

# SCIENTIFIC REPORTS

OPEN

## Role of ectonucleotide pyrophosphatase/phosphodiesterase 2 in the midline axis formation of zebrafish

Frisca Frisca<sup>1,2</sup>, Daniel Colquhoun<sup>1</sup>, Yona Goldshmit<sup>1,3</sup>, Minna-Liisa Änkö<sup>4</sup>, Alice Pébay<sup>2,\*</sup> & Jan Kaslin<sup>1,\*</sup>

Received: 13 April 2016

Accepted: 01 November 2016

Published: 24 November 2016

Lysophosphatidic acid (LPA) is a unique bioactive lysophospholipid that induces pleiotropic effects in various cell types and organisms by acting on its specific receptors. LPA is mainly synthesised extracellularly by the ectonucleotide pyrophosphatase/phosphodiesterase 2/autotaxin (enpp2). Altered LPA signalling is associated with embryonic abnormalities, suggesting critical roles for LPA during development. However, the role of LPA signalling during early embryogenesis is not well established. We demonstrate that enpp2/LPA signalling in the early zebrafish embryo results in altered axis and midline formation, defects in left right (L-R) patterning, ciliogenesis of the Kupffer's vesicle (KV), through the modulation of cell migration during gastrulation in a *lpar*<sub>1-3</sub> Rho/ROCK-dependant manner. Overall, this study demonstrates an essential role of enpp2/LPA signalling during early embryogenesis.

The midline is an essential embryonic structure during vertebrate embryogenesis. It provides structural support of vertebrates, and plays a role in tissue patterning and in the establishment of the left right asymmetry (L-R) during internal organogenesis<sup>1-3</sup>. The development of the midline/body axis occurs from the onset of gastrulation. The process is initially marked by mesoderm and endoderm progenitor cell compaction at the dorsal side of the gastrula (dorsal organizer shield) which then undergoes simultaneous morphological changes, cellular intercalation and migration (convergent extension, CE) that is governed by specific signalling pathways to form the body axis<sup>4,5</sup>. Failure in this process and the associated signalling pathways alter midline formation and result in developmental abnormalities including impaired elongation of the body length<sup>6-8</sup>, establishment of L-R asymmetry<sup>9</sup> and tissue patterning<sup>10</sup>. Many key signalling molecules and signalling pathways regulate midline formation during embryogenesis, including Platelet-Derived Growth Factor (PDGF)<sup>11,12</sup>, the non-canonical Wnt/Planar Cell Polarity (PCP) pathway<sup>13</sup>, Bone Morphogenetic Protein (BMPs)<sup>14</sup> and Fibroblast Growth Factor (FGFs)<sup>15</sup>. However, the role of phospholipid signalling in this context remains poorly understood.

Lysophosphatidic acid (LPA) is a unique bioactive lysophospholipid that induces pleiotropic effects through binding to its specific G-protein coupled receptors *lpa*<sub>1-6</sub> (reviewed in refs 16 and 17) in various cell types. LPA signalling affects proliferation, cell survival, motility, morphological rearrangements and differentiation. Dysregulation of LPA signalling is associated with early and late embryonic abnormalities, suggesting critical roles for LPA during development<sup>18-21</sup>. Studies in LPA receptor knockout mouse models have shown a prominent role of LPA signalling in particular in neural and vascular development<sup>19,22</sup>. *Lpa*<sub>1</sub><sup>(-/-)</sup> mice die postnatally due to an impaired suckling behavior likely associated with a defective olfaction and impaired CNS development<sup>19</sup>. *Lpa*<sub>2</sub><sup>(-/-)</sup> mice do not display severe phenotypes although some intracellular signalling pathways such as PLC activation, Ca<sup>2+</sup> mobilization, and stress fiber formation are altered<sup>23</sup>. *Lpa*<sub>3</sub><sup>(-/-)</sup> female mice display a reproductive impairment, with delayed embryo implantation and embryo spacing alterations<sup>24</sup> while *Lpa*<sub>4</sub><sup>(-/-)</sup> mice have defects in blood vessel formation leading to haemorrhages in many organs at different embryonic stages<sup>22</sup>.

<sup>1</sup>Australian Regenerative Medicine Institute, Building 75, Monash University, Australia. <sup>2</sup>Centre for Eye Research Australia, Royal Victorian Eye and Ear Hospital & Ophthalmology, the University of Melbourne, Department of Surgery, Australia. <sup>3</sup>Department of Neurobiology, Tel-Aviv University, Israel. <sup>4</sup>Monash Biomedicine Discovery Institute, Department of Anatomy and Developmental Biology, Biomedicine Discovery Institute, Monash University, Australia. \*These authors contributed equally to this work. Correspondence and requests for materials should be addressed to J.K. (email: jan.kaslin@monash.edu)

Lpa<sub>5</sub><sup>(-/-)</sup> mice show deficit in response to neuropathic pain (reviewed in ref. 25) and the loss of lpa<sub>6</sub> in *Xenopus* embryo disrupts forebrain development<sup>26</sup>.

