endozoicomonas* spp.⁴⁶. Urgent knowledge in the field is required to reveal other potential approaches, tools and microbial candidates that can be applied for coral rehabilitation²⁸. To fill these gaps, a more diverse array of microorganisms should be explored, including members of the coral dark matter (i.e., yet to be cultured, see refs. 47–49). Additionally, different approaches have been suggested to accelerate and improve the selection of additional putative beneficial functions and taxa^{50–53}, and enhance the selection of beneficial microbes and implement even more efficient or alternative microbial therapies^{51,54,55}, including the selection of new probiotic candidates and beneficial functions²⁸, such as the use of phototrophic bacteria.

Phototrophic bacteria are organisms that can use light as their energy source, and are integral components of the coral-associated microbiome in various habitats and bioregions (e.g. refs. 18,56–58,59). These bacteria contain chlorophyll (B-Chl) and abundant carotenoids (Fig. 1), which allow them to harness light through photosynthesis⁶⁰. While their photosynthetic capabilities could theoretically play a similar role of the algal symbionts in supplementing the host's diet during nutrient-poor periods like bleaching^{57,61–63}, this hypothesis remains underexplored. Additionally, phototrophic bacteria produce carotenoids, a diverse class of pigments that play crucial roles as antioxidants and photoprotective agents. These pigments can mitigate oxidative stress by scavenging an array of free radicals, which could protect coral cells from damage^{22,64,65,66}. Carotenoids may also specifically shield the host from reactive oxygen species (ROS) during heat stress, further enhancing coral resilience^{20,21,66}. Although these functional

traits have not yet been fully explored, they place phototrophic bacteria as promising candidates, alongside other taxa, for potentially enhancing coral thermal resilience under climate change. Despite the incomplete understanding of their functional roles within the coral holobiont, this article aims to explore the current knowledge on phototrophic bacteria and their potential contributions to reduce stress by providing additional energy supplies, acting as a barrier against potential light damage, and regulating ROS within the coral holobiont. We also present a framework in which phototrophic bacteria can be considered as putative probiotics, to be further explored in conservation initiatives to improve coral stress resilience.

Who are these “phototrophic bacteria”?

Phototrophic bacteria can be autotrophs or heterotrophs, and the two main mechanisms for autotrophy are retinalphototrophy and chlorophototrophy⁶⁷. Retinalphototrophs use retinal-binding proteins called rhodopsins to convert light energy into chemical energy by generating ion gradients, such as protons, sodium, or chloride (e.g., *Halobacteria* spp. and *Pelagibacter ubique*). Chlorophototrophs use chlorophylls (Chls) and/or bacteriochlorophylls (B-Chls) to harness light energy and facilitate light-driven redox reactions, referred to as photochemistry, to fix carbon dioxide⁶⁸. Notably, while all photosynthetic organisms are chlorophototrophs, some also use organic compounds and are thus classified as photoorganoheterotrophs, while some chlorophototrophs, primarily those that are anoxygenic, also possess the ability to produce retinal-binding proteins. Therefore, not all chlorophototrophs are photosynthetic and certain chlorophototrophs may also function as retinalphototrophs⁶⁷.

Phototrophs include cyanobacteria, sulfur/nonsulfur purple and green bacteria, and heliobacteria. Based on the structure of their photosynthetic machinery and capacity to release oxygen during photosynthesis, they are further divided into other two groups: anoxygenic and oxygenic groups.

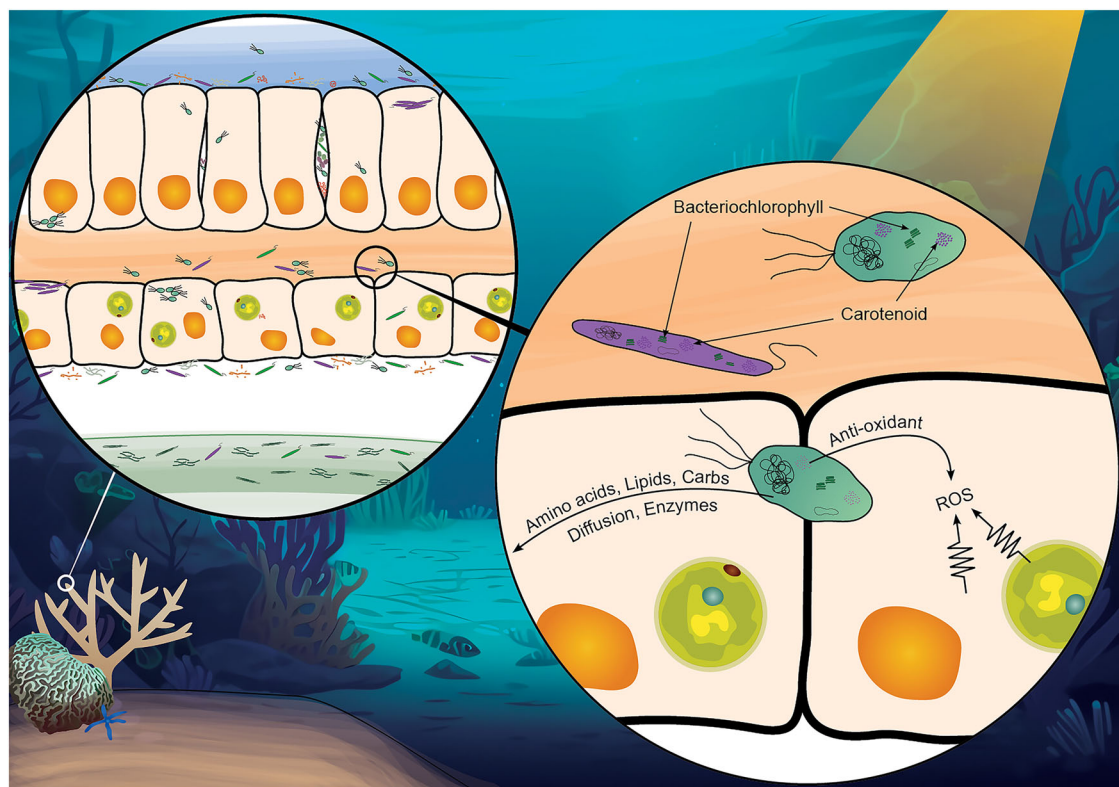


Fig. 1 | Schematic hypothetical illustration of phototrophic bacteria and their potential role in the coral holobiont. Phototrophic bacteria have been recovered from coral mucus and skeleton samples, although it also seems to be associated with coral tissue samples. Phototrophic bacteria contain bacterial chlorophyll and carotenoids. These bacteria fix carbon, to produce organic carbon which can potentially support the host's diet during and after bleaching-induced starvation

periods. However, their contribution to corals' energy budget and mechanisms of nutrient transfer remains unclear. Additionally, many phototrophic bacteria produce large amounts of carotenoids that could act as antioxidants, scavenging excess reactive oxygen species (ROS) during stressful conditions. These bacteria belong to different taxa and therefore may have varying potential capacities for carbon fixation and antioxidant production.

Cyanobacteria can have both photosystems I and II, similar to algae and plants, and can release oxygen during photosynthesis, being the only oxygenic chloroautotrophs⁶⁷. Phototrophic bacteria possess functional photosynthetic systems with well-documented and recently explored carbon fixation pathways that include, but are not restricted to, the Calvin–Benson–Bassham cycle^{69,70}, although some species may lack functional photosynthetic capabilities^{71,72}. Furthermore, most phototrophic bacteria (i.e., purple, green, and heliobacteria) have a single photosystem (either bacteriochlorophyll I or II) and are therefore unable to split water to release oxygen (i.e., anoxygenic). Alternatively, they use simple organic acids (non-sulfur bacteria) and/or hydrogen and sulfide (sulfur bacteria) as electron donors. As such, phototrophic bacteria, which are prevalent members of bacterioplankton communities, play a vital role in the global carbon, nitrogen, and sulfur cycles^{73,74}. Phototrophy facilitated by bacteriochlorophyll (B-Chl) has been estimated to contribute around 5 to 10% of the energy generation in the upper layers of tropical oceans^{75,76} (reviewed in ref. 77).

Phototrophic bacteria have been documented in a wide range of environments, including terrestrial, freshwater, and marine ecosystems^{78,79}. In marine settings, these bacteria have been found in both shallow (e.g., green sulfur bacteria⁸⁰) and deep-sea environments⁸¹, as well as in samples collected across multiple locations, such as hydrothermal vents⁸² and cold seeps⁸³. They can exist as free-living organisms or in association with various hosts and microbes, including endolithic algae of the genus *Ostreobium*⁸⁴, diatoms⁸⁵, sponges⁸⁶, corals¹⁸, oysters⁸⁷, starfish⁸⁸, rhizosphere of mangrove plants⁸⁹, and seagrass leaves⁹⁰. Additionally, phototrophic bacteria have also been found in extreme habitats, such as hot springs with high temperatures, hypersaline lakes, and acidic lakes (see ref. 91). One of the reasons for their ecological success may be explained by their ability to utilize a broad spectrum of light wavelengths (350–1100 nm) to sustain their metabolic processes, due to structural variations in their chlorophylls and bacteriochlorophylls contents^{92,93}. Notably, some of the green sulfur bacteria can utilize either blue light at mesophotic depths⁹⁴ or infrared light emitted near hydrothermal vents for photosynthesis^{82,95}.

As for their identity, phototrophic bacteria containing bacterial chlorophyll are classified into several phyla, including *Cyanobacteria*, *Proteobacteria*, *Chlorobi*, *Chloroflexi*, *Firmicutes*, *Acidobacteria*, and *Gemmatimonadetes*⁹⁶. These groups are widespread in seawater and also associated with corals, residing in various compartments (i.e., tissue, skeleton, or mucus). However, their precise niches within the coral holobiont are still uncertain due to limited empirical data. Phototrophic bacteria, as a functional group, have been predominantly found within the coral skeleton among endolithic communities. They have also been documented, albeit less frequently, in coral tissues and mucus layers, suggesting a more complex spatial distribution within the coral holobiont. For instance, assessments of diazotrophic diversity using the *nifH* gene suggest that while these diazotrophs are primarily abundant in the skeleton, they are also present in coral tissue and mucus⁹⁷. Similarly, cyanobacteria seem to be found in the coral tissue⁹⁸, although their presence is less common compared to the skeleton⁹⁹ and mucus¹⁰⁰. Whether these associations in tissue or mucus are species-specific or persistent, transient, or perhaps incidental remains an open question. Identifying the exact locations of these microbial groups within coral compartments is crucial, as it directly improves our understanding of their physiological roles and their potential for coral health benefits.

Cyanobacteria, in particular, are recognized by their symbiotic associations with corals, exhibiting nitrogen-fixing abilities and transferring photosynthetic assimilates or nitrogen to support the coral host diet^{101–104}. They may use glycerol from the coral's symbiotic algae for metabolic needs, which could play a crucial role in nutrient cycling in tropical nutrient-poor environments⁹⁸. Cyanobacteria also form symbiotic associations and colonize the entire skeletal structure in cold-water corals¹⁰⁵ that show a complete nitrogen cycle similar to those in tropical reefs¹⁰⁶. Interestingly, some cyanobacteria (i.e., *Acaryochloris* sp.) can utilize the red-shifted chlorophyll-d as their main photosynthetic pigment, which may allow them to effectively utilize near-infrared light conditions¹⁰⁷. This infers that certain cyanobacteria strains could play a key role on the coral holobiont physiology

under different light conditions¹⁰⁸. In addition, some cyanobacteria co-evolved with their coral host, although this is species-specific, and their interactions are yet to be fully explored¹⁰⁹. Despite these potentially beneficial interactions, it is important to emphasize that cyanobacteria, which are more resistant to increased light incidence and are becoming more abundant in degraded coral reefs^{110,111}, have also been found associated with coral disease and disease susceptibility^{112,113}.

Unlike cyanobacteria, other phototrophic phyla have received less attention despite their frequent association with corals across different compartments. For example, various taxa of the genus *Erythrobacter* (phylum Proteobacteria) also seems to be found and eventually isolated from both coral mucus¹⁸ and tissue of several reef-building^{114–116} and soft corals^{88,117}. Furthermore, phototrophic bacteria play a significant role in the endolithic community of coral skeletons⁵⁷, as observed in the skeleton of *Isopora palifera* where *Chlorobi* sp. and *Prosthecochloris* sp. are dominant species¹¹⁸. Cai et al.⁸⁰, showed a potential symbiotic relationship between *Prosthecochloris korallensis* and its coral hosts, in which the bacteria potentially offer organic and nitrogenous nutrients and assist the host in sulfide detoxification. In return, the host could create an anaerobic environment suitable for the bacteria's survival, provide carbon dioxide and acetate for their growth, and offer hydrogen sulfide as an electron donor for photosynthesis⁸⁰. Moreover, *Prosthecochloris* sp. associated with *I. palifera* coexists with sulfate-reducing bacteria, further indicating a potential synergistic relationship and functional role within the coral skeleton¹¹⁹. Several other photosynthetic microorganisms, such as *Chromera velia*¹²⁰ and *Vitrella brassicaformis* (both members of the superphylum *Alveolata*)¹²¹ are widespread in corals and phylogenetically close to photosynthetic members of the phylum Apicomplexan, which are also frequently found in coral samples^{120,122}. It is important to highlight, though, that in many cases, the roles of these groups have not been elucidated¹²³. In fact, some studies indicate that *Chromera* spp. may not be beneficial to *Acropora digitifera* larvae, implying they may act as parasites, commensals, or incidental associates¹²⁴. In other words, despite the potential beneficial roles of some phototrophic bacteria, there is still much to discover about their specific functions, and the identification of sub-groups that are not consistently defined as beneficial. The underlying mechanisms of interactions between these microbial groups and the other members of the holobiont also need to be further explored, particularly photosynthesis and antioxidant traits, and whether and how these activities could potentially contribute to coral health.

Photosynthetic capacity of phototrophs and their contribution to holobiont nutrition

Bacteria can transfer beneficial molecules to their hosts across various biological systems. For instance, rhizobacteria fix atmospheric nitrogen for plants, enhancing their growth¹²⁵. In the human gut, bacteria produce short-chain fatty acids that support intestinal health and metabolism¹²⁶. In deep-sea environments, chemosynthetic bacteria may provide essential nutrients to their hosts, such as tube worms, through symbiotic relationships^{127,128}. In coral hosts, phototrophic members of the coral-associated endolithic community (including algae, cyanobacteria, and various bacteria) may also play crucial roles in the primary productivity of coral reefs (reviewed in ref. 57,59,84), although the contribution of phototrophic bacteria to the coral holobiont's energy budget and the mechanisms involved are not yet understood.

Endolithic algae, such as members of the genus *Ostreobium*, have been documented to assimilate inorganic carbon and fix up to 40 $\mu\text{g C cm}^{-2}$ during the day, transferring photoassimilates into the tissues of azooxanthellate corals, such as *Tubastrea micranthus*^{129,130}. In zooxanthellate corals, the endolithic community can also transfer photosynthates to coral tissues during coral bleaching, providing an additional or alternative source of energy and nutrients in the absence of symbiotic dinoflagellates^{61,62,129,131,132}. Additionally, endolithic communities participate in carbon and nitrogen assimilation processes in both healthy and bleached corals, resulting in the translocation of organic

carbon and nitrogen to host tissue^{84,130}. Diazotrophic cyanobacteria, for example, can contribute to coral nitrogen requirements via nitrogen fixation^{130–132} and their abundance is associated with coral health¹³³. Phototrophic bacteria might, therefore, similarly provide the host with photoassimilates through unexplored mechanisms. Cárdenas et al.¹³⁴ proposed a functional connection between endolithic microbiome composition, metabolic activity, and bleaching vulnerability, suggesting that the prevalence of endolithic photoautotrophs could facilitate the translocation of surplus nutrients to coral tissues.

For a long time, cyanobacteria and the green algae *Ostreobium* spp. were believed to be the only contributors to endolithic microbial communities due to their prevalence in coral skeletons⁹⁸. However, more recent studies have revealed that a significant portion of the taxa in these communities are bacteria belonging to various groups, including both anaerobic (members of the phylum *Chloroflexi*) and aerobic (*Chlorobi* spp.) green sulfur bacteria, as well as members of the phyla *Actinobacteria* and *Firmicutes*¹³⁵. Among these, green sulfur bacteria, such as *Prosthecochloris* spp., have been widely reported associated with the coral skeleton^{118,136}.

Among the groups mentioned above, members of the phylum *Chloroflexi* and *Chlorobi* spp. are predominantly found as part of the coral endolithic community¹³⁷, while they have also been reported in the coral tissue^{138,139}. They are typically found in low abundance within corals, but their presence becomes more pronounced when corals inhabit fluctuating¹⁴⁰ or extreme¹⁴¹ environmental conditions. These phototrophic bacteria may contribute significantly to the coral's nitrogen needs, potentially supplying 55–60% of the nitrogen required by their coral host¹⁴². Specifically, green sulfur bacteria, which are potential nitrogen fixers, may provide essential nitrogen and carbon sources to the coral holobiont¹¹⁸. Members of the genus *Prosthecochloris*, for example, possess functional genes related to nitrogen and sulfur metabolism and fixation, suggesting important roles in coral health¹³⁷. These bacteria thrive exclusively in oxygen-depleted skeletal environments, where their metabolic activities influence nitrogen and sulfur cycling. This, in turn, impacts the broader microbial community and enhances coral health¹³⁷. Phototrophic bacteria are could therefore contribute to the input of organic carbon and nitrogen to the coral host (Fig. 1), which plays a vital role in its physiology, particularly as corals typically inhabit oligotrophic waters where nutrient concentrations are low¹⁴³.

Antioxidant capacity

Corals are naturally exposed to temperature and light stress, which may trigger oxidative stress due to excessive production of reactive oxygen species (ROS) within the coral holobiont⁴. When ROS production exceeds the host capacity, this results in damage to the cellular machinery of both the host and symbionts, leading to dysbiosis and contributing to coral bleaching (reviewed in ref. 144). Therefore, corals use several mechanisms to neutralize excess ROS and to minimize damage. The coral microbiome has demonstrated its ability to scavenge ROS¹⁴⁵, for example, which could be due to production of ROS-reactive pigments such as carotenoids¹⁴⁶. This can be achieved by either quenching chlorophyll triplet states, preventing the formation of singlet state oxygen molecules, or directly scavenging singlet oxygen¹⁴⁷. Many bacterial taxa, including photobacteria, possess various pathways for carotenoid production that can dissipate the excess free radicals leaking from the photosynthetic processes of the algal symbionts. This suggests that phototrophic bacteria, which naturally contain large amounts of carotenoids, may play a protective role against oxidative damage and potentially enhance coral fitness, making them promising candidates for probiotics.