The ectonucleotide pyrophosphatase/phosphodiesterase 2 (enpp2), also known as autotaxin (atx) or lysophospholipase D, is the main enzyme responsible for the synthesis of extracellular LPA<sup>27,28</sup>. The loss of enpp2 in mice is lethal between E9.5–E10.5 because of severe vascular defects in the yolk sac and embryo<sup>25,29,30</sup>, hampering the characterization of the LPA function in the early mouse embryo. In the early zebrafish embryo, the *enpp2/lpa<sub>3</sub>* axis regulates the L-R internal organ asymmetry but the molecular mechanisms responsible are unknown<sup>31</sup>. Furthermore, overexpression of *enpp2* in zebrafish induces cardia bifida<sup>32</sup>. Enpp2/LPA signalling also regulates oligodendrocyte production and differentiation of the zebrafish hindbrain<sup>33</sup>. Although these studies suggest essential roles for LPA signalling during embryogenesis, the understanding of LPA functions remains limited.

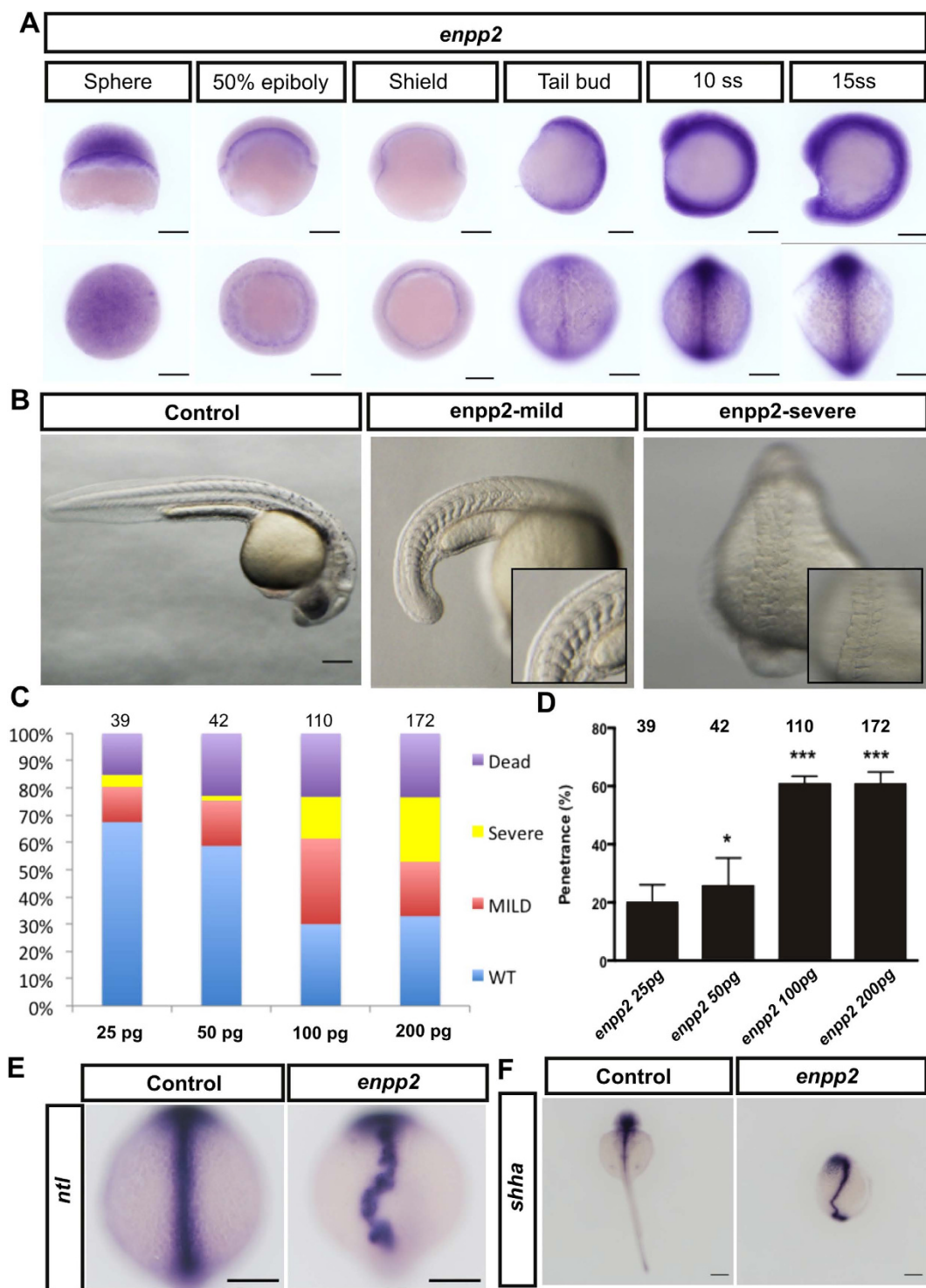
To determine the function of LPA during embryogenesis, we transiently overexpressed the LPA-producing enzyme enpp2 in developing zebrafish. Our gain of function study of *enpp2* in the zebrafish embryo shows for the first time the unique role of *enpp2* in regulating the cell migration during gastrulation and subsequent formation of the axial midline, as well as the establishment of L-R asymmetry. Furthermore, rescue experiments show that these effects are lpa<sub>1-3</sub>-dependent and mediated by the Rho/ROCK pathway.