In fact, antioxidant capabilities to mitigate toxic compounds and oxidative stress have been proposed as critical traits for selecting beneficial microbial consortia (BMCs), through ROS scavenging mechanisms²², or carotenoids production²⁰, both of which have been explored when selecting and monitoring BMCs^{7,34,51}. ROS scavenging as a tool for selecting efficient BMCs was recently validated as beneficial for corals¹⁴⁵. Further, the presence and activity of carotenoids and ROS scavenging genes have been documented in coral-associated microbes^{7,42,51,64,117,148}. This suggests that some

bacteria associated with corals have the potential and capacity to detoxify free radicals (e.g. ref. 50), which still needs to be further explored and validated. Diaz et al.¹⁴⁹, demonstrated the ability of corals and/or their microbiomes to regulate superoxide in their immediate environment. Similarly, *Flavobacteriaceae* strain (GF1) produced a robust antioxidant zeaxanthin that played a significant protective role for *Symbiodiniaceae* against thermal and light stress¹⁵⁰. Further, microbial manipulation in *Symbiodiniaceae* cultures (using antibiotics) to enhance the prevalence of pigmented mutualistic bacteria has been proven effective in protecting algae from light stress by the production of carotenoids¹⁵¹. Even though they are still exploratory, these results suggest that photobacteria with inherent high levels of carotenoids could play crucial roles in neutralizing reactive oxygen species within the coral holobiont (Fig. 1).

In addition to carotenoids, the ability of the microbiome to control ROS may involve other mechanisms, some of which may or may not be found in phototrophic bacteria. For example, some bacteria have the capacity to produce antioxidant enzymes such as catalase, peroxidase, and superoxide dismutase to protect themselves from oxidative stress¹⁵². Coral-associated bacteria can also synthesize dimethylsulfoniopropionate (DMSP), a crucial chemical compound known for its antioxidant properties (and implication in other key processes), that helps improve coral physiology^{153,154}. For example, members of the class *Alphaproteobacteria* produce DMSP¹⁵⁵, many of which are known to harbor *dsyB* gene (involved in the synthesis of DMSP) which is present in up to 50% of bacterial communities associated with some reef-building coral species¹⁵⁶. Other bacteria are also known to synthesize vitamin B12¹⁵⁷, an important cofactor for the biosynthesis of the amino acid methionine, and is involved in various metabolic pathways, including the generation of antioxidants such as glutathione and DMSP¹⁵⁸. The genes responsible for Vitamin B12 biosynthesis have been identified in coral-associated bacteria¹⁵⁹ and were proposed as beneficial microbes for corals^{21,50}. The presence of similar mechanisms in phototrophic bacteria remains an area for future exploration.

Potential benefits of phototrophic bacteria as probiotics

Phototrophic bacteria have been demonstrated to be beneficial for agricultural crops and colleagues¹⁶⁰ demonstrated that the inoculation of purple non-sulfur bacteria (PNSB) on plants can enhance their growth through various mechanisms, including nutrient acquisition, phytohormone production, and induction of immune system responses. Moreover, PNSB can alleviate abiotic stress in plants through the production of endogenous 5-aminolevulinic acid and induce systemic resistance against pathogens under biotic stress¹⁶⁰. In addition, PNSB can improve soil fertility by increasing nutrients (see ref. 161). Recently, the strain YH-07T of the genus *Erythrobacter* was identified as a promising plant growth-promoting bacterium for future agricultural applications as it can increase nutrient availability, antibiotic biosynthesis, siderophore production, root colonization, and tolerance to harsh environments¹⁶². These findings highlight the potential of phototrophic bacteria as a sustainable probiotic solution in agriculture that can enhance crop production and mitigate environmental stress.

Additionally, photobacteria have been extensively detected in sponge-associated bacterial communities and have the capacity to transfer carbon and nitrogen to the host, providing another example of their significant role in benthic holobiont organisms. Net primary productivity and stable isotope analyses revealed nutrient translocation of photosynthates, including glycerol and organic phosphate, from bacterial and algal symbionts to their sponge hosts¹⁶³. This translocation significantly contributes to host metabolism and growth^{164,165}. Phototrophic cyanobacteria can also contribute up to 80% of the carbon assimilation in sponges^{166,167} enabling them to thrive in low-nutrient areas. In addition, sponge living in low light environment showed that photosynthetically fixed carbon produced by its symbiotic cyanobacteria provides up to 52% of sponge holobiont's respiratory demand and contributed by 7% to total daily carbon uptake when considering the total mixotrophic community¹⁶³. Fixed carbon and nitrogen by phototrophic bacteria could be similarly transferred into coral hosts.

Several phototrophic bacteria members have also demonstrated the potential to enhance host health, although the underlying mechanisms remain unexplored. For instance, co-culture of members of the genus *Erythrobacter* with microalgae (*Marinichlorella kaistiae*) showed a threefold increase in microalgal growth rates and a 20% increase in electron transport rates, improving microalgal photosynthetic rates¹⁶⁸. It is hypothesized that the tested *Erythrobacter* strain may provide *M. kaistiae* with inorganic carbon sources for photosynthesis, as well as vitamins or growth hormones¹⁶⁸. Additionally, *Mameliella alba* (a phototrophic bacterium of the family *Roseobacteraceae*) consistently isolated from dinoflagellate cultures enhanced the dinoflagellate growth rates, suggesting the production of growth-promoting hormone^{146,169}. However, it is important to note that these studies were conducted in vitro, not within the holobiont, where the dynamics may be more complex. Further research is required to elucidate how bacterial contributions may shift from beneficial to harmful, as well as how this dynamic will play out when corals and algae are exposed to stressful conditions. Furthermore, analyses of the genome and core metabolic pathways have revealed that the bacterium “Candidatus *Prosthecochloris korallensis*,” which is associated with coral skeletons, has photoautotrophic capacities⁸⁰. Specifically, a hypothetical mutualistic interaction between the coral host and this microbe, could involve “*Ca. P. korallensis*” providing organic and nitrogenous nutrients to the host, while benefiting from anaerobic conditions, carbon dioxide, acetate for growth, and hydrogen sulfide as an electron donor for photosynthesis provided by the host⁸⁰. In addition, phototrophic bacteria can produce terpenoids which can be important for the coral host¹⁷⁰. Altogether, these mechanisms could fulfill some of the metabolic needs of corals, although targeted studies investigating these specific interactions are needed.

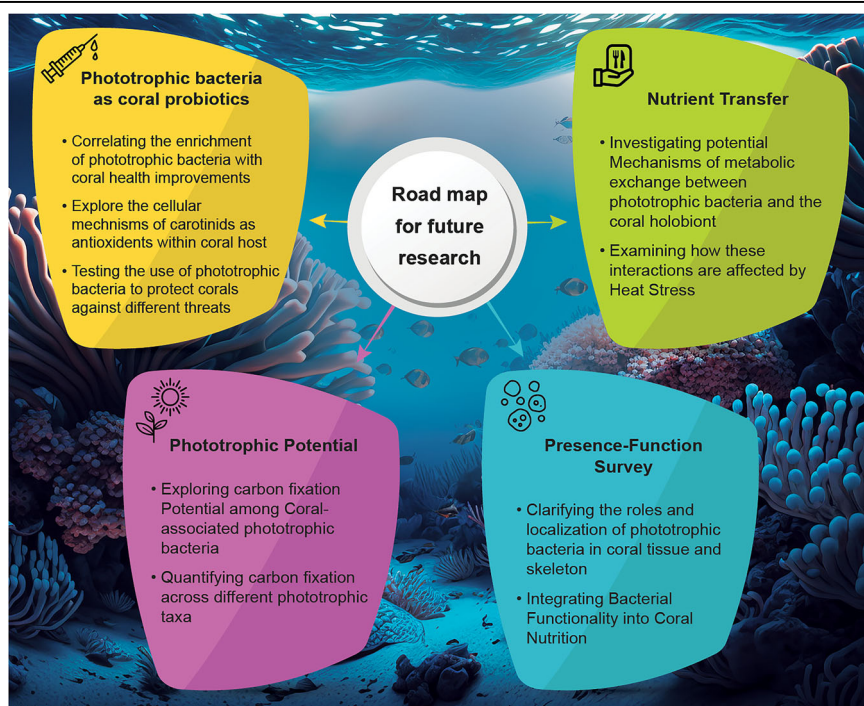
Caveats, challenges, and opportunities

In contrast to the possible benefits described above, studies have suggested that some phototrophic microbes may also have a negative impact on coral health. Algae of the genus *Ostreobium* can penetrate both dead carbonate substrates and live corals which potentially increase the susceptibility of coral colonies to physical damage⁹⁹. In fact, *Ostreobium* can dissolve up to 0.9 kg of CaCO₃ per m² of reef per year¹⁷¹. Further, phototrophic bacteria, such as some *Erythrobacter* spp. strains, have been suggested to be opportunistic pathogens in corals based on their genetic

characteristics and abundance increase correlated with coral disease, such as the white plague^{15,172,173}. *Erythrobacter* spp. strains have been demonstrated to increase the expression of genes related to pathogenicity, such as genes related to the production of membrane disrupting cytotoxins (TlyA and TlyC) and to siderophore scavengers when associated to corals in sugar-rich environments¹⁷⁴. Further, it has been shown that *Erythrobacter* spp. carries genes that encode proteins responsible for the breakdown of multiple plant and algae cell-wall components, therefore, having characteristics of plants and algae pathogens or scavengers¹⁷⁵. Lastly, bacteria of the phylum, *Cyanobacteria* can also negatively impact coral health by outcompeting corals for space on the reef substrate, leading to reduced coral growth and survival¹⁷⁶. Additionally, some *Cyanobacteria* species can produce harmful toxins to corals, causing tissue damage and diseases^{177,178}. This implies that not all phototrophic bacteria are beneficial, underscoring the need for caution in selecting target members (and applying the appropriate dosage), following careful risk assessment steps for the use of microbial therapies²⁸ to minimize potential risks. Furthermore, targeted research on the beneficial members, doses, regime of application and underlying mechanisms promoting coral health may advance and accelerate research in this field (Fig. 2).

The link between phototrophic bacterial abundance and their functions to the host also remains controversial. As such, it might be assumed that the overall low relative abundance of phototrophic bacteria in corals (e.g., *Erythrobacter* sp.¹⁸) indicates a limited contribution to the coral hosts’ diet. We argue here that despite their low relative abundance, the absolute abundance in coral tissue can still be enough to play a vital role in corals (e.g. ref. 179). Low bacterial abundance does not necessarily predict their functional significance, and some of the least abundant microbes in lake water, for example, contribute significantly to nutrient uptake¹⁸⁰. Similarly, low-abundance and core intracellular bacteria have been linked to nitrogen fixation, phytohormone production, and growth promotion in corals⁸¹. Importantly, most probiotic bacteria tested so far are found in low abundances in the coral microbiome and, still, their enrichment is associated with significant improvements in the holobiont health^{7,34,42}. This warrants empirical research and could be a promising avenue for future research (Fig. 2). Understanding the functional roles of phototrophic bacteria, even at low abundances, could inform strategies to improve coral health, particularly in stressed environments.

Fig. 2 | Roadmap highlighting key research topics to improve our understanding of the role of phototrophic bacteria in coral health. This roadmap emphasizes four main research priorities: (1) determining the potential use of phototrophic bacteria as probiotics for corals and underlying mechanisms, (2) exploring the potential of various phototrophic taxa and quantifying their capacity for carbon fixation, (3) investigating nutrient transfer mechanisms to determine whether photoassimilates are transported from bacteria to corals, and (4) identify locations and understanding the role of phototrophic bacteria within the coral holobiont and their contribution to the coral’s energy budget. Addressing these research priorities is crucial for enhancing our understanding of the interactions between phototrophic bacteria and corals.



Another area of uncertainty is whether hosts intentionally increase the abundance of phototrophic (or any other) bacteria during heat stress to benefit from their food resources or whether this is a stochastic response. Evidence suggests that endolithic algae, notably *Ostreobium spp.*, increase in abundance during bleaching and help supplement the coral diet⁶¹. Consequently, biomass, photosynthetic pigments, and the rates of photoassimilate translocation is higher in the skeletons of bleached corals compared to those of non-bleached corals^{61,63} (reviewed in ref. 57,84). Increases in endolithic algae in the coral skeletons could be a result of bleaching, which increases light penetration into the skeleton and drives greater autotrophic nutrients acquisition and higher cell numbers (i.e., the loss of Symbiodiniaceae facilitates greater light penetration, leading to higher phototroph proliferation). This mechanism may be similarly applied to bacteria where certain taxa, including phototrophic bacteria, may increase in abundance (or become more active) due to their functional traits during stress. The correlation between microbial composition shifts and extended coral survival in suboptimal conditions is a well-documented pattern (e.g. refs. 65,182). This relationship is nonetheless complex and although some molecular mechanisms promoted by bacteria have been correlated with coral heat resistance¹⁸³ and survivorship⁷, most of them are yet to be understood. For example, an increased abundance of *Erythrobacter spp.* has been observed following exposure to extreme thermal stress¹⁸⁴ or with rising sea surface temperatures across latitudinal gradients¹⁸, but whether they actually provide functional benefits for corals or are just opportunists remains unknown. Additionally, it is not clear if such regulation is actively driven by the host or the surrounding environment and how it correlates with health improvements. Despite this, chemoautotrophic bacteria seem to be prevalent members of the microbiome associated with deep-sea organisms (e.g., the coral *Callogorgia delta*, the sponge *Aphrocallistes beatrix* and a sea anemone *Ostiactis pearseae*^{19,185,186,186}, and might serve as the primary nutritional source to their hosts, through the translocation of nutrients (reviewed in ref. 128). Based on this observation, a similar nutrient translocation mechanism could be used for phototrophic bacteria in shallow-water corals, potentially enhancing coral resilience and nutrient acquisition in these environments, although this has yet to be explored.

The mechanisms underlying nutrient transfer between phototrophs and corals, as well as the influencing factors, also remain to be elucidated. Phototrophic bacteria may release photoassimilates such as amino acids, glycerol, and fatty acids to coral tissue, similar to algal symbionts¹⁸⁷. These compounds may be transported through diffusion or facilitated by enzymes or transport proteins¹⁸⁸. While members of the phylum *Cyanobacteria* can fix nitrogen and produce organic compounds^{102,189}, the details of nutrient assimilation by the host are still unknown. We propose that mechanisms analogous to those of algal symbionts and cyanobacteria may also apply to phototrophic bacteria. However, this hypothesis is speculative due to the lack of direct evidence testing these pathways. As such, it remains unclear whether phototrophic bacteria employ a similar approach as other symbionts to transfer photoassimilates to their host. Addressing this gap represents a critical opportunity for future research to uncover the potential mechanisms underlying nutrient exchange between phototrophs and coral hosts (Fig. 2).

Additionally, the variability in photosynthetic capabilities across different bacterial phylotypes^{190,191}, hinder our ability to identify which ones can contribute positively to coral health (Fig. 2). Polyphasic approaches combining improved culture-based techniques^{48,49} that can culture different taxa of phototrophic bacteria, multi-omics, and experimental assessments of the effects of phototrophic bacteria on corals, are crucial to explore and correlate their presence, abundance, and function with holobiont health.

Finally, the primary goal of our review is to discuss the largely overlooked potential importance of phototrophic bacteria and their interactions with corals. By expanding the studies to understand their role, localization and mechanisms of interaction and action, we can expand our knowledge on the coral microbiome. Additionally, we can explore the possibility of testing their use as potential probiotics, although such use will also need to overcome other challenges. For

example, the active enrichment of phototrophic bacteria in corals due to their application, or their ability to promote microbiome restructuring following such inoculation, should be among the first steps to be evaluated.

Conclusion

Phototrophic bacteria exhibit functional traits that could be beneficial and play a crucial role to enhance coral survival under stressful conditions. Their active phototrophic capacity suggests an underexplored potential contribution to coral nutrition or competition advantages against coral pathogens. Despite such potential, the extent of the contribution of phototrophic bacteria to the coral diet and overall health remains unclear, indicating a clear opportunity for studies incorporating experimental and isotopic analyses that can provide a more comprehensive understanding of their role. Furthermore, the significant production of anti-oxidant compounds (e.g., carotenoids) by phototrophic bacteria could also contribute to minimizing the concentration of excessive reactive oxygen species (ROS) within the coral holobiont during heat stress. These functional traits highlight the potential role of phototrophic bacteria as coral probiotics and encourage further studies targeting the specific contributions of phototrophic bacteria to the health of the coral holobiont and their underlying mechanisms.

Data availability

No datasets were generated or analyzed during the current study.

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References

- Hughes, T. P. et al. Spatial and temporal patterns of mass bleaching of corals in the Anthropocene. *Science* ((1979)) **359**, 80–83 (2018).
- Reimer, J. D. et al. The fourth global coral bleaching event: Where do we go from here? *Coral Reefs* **43**, 1121–1125 (2024).
- Rädecker, N. et al. Heat stress destabilizes symbiotic nutrient cycling in corals. *Proc. Natl. Acad. Sci. USA* **118**, e2022653118 (2021).
- Suggett, D. J. & Smith, D. J. Coral bleaching patterns are the outcome of complex biological and environmental networking. *Glob. Chang Biol.* **26**, 68–79 (2020).
- Muscantine, L., R. McCloskey, L. & E. Marian, R. Estimating the daily contribution of carbon from zooxanthellae to coral animal respiration1. *Limnol. Oceanogr.* **26**, 601–611 (1981).
- Cui, G. et al. Molecular insights into the Darwin paradox of coral reefs from the sea anemone *Aiptasia*. *Sci. Adv.* **9**, eadf7108 (2023).
- Santoro, E. P. et al. Coral microbiome manipulation elicits metabolic and genetic restructuring to mitigate heat stress and evade mortality. *Sci. Adv.* **7**, 3088–3101 (2021).
- Doering, T. et al. Comparing the role of ROS and RNS in the thermal stress response of two cnidarian models, *Exaiptasia diaphana* and *Galaxea fascicularis*. *Antioxidants* **12**, 1057 (2023).
- Schlottheuber, M. et al. High temporal resolution of hydrogen peroxide (H₂O₂) dynamics during heat stress does not support a causative role in coral bleaching. *Coral Reefs* **43**, 119–133 (2024).
- Grottoli, A. G., Rodrigues, L. J. & Palardy, J. E. Heterotrophic plasticity and resilience in bleached corals. *Nature* **440**, 1186–1189 (2006).
- Mohamed, A. R., Ochsenschühn, M. A., Kazlak, A. M., Moustafa, A. & Amin, S. A. The coral microbiome: towards an understanding of the molecular mechanisms of coral–microbiota interactions. *FEMS Microbiol. Rev.* **47**, fuad005 (2023).
- Voolstra, C. R. et al. The coral microbiome in sickness, in health and in a changing world. *Nat. Rev. Microbiol.* **22**, 460–475 (2024).
- Palacio-Castro, A. M., Rosales, S. M., Dennison, C. E. & Baker, A. C. Microbiome signatures in *Acropora cervicornis* are associated with

- genotypic resistance to elevated nutrients and heat stress. *Coral Reefs* **41**, 1389–1403 (2022).
14. Dubé, C. E. et al. Naturally occurring fire coral clones demonstrate a genetic and environmental basis of microbiome composition. *Nat. Commun.* **12**, 1–12 (2021).
 15. Pollock, F. J. et al. Coral-associated bacteria demonstrate phyllosymbiosis and cophylogeny. *Nat. Commun.* **9**, 4921 (2018).
 16. Hill, L. J. et al. Bacteria associated with the in hospite Symbiodiniaceae's phycosphere. *iScience* **27**, 109531 (2024).
 17. Garrido, A. et al. Antibacterial activity of volatile organic compounds produced by the octocoral-associated bacteria *Bacillus* sp. BO53 and *Pseudoalteromonas* sp. GA327. *Antibiotics* **9**, 1–10 (2020).
 18. Osman, E. O. et al. Coral microbiome composition along the northern Red Sea suggests high plasticity of bacterial and specificity of endosymbiotic dinoflagellate communities. *Microbiome* **8**, 8 (2020).
 19. Osman, E. O. et al. Capacity of deep-sea corals to obtain nutrition from cold seeps aligned with microbiome reorganization. *Glob. Chang Biol.* **29**, 189–205 (2023).
 20. Peixoto, R. S., Harkins, D. M. & Nelson, K. E. Advances in microbiome research for animal health. *Ann. Rev. Anim. Biosci.* **9**, 289–311 (2021).
 21. Peixoto, R. S. et al. Coral probiotics: premise, promise, prospects. *Ann. Rev. Anim. Biosci.* **9**, 265–288 (2021).
 22. Peixoto, R. S. et al. Beneficial microorganisms for corals (BMC): proposed mechanisms for coral health and resilience. *Front. Microbiol.* **8**, 341 (2017).
 23. Voolstra, C. R. & Ziegler, M. Adapting with microbial help: microbiome flexibility facilitates rapid responses to environmental change. *Bioessays* **42**, e2000004 (2020).
 24. Pogoreutz, C. et al. Dominance of *Endozoicomonas* bacteria throughout coral bleaching and mortality suggests structural inflexibility of the *Pocillopora verrucosa* microbiome. *Ecol. Evol.* **8**, 2240–2252 (2018).
 25. Leite, D. C. A. et al. Coral bacterial-core abundance and network complexity as proxies for anthropogenic pollution. *Front. Microbiol.* **9**, 360247 (2018).
 26. McDevitt-Irwin, J. M., Baum, J. K., Garren, M. & Vega Thurber, R. L. Responses of coral-associated bacterial communities to local and global stressors. *Front. Mar. Sci.* **4**, 286253 (2017).
 27. Farias, L. et al. Heat-stressed coral microbiomes are stable and potentially beneficial at the level of taxa and functional genes. *Authorea*. <https://doi.org/10.22541/AU.167542930.03517639/V1> (2023).
 28. Peixoto, R. S. et al. Harnessing the microbiome to prevent global biodiversity loss. *Nat. Microbiol.* **7**, 1726–1735 (2022).
 29. Peixoto, R. S. & Voolstra, C. R. The baseline is already shifted: marine microbiome restoration and rehabilitation as essential tools to mitigate ecosystem decline. *Front. Mar. Sci.* **10**, 1218531 (2023).
 30. Raquel, P. et al. Microbial solutions must be deployed against climate catastrophe. *Nat. Commun.* **15**, <https://doi.org/10.1038/s41467-024-53680-w> (2024).
 31. Reshef, L., Koren, O., Loya, Y., Zilber-Rosenberg, I. & Rosenberg, E. The coral probiotic hypothesis. *Environ. Microbiol.* **8**, 2068–2073 (2006).
 32. Apprill, A., Marlow, H. Q., Martindale, M. Q. & Rappe, M. S. Specificity of associations between bacteria and the coral *Pocillopora meandrina* during early development. *Appl. Environ. Microbiol.* **78**, 7467–7475 (2012).
 33. Wilkinson, S. P. et al. Intra-genomic variation in symbiotic dinoflagellates: recent divergence or recombination between lineages? *BMC Evol. Biol.* **15**, 46 (2015).
 34. Rosado, P. M. et al. Marine probiotics: increasing coral resistance to bleaching through microbiome manipulation. *ISME J.* **13**, 921–936 (2019).
 35. Morgans, C. A., Hung, J. Y., Bourne, D. G. & Quigley, K. M. Symbiodiniaceae probiotics for use in bleaching recovery. *Restor. Ecol.* **28**, 282–288 (2020).
 36. Tang, K. et al. Antagonism between coral pathogen *Vibrio coralliilyticus* and other bacteria in the gastric cavity of scleractinian coral *Galaxea fascicularis*. *Sci. China Earth Sci.* **63**, 157–166 (2020).
 37. Silva, D. P. et al. Multi-domain probiotic consortium as an alternative to chemical remediation of oil spills at coral reefs and adjacent sites. *Microbiome* **9**, 1–19 (2021).
 38. Zhang, Y. et al. Shifting the microbiome of a coral holobiont and improving host physiology by inoculation with a potentially beneficial bacterial consortium. *BMC Microbiol.* **21**, 1–14 (2021).
 39. Ushijima, B. et al. Chemical and genomic characterization of a potential probiotic treatment for stony coral tissue loss disease. *Commun. Biol.* **6**, 1–13 (2023).
 40. Moradi, M. et al. Probiotics mitigate thermal stress- and pathogen-driven impacts on coral skeleton. *Front. Mar. Sci.* **10**, 1212690 (2023).
 41. Li, J. et al. A coral-associated actinobacterium mitigates coral bleaching under heat stress. *Environ. Microbiome* **18**, 83 (2023).
 42. Cardoso, P. M. et al. Localization and symbiotic status of probiotics in the coral holobiont. *mSystems* **9**, e0026124 (2024).
 43. Delgado-Ordoñez, N. et al. Probiotics reshape the coral microbiome in situ without detectable off-target effects in the surrounding environment. *Commun. Biol.* **7**, 1–16 (2024).
 44. Neave, M. J., Apprill, A., Ferrier-Pagès, C. & Voolstra, C. R. Diversity and function of prevalent symbiotic marine bacteria in the genus *Endozoicomonas*. *Appl. Microbiol. Biotechnol.* **100**, 8315–8324 (2016).
 45. Maire, J. et al. Colocalization and potential interactions of *Endozoicomonas* and *Chlamydiae* in microbial aggregates of the coral *Pocillopora acuta*. *Sci. Adv.* **9**, eadg0773 (2023).
 46. Pogoreutz, C. & Ziegler, M. Frenemies on the reef? Resolving the coral–*Endozoicomonas* association. *Trends Microbiol.* <https://doi.org/10.1016/J.TIM.2023.11.006> (2024).
 47. Sweet, M. et al. Insights into the cultured bacterial fraction of corals. *mSystems* **6**, e0124920 (2021).
 48. Schultz, J., Modolon, F., Peixoto, R. S. & Rosado, A. S. Shedding light on the composition of extreme microbial dark matter: alternative approaches for culturing extremophiles. *Front. Microbiol.* **14**, 1167718 (2023).
 49. Modolon, F. et al. In situ devices can culture the microbial dark matter of corals. *iScience* **26**, 108374 (2023).
 50. Rosado, P. M. et al. Exploring the potential molecular mechanisms of interactions between a probiotic consortium and its coral host. *mSystems* **8**, e0092122 (2023).
 51. Doering, T. et al. Advancing coral microbiome manipulation to build long-term climate resilience. *Microbiol. Aust.* **44**, 36–40 (2023).
 52. Raimundo, I., Rosado, P. M., Barno, A. R., Antony, C. P. & Peixoto, R. S. Unlocking the genomic potential of Red Sea coral probiotics. *Sci. Rep.* **14**, 1–19 (2024).
 53. Staab, S., Cardénas, A., Peixoto, R. S., Schreiber, F. & Voolstra, C. R. Coracle—a machine learning framework to identify bacteria associated with continuous variables. *Bioinformatics* **40**, btad749 (2024).
 54. Maire, J. & van Oppen, M. J. H. A role for bacterial experimental evolution in coral bleaching mitigation? *Trends Microbiol.* **30**, 217–228 (2022).
 55. Garcias-Bonet, N. et al. Horizon scanning the application of probiotics for wildlife. *Trends Microbiol.* **32**, 252–269 (2024).
 56. Ritchie, K. Regulation of microbial populations by coral surface mucus and mucus-associated bacteria. *Mar. Ecol. Prog. Ser.* **322**, 1–14 (2006).
 57. Pernice, M. et al. Down to the bone: the role of overlooked endolithic microbiomes in reef coral health. *ISME J.* **14**, 325–334 (2019).
 58. Li, M., Ning, P., Sun, Y., Luo, J. & Yang, J. Characteristics and application of *Rhodospseudomonas palustris* as a microbial cell factory. *Front. Bioeng. Biotechnol.* **10**, 897003 (2022).

59. Saper, J., Raina, J. B., Humphrey, C., Høj, L. & Bourne, D. G. Microbial processes and nutrient uptake in the coral holobiont and reef ecosystems. In *Coral Reef Microbiome*. Coral Reefs of the World, Vol. 20 (eds Peixoto, R. S. & Voolstra, C. R.) https://doi.org/10.1007/978-3-031-76692-3_9 (Springer, Cham, 2025).
60. Thiel, V., Tank, M. & Bryant, D. A. Diversity of chlorophototrophic bacteria revealed in the omics era. *Annu. Rev. Plant Biol.* **69**, 21–49 (2018). <https://doi.org/10.1146/annurev-arplant-042817-040500>
61. Fine, M. & Loya, Y. Endolithic algae: an alternative source of photoassimilates during coral bleaching. *Proc. Biol. Sci. R. Soc.* **269**, 1205–1210 (2002).
62. Ricci, F. et al. Beneath the surface: community assembly and functions of the coral skeleton microbiome. *Microbiome* **7**, 1–10 (2019).
63. Galindo-Martínez, C. T. et al. The role of the endolithic alga *Ostreobium* spp. during coral bleaching recovery. *Sci. Rep.* **12**, 2977 (2022).
64. Dungan, A. M., Bulach, D., Lin, H., van Oppen, M. J. H. & Blackall, L. L. Development of a free radical scavenging bacterial consortium to mitigate oxidative stress in cnidarians. *Microb. Biotechnol.* **14**, 2025 (2021).
65. Dörr, M. et al. Microbial-based therapies to restore and rehabilitate disrupted coral health. In *Coral Reef Microbiome*. Coral Reefs of the World, Vol. 20 (eds Peixoto, R. S. & Voolstra, C. R.) https://doi.org/10.1007/978-3-031-76692-3_13 (Springer, Cham, 2025).
66. Dungan, A. M., Hartman, L. M., Blackall, L. L. & van Oppen, M. J. H. Exploring microbiome engineering as a strategy for improved thermal tolerance in *Exaiptasia diaphana*. *J. Appl. Microbiol.* **132**, 2940–2956 (2022).
67. Bryant, D. A. Phototrophy and phototrophs. *Encycl. Microbiol.* <https://doi.org/10.1016/B978-0-12-809633-8.20672-9> (2019).
68. Overmann, J. & Garcia-Pichel, F. The phototrophic way of life. *Prokaryotes* https://doi.org/10.1007/0-387-30742-7_3 (2006).
69. Garritano, A. N., Song, W. & Thomas, T. Carbon fixation pathways across the bacterial and archaeal tree of life. *PNAS Nexus* **1**, 1–12 (2022).
70. Santos Correa, S., Schultz, J., Lauersen, K. J. & Soares Rosado, A. Natural carbon fixation and advances in synthetic engineering for redesigning and creating new fixation pathways. *J. Adv. Res.* **47**, 75–92 (2023).
71. Sattley, W. M. & Blankenship, R. E. Insights into heliobacterial photosynthesis and physiology from the genome of *Heliobacterium modesticaldum*. *Photosynth Res.* **104**, 113–122 (2010).
72. Orf, G. S. & Redding, K. E. Photosynthesis|the Heliobacteria. *Encycl. Biol. Chem. III* **2**, 352–364 (2021).
73. Fuchs, B. M. et al. Characterization of a marine gammaproteobacterium capable of aerobic anoxygenic photosynthesis. *Proc. Natl. Acad. Sci. USA* **104**, 2891–2896 (2007).
74. Graham, E. D., Heidelberg, J. F. & Tully, B. J. Potential for primary productivity in a globally-distributed bacterial phototroph. *ISME J.* **2018** *12:7* **12**, 1861–1866 (2018).
75. Kolber, Z. S., Van Dover, C. L., Niederman, R. A. & Falkowski, P. G. Bacterial photosynthesis in surface waters of the open ocean. *Nature* **407**, 177–179 (2000).
76. Kolber, Z. S. et al. Contribution of aerobic photoheterotrophic bacteria to the carbon cycle in the ocean. *Science* **292**, 2492–2495 (2001).
77. Kirchman, D. L. Microbial primary production and phototrophy. *Proc. Microb. Ecol.* <https://doi.org/10.1093/OSO/9780198789406.003.0006> (2018).
78. Ferrera, I. et al. Diversity and distribution of freshwater aerobic anoxygenic phototrophic bacteria across a wide latitudinal gradient. *Front. Microbiol.* **8**, 175 (2017).
79. Tahon, G., Tytgat, B. & Willems, A. Diversity of phototrophic genes suggests multiple bacteria may be able to exploit sunlight in exposed soils from the Sør Rondane Mountains, East Antarctica. *Front. Microbiol.* **7**, 232627 (2016).
80. Cai, L. et al. Metagenomic analysis reveals a green sulfur bacterium as a potential coral symbiont. *Sci. Rep.* **7**, 1–11 (2017).
81. Morrow, K. M., Fiore, C. L. & Lesser, M. P. Environmental drivers of microbial community shifts in the giant barrel sponge, *Xestospongia muta*, over a shallow to mesophotic depth gradient. *Environ. Microbiol.* **18**, 2025–2038 (2016).
82. Chen, H. et al. Metagenomic analysis reveals wide distribution of phototrophic bacteria in hydrothermal vents on the ultraslow-spreading Southwest Indian Ridge. *Mar. Life Sci. Technol.* **4**, 255–267 (2022).
83. Shan, Y. et al. A deep-sea bacterium senses blue light via a BLUF-dependent pathway. *mSystems* **7**, e0127921 (2022).
84. Kshitij, T., Juntong, H., Marisa, M. & Pasella, B. L. D. Uthpala Pushpakumara & Heroen Verbruggen Coral Reef Microbiome. 41–46 (Springer Nature Switzerland: Cham, 2025).
85. Kahla, O. et al. Efficiency of benthic diatom-associated bacteria in the removal of benzo(a)pyrene and fluoranthene. *Sci. Total Environ.* **751**, 141399 (2021).
86. Erwin, P. M. & Thacker, R. W. Incidence and identity of photosynthetic symbionts in Caribbean coral reef sponge assemblages. *J. Mar. Biol. Assoc. U. Kingd.* **87**, 1683–1692 (2007).
87. Pira, H., Risdian, C., Müsken, M., Schupp, P. J. & Wink, J. *Pacificimonas pallium* sp. nov., an isolated bacterium from the mantle of pacific oyster *Crassostrea gigas* in Germany, and prediction of one-carbon metabolism. *Diversity ((Basel))* **14**, 181 (2022).
88. Ivanova, E. P. et al. *Erythrobacter vulgaris* sp. nov., a novel organism isolated from the marine invertebrates. *Syst. Appl. Microbiol.* **28**, 123–130 (2005).
89. Borrego, B. B., Melo, L. B. U., Gracioso, L. H., Cardoso, L. O. B. & Perpetuo, E. A. Photoautotrophic microorganisms from mangroves: a review of the ecological role and bioproducts of commercial interest. *Biofuels Bioprod. Bioref.* **17**, 1457–1477 (2023).
90. Hurtado-McCormick, V. et al. Regional and microenvironmental scale characterization of the *Zostera muelleri* seagrass microbiome. *Front. Microbiol.* **10**, 1011 (2019).
91. Madigan, M. T. Anoxygenic phototrophic bacteria from extreme environments. *Photosynth Res.* **76**, 157–171 (2003).
92. Bryant, D. A. & Canniffe, D. P. How nature designs light-harvesting antenna systems: design principles and functional realization in chlorophototrophic prokaryotes. *J. Phys. B: At., Mol. Optical Phys.* **51**, 033001 (2018).
93. Chen, M. Chlorophyll modifications and their spectral extension in oxygenic photosynthesis. *Annu. Rev. Biochem.* **83**, 317–340 (2014).
94. Ritchie, R. J. & Runcie, J. W. Photosynthetic electron transport in an anoxygenic photosynthetic bacterium *Afifella* (*Rhodospseudomonas*) *marina* measured using PAM fluorometry. *Photochem. Photobiol.* **89**, 370–383 (2013).
95. Beatty, J. T. et al. An obligately photosynthetic bacterial anaerobe from a deep-sea hydrothermal vent. *Proc. Natl. Acad. Sci. USA* **102**, 9306 (2005).
96. Raymond, J. Coloring in the tree of life. *Trends Microbiol.* **16**, 41–43 (2008).
97. Lema, K. A., Willis, B. L. & Bourne, D. G. Corals form characteristic associations with symbiotic nitrogen-fixing bacteria. *Appl Environ. Microbiol.* **78**, 3136–3144 (2012).
98. Lesser, M. P., Mazel, C. H., Gorbunov, M. Y. & Falkowski, P. G. Discovery of symbiotic nitrogen-fixing cyanobacteria in corals. *Science* **305**, 997–1000 (2004).
99. Le Campion-Alsumard, T., Golubic, S. & Hutchings, P. Microbial endoliths in skeletons of live and dead corals: *Porites lobata* (Moorea, French Polynesia). *Mar. Ecol. Prog. Ser.* **117**, 149–157 (1995).

100. Naumann, M. S., Richter, C., El-Zibdah, M. & Wild, C. Coral mucus as an efficient trap for picoplanktonic cyanobacteria: implications for pelagic–benthic coupling in the reef ecosystem. *Mar. Ecol. Prog. Ser.* **385**, 65–76 (2009).
101. Lesser, M. P. et al. Nitrogen fixation by symbiotic cyanobacteria provides a source of nitrogen for the scleractinian coral *Montastraea cavernosa*. *Mar. Ecol. Prog. Ser.* **346**, 143–152 (2007).
102. Benavides, M., Bednarz, V. N. & Ferrier-Pagès, C. Diazotrophs: Overlooked key players within the coral symbiosis and tropical reef ecosystems? *Front Mar. Sci.* **4**, 233767 (2017).
103. Bednarz, V. N., Grover, R., Maguer, J. F., Fine, M. & Ferrier-Pagès, C. The assimilation of diazotroph-derived nitrogen by scleractinian corals depends on their Metabolic Status. *mBio* **8**, e02058-16 (2017).
104. Mutalipassi, M. et al. Symbioses of cyanobacteria in marine environments: ecological insights and biotechnological perspectives. *Mar. Drugs* **19**, 227 (2021).
105. Försterra, G. & Häussermann, V. Unusual symbiotic relationships between microendolithic phototrophic organisms and azooxanthellate cold-water corals from Chilean fjords. *Mar. Ecol. Prog. Ser.* **370**, 121–125 (2008).
106. Middelburg, J. J. et al. Discovery of symbiotic nitrogen fixation and chemoautotrophy in cold-water corals. *Sci. Rep.* **5**, 1–9 (2015).
107. Swingley, W. D. et al. Niche adaptation and genome expansion in the chlorophyll d-producing cyanobacterium *Acaryochloris marina*. *Proc. Natl. Acad. Sci. USA* **105**, 2005–2010 (2008).
108. Behrendt, L. et al. Endolithic chlorophyll d-containing phototrophs. *ISME J.* **5**, 1072–1076 (2011).
109. Lema, K. A., Bourne, D. G. & Willis, B. L. Onset and establishment of diazotrophs and other bacterial associates in the early life history stages of the coral *Acropora millepora*. *Mol. Ecol.* **23**, 4682–4695 (2014).
110. Morrow, K. M., Paul, V. J., Liles, M. R. & Chadwick, N. E. Allelochemicals produced by Caribbean macroalgae and cyanobacteria have species-specific effects on reef coral microorganisms. *Coral Reefs* **30**, 309–320 (2011).
111. Charpy, L., Casareto, B. E., Langlade, M. J. & Suzuki, Y. Cyanobacteria in coral reef ecosystems: a review. *J. Mar. Sci.* **2012**, 259571 (2012).
112. Laurent, D. et al. Are cyanobacteria involved in Ciguatera Fish Poisoning-like outbreaks in New Caledonia? *Harmful Algae* **7**, 827–838 (2008).
113. Ford, A. K. et al. Reefs under siege—the rise, putative drivers, and consequences of benthic cyanobacterial mats. *Front Mar. Sci.* **5**, 320011 (2018).
114. Wusqy, K. N., Limantara, L. & Fredy Karwur, F. Exploration, isolation and quantification of β -carotene from bacterial symbiont of *Acropora* sp. *Microbiol Indones.* **8**, 58–64 (2014).
115. Pantos, O. et al. The bacterial ecology of a plague-like disease affecting the Caribbean coral *Montastrea annularis*. *Environ. Microbiol.* **5**, 370–382 (2003).
116. Jaafar, S. N. T. et al. Isolation and identification of bacterial communities from coral tissue affected by black band disease at Pulau Redang, Terengganu, Malaysia. *Appl Ecol. Environ. Res.* **21**, 1823–1835 (2023).
117. Kusmita, L. et al. Characterization of carotenoid pigments from bacterial symbionts of soft-coral *Sarcophyton* sp. from North Java Sea. *Int Aquat. Res.* **9**, 61–69 (2017).
118. Yang, S. H. et al. Prevalence of potential nitrogen-fixing, green sulfur bacteria in the skeleton of reef-building coral *Isopora palifera*. *Limnol. Oceanogr.* **61**, 1078–1086 (2016).
119. Chen, Y. H. et al. Potential syntrophic relationship between coral-associated *Prosthecochloris* and its companion sulfate-reducing bacterium unveiled by genomic analysis. *Micro. Genom.* **7**, 000574 (2021).
120. Moore, R. B. et al. A photosynthetic alveolate closely related to apicomplexan parasites. *Nature* **451**, 959–963 (2008).
121. Obornik, M. et al. Morphology, ultrastructure and life cycle of *Vitrella brassicaformis* n. sp., n. gen., a novel chromerid from the great barrier reef. *Protist* **163**, 306–323 (2012).
122. Vohsen, S. A. et al. Deep-sea corals provide new insight into the ecology, evolution, and the role of plastids in widespread apicomplexan symbionts of anthozoans. *Microbiome* **8**, 34 (2020).
123. Obornik, M. & Lukeš, J. The organellar genomes of chromera and vitrella, the phototrophic relatives of apicomplexan parasites. *Annu. Rev. Microbiol.* **69**, 129–144 (2015).
124. Mohamed, A. R. et al. Deciphering the nature of the coral–Chromera association. *ISME J.* **12**, 776 (2018).
125. Kuan, K. B., Othman, R., Rahim, K. A. & Shamsuddin, Z. H. Plant growth-promoting rhizobacteria inoculation to enhance vegetative growth, nitrogen fixation and nitrogen remobilisation of maize under greenhouse conditions. *PLoS One* **11**, e0152478 (2016).
126. Fusco, W. et al. Short-chain fatty-acid-producing bacteria: key components of the human gut microbiota. *Nutrients* **15**, 2211 (2023).
127. DeChaine, E. G., Bates, A. E., Shank, T. M. & Cavanaugh, C. M. Off-axis symbiosis found: characterization and biogeography of bacterial symbionts of *Bathymodiulus* mussels from Lost City hydrothermal vents. *Environ. Microbiol.* **8**, 1902–1912 (2006).
128. Osman, E. O. & Weinnig, A. M. Microbiomes and obligate symbiosis of deep-sea animals. *Annu. Rev. Anim. Biosci.* **10**, 151–176 (2022).
129. Schlichter, D., Zscharnack, B. & Krisch, H. Transfer of photoassimilates from endolithic algae to coral tissue. *Naturwissenschaften* **82**, 561–564 (1995).
130. Sangsawang, L. et al. 13C and 15N assimilation and organic matter translocation by the endolithic community in the massive coral *Porites lutea*. *R. Soc. Open Sci.* **4**, 171201 (2017).
131. Massé, A. et al. Functional diversity of microboring *Ostreobium algae* isolated from corals. *Environ. Microbiol.* **22**, 4825–4846 (2020).
132. Tityanov, E. A., Kiyashko, S. I., Tityanova, T. V., Kalita, T. L. & Raven, J. A. $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ values in reef corals *Porites lutea* and *P. cylindrica* and in their epilithic and endolithic algae. *Mar. Biol.* **155**, 353–361 (2008).
133. Santos, H. F. et al. Climate change affects key nitrogen-fixing bacterial populations on coral reefs. *ISME J.* **8**, 2272–2279 (2014).
134. Cárdenas, A. et al. Greater functional diversity and redundancy of coral endolithic microbiomes align with lower coral bleaching susceptibility. *ISME J.* **16**, 2406–2420 (2022).
135. Marcelino, V. R. & Verbruggen, H. Multi-marker metabarcoding of coral skeletons reveals a rich microbiome and diverse evolutionary origins of endolithic algae. *Sci. Rep.* **6**, 1–9 (2016).
136. Roush, D., Couradeau, E., Guida, B., Neuer, S. & Garcia-Pichel, F. A new niche for anoxygenic phototrophs as endoliths. *Appl. Environ. Microbiol.* **84**, e02055-17 (2018).
137. Yang, S. H. et al. Metagenomic, phylogenetic, and functional characterization of predominant endolithic green sulfur bacteria in the coral *Isopora palifera*. *Microbiome* **7**, 1–13 (2019).
138. Li, J. et al. Highly heterogeneous bacterial communities associated with the south China Sea reef corals *Porites lutea*, *Galaxea fascicularis* and *Acropora millepora*. *PLoS One* **8**, e71301 (2013).
139. Liu, Y. C. et al. The unexpected diversity of microbial communities associated with black corals revealed by high-throughput Illumina sequencing. *FEMS Microbiol. Lett.* **365**, 167 (2018).
140. Liang, J. et al. Diazotroph diversity associated with scleractinian corals and its relationships with environmental variables in the south China Sea. *Front Physiol.* **11**, 518543 (2020).
141. Maggioni, F., Stenger, P. L., Jourand, P. & Majorel, C. The phylum Chloroflexi and their SAR202 clade dominate the microbiome of two marine sponges living in extreme environmental conditions. *Mar. Ecol.* **44**, e12757 (2023).
142. Ferrer, L. & Szmant, A. Nutrient regeneration by the endolithic community in coral skeletons. In *Proc. 6th International Coral Reef Symposium* 1–4 (1988).

143. Tanaka, Y. et al. Distribution of dissolved organic carbon and nitrogen in a coral reef. *Coral Reefs* **30**, 533–541 (2011).
144. Helgoe, J., Davy, S. K., Weis, V. M. & Rodriguez-Lanetty, M. Triggers, cascades, and endpoints: connecting the dots of coral bleaching mechanisms. *Biol. Rev.* **99**, 715–752 (2024).
145. Tang, X. et al. Validating the use of ROS-scavenging bacteria as probiotics to increase coral resilience to thermal stress. *J. Oceano. Limnol.* **42**, 1242–1260 (2024).
146. Varasteh, T. et al. Conserved pigment profiles in phylogenetically diverse symbiotic bacteria associated with the corals *Montastraea cavernosa* and *Mussismilia braziliensis*. *Micro. Ecol.* **81**, 267–277 (2021).
147. Kiokias, S. & Gordon, M. H. Antioxidant properties of carotenoids in vitro and in vivo. *Food Rev. Int.* **20**, 99–121 (2004).
148. Rosado, P. M. et al. Marine probiotics: increasing coral resistance to bleaching through microbiome manipulation. *ISME J.* **13**, 921–936 (2018).
149. Diaz, J. M. et al. Species-specific control of external superoxide levels by the coral holobiont during a natural bleaching event. *Nat. Commun.* **7**, 13801 (2016).
150. Motone, K. et al. A zeaxanthin-producing bacterium isolated from the algal phycosphere protects coral endosymbionts from environmental stress. *mBio* **11**, e01019-19 (2020).
151. Takagi, T. et al. Mutualistic interactions between dinoflagellates and pigmented bacteria mitigate environmental stress. *Microbiol. Spectr.* **11**, e0246422 (2023).
152. Imlay, J. A. Where in the world do bacteria experience oxidative stress? *Environ. Microbiol.* **21**, 521–530 (2019).
153. Kuek, F. W. I. et al. DMSP production by coral-associated bacteria. *Front. Mar. Sci.* **9**, 869574 (2022).
154. Sunda, W., Kieber, D. J., Kiene, R. P. & Huntsman, S. An antioxidant function for DMSP and DMS in marine algae. *Nature* **418**, 317–320 (2002).
155. Curson, A. R. J. et al. Dimethylsulfoniopropionate biosynthesis in marine bacteria and identification of the key gene in this process. *Nat. Microbiol.* **2**, 1–9 (2017).
156. Luo, D. et al. Population differentiation of Rhodobacteraceae along with coral compartments. *ISME J.* **15**, 3286–3302 (2021).
157. Raux, E., Schubert, H. L. & Warren, M. J. Biosynthesis of cobalamin (vitamin B12): a bacterial conundrum. *Cell Mol. Life Sci.* **57**, 1880–1893 (2000).
158. Croft, M. T., Lawrence, A. D., Raux-Deery, E., Warren, M. J. & Smith, A. G. Algae acquire vitamin B12 through a symbiotic relationship with bacteria. *Nature* **438**, 90–93 (2005).
159. Robbins, S. J. et al. A genomic view of the reef-building coral *Porites lutea* and its microbial symbionts. *Nat. Microbiol.* **4**, 2090–2100 (2019).
160. Lee, S. K., Lur, H. S. & Liu, C. Te. From lab to farm: elucidating the beneficial roles of photosynthetic bacteria in sustainable agriculture. *Microorganisms* **9**, 2453 (2021).
161. Sakarika, M. et al. Purple non-sulphur bacteria and plant production: benefits for fertilization, stress resistance and the environment. *Microb. Biotechnol.* **13**, 1336–1365 (2020).
162. Tang, T., Sun, X., Dong, Y. & Liu, Q. *Erythrobacter aureus* sp. nov., a plant growth-promoting bacterium isolated from sediment in the Yellow Sea, China. *3 Biotech* **9**, 1–9 (2019).
163. Hudspith, M. et al. Harnessing solar power: photoautotrophy supplements the diet of a low-light dwelling sponge. *ISME J.* **16**, 2076–2086 (2022).
164. Koopmans, M. et al. Carbon conversion and metabolic rate in two marine sponges. *Mar. Biol.* **158**, 9–20 (2011).
165. Weisz, J. B., Massaro, A. J., Ramsby, B. D. & Hill, M. S. Zooxanthellar symbionts shape host sponge trophic status through translocation of carbon. *Bio. Bull.* **219**, 189–197 (2010).
166. Thacker, R. W. Impacts of shading on sponge-cyanobacteria symbioses: a comparison between host-specific and generalist associations. *Integr. Comp. Biol.* **45**, 369–376 (2005).
167. Wilkinson, C. R. Net primary productivity in coral reef sponges. *Science* **219**, 410–412 (1983).
168. Martin, N. et al. Synthetic algal-bacteria consortia for space-efficient microalgal growth in a simple hydrogel system. *J. Appl. Phycol.* **33**, 2805–2815 (2021).
169. Ren, C. Z. et al. Taxonomic and bioactivity characterizations of *Mamieliella alba* strain LZ-28 isolated from highly toxic marine dinoflagellate *Alexandrium catenella* LZT09. *Mar. Drugs* **20**, 321 (2022).
170. Klaus, O. et al. Engineering phototrophic bacteria for the production of terpenoids. *Curr. Opin. Biotechnol.* **77**, 102764 (2022).
171. Grange, J. S., Rybarczyk, H. & Tribollet, A. The three steps of the carbonate biogenic dissolution process by microborers in coral reefs (New Caledonia). *Environ. Sci. Pollut. Res.* **22**, 13625–13637 (2015).
172. Cardenas, A., Rodriguez, R. L., Pizarro, V., Cadavid, L. F. & Arevalo-Ferro, C. Shifts in bacterial communities of two Caribbean reef-building coral species affected by white plague disease. *ISME J.* **6**, 502–512 (2012).
173. Sunagawa, S., DeSalvo, M. K., Voolstra, C. R., Reyes-Bermudez, A. & Medina, M. Identification and gene expression analysis of a taxonomically restricted cysteine-rich protein family in reef-building corals. *PLoS One* **4**, e4865 (2009).
174. Cárdenas, A. et al. Excess labile carbon promotes the expression of virulence factors in coral reef bacterioplankton. *ISME J.* **12**, 59–76 (2018).
175. Cho, S. H. et al. Assessment of erythrobacter species diversity through pan-genome analysis with newly isolated erythrobacter sp. 3-20A1M. *J. Microbiol. Biotechnol.* **31**, 601 (2021).
176. Kuffner, I. B. et al. Inhibition of coral recruitment by macroalgae and cyanobacteria. *Mar. Ecol. Prog. Ser.* **323**, 107–117 (2006).
177. Myers, J. L., Sekar, R. & Richardson, L. L. Molecular detection and ecological significance of the cyanobacterial genera *Geitlerinema* and *Leptolyngbya* in black band disease of corals. *Appl Environ. Microbiol.* **73**, 5173–5182 (2007).
178. Gantar, M., Sekar, R. & Richardson, L. L. Cyanotoxins from black band disease of corals and from other coral reef environments. *Micro. Ecol.* **58**, 856–864 (2009).
179. Prioux, C. et al. Unveiling microbiome changes in Mediterranean octocorals during the 2022 marine heatwaves: quantifying key bacterial symbionts and potential pathogens. *Microbiome* **11**, 1–19 (2023).
180. Musat, N. et al. A single-cell view on the ecophysiology of anaerobic phototrophic bacteria. *Proc. Natl. Acad. Sci. USA* **105**, 17861–17866 (2008).
181. Ainsworth, T. D. et al. The coral core microbiome identifies rare bacterial taxa as ubiquitous endosymbionts. *ISME J.* <https://doi.org/10.1038/ismej.2015.39> (2015).
182. Ziegler, M., Seneca, F. O., Yum, L. K., Palumbi, S. R. & Voolstra, C. R. Bacterial community dynamics are linked to patterns of coral heat tolerance. *Nat. Commun.* **8**, 14213 (2017).
183. Erika, P. et al. Inherent differential microbial assemblages and functions associated with corals exhibiting different thermal phenotypes. *Sci. Adv.* **11**, <https://doi.org/10.1126/sciadv.adq2583> (2025).
184. Lee, S. T. M. et al. Successive shifts in the microbial community of the surface mucus layer and tissues of the coral *Acropora muricata* under thermal stress. *FEMS Microbiol. Ecol.* **91**, 289–296 (2015).
185. Alessandro, N. G. et al. Torsten Thomas, Simple Porifera holobiont reveals complex interactions between the host, an archaeon, a bacterium, and a phage, *The ISME J.* **18**, wrae197 (2024).
186. Goffredi, S. K. et al. Mixotrophic chemosynthesis in a deep-sea anemone from hydrothermal vents in the Pescadero Basin, Gulf of California. *BMC Biol.* **19**, 1–18 (2021).
187. Trench, R. K. Microalgal-invertebrate symbioses—a review. *Endocytobiosis Cell Res.* **9**, 135–175 (1993).
188. Yellowlees, D., Rees, T. A. V. & Leggat, W. Metabolic interactions between algal symbionts and invertebrate hosts. *Plant Cell Environ.* **31**, 679–694 (2008).

189. Cardini, U. et al. Functional significance of dinitrogen fixation in sustaining coral productivity under oligotrophic conditions. *Proc. Biol. Sci.* **282**, 20152257 (2015).
190. Kushkevych, I., Procházka, J., Gajdács, M., Rittmann, S. K. M. R. & Vítězová, M. Molecular physiology of anaerobic phototrophic purple and green sulfur bacteria. *Int. J. Mol. Sci.* **22**, 6398 (2021).
191. Rasmussen, M. & Minteer, S. D. Photobioelectrochemistry: solar energy conversion and biofuel production with photosynthetic catalysts. *J. Electrochem. Soc.* **161**, H647–H655 (2014).

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Author contributions

E.O.O. and R.S.P. developed the review outline, while E.O.O. drafted the initial MS version, and all other authors contributed to writing, editing, and approving the final manuscript.

Competing interests

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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