## Results

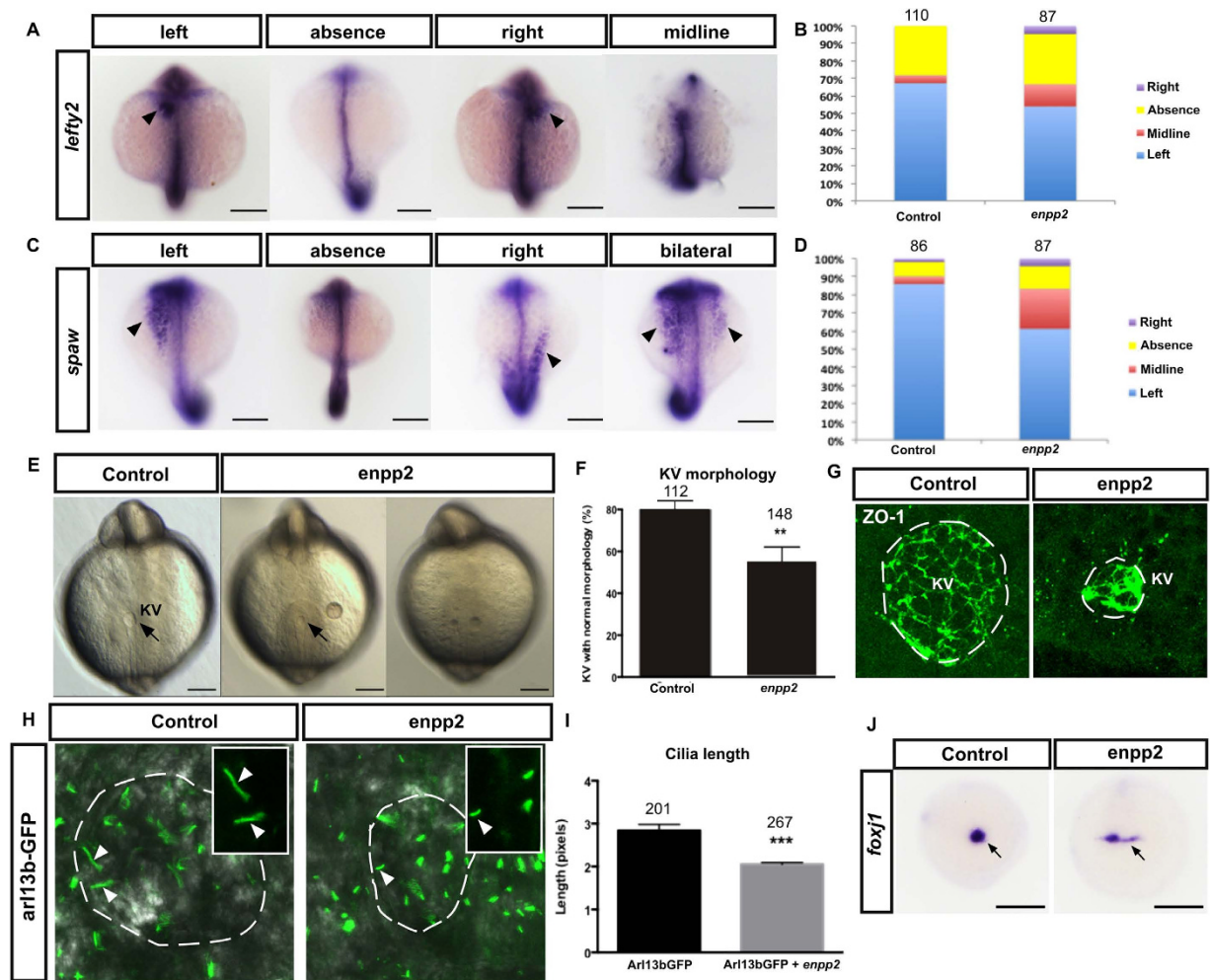
**Overexpression of enpp2 alters axis formation in the zebrafish embryo and modulates expression of the midline axis genes *shha* and *ntl*.** We cloned the full-length *enpp2* (NM\_200603.1)<sup>34</sup> to generate probes for whole mount *in situ* hybridization (WISH). WISH analysis of the developing zebrafish embryos showed that *enpp2* mRNA expression is dynamically regulated during development, which was also confirmed by quantitative RT-PCR analysis (Fig. 1A, Suppl. Fig. 1). *Enpp2* was maternally deposited (sphere) and expressed at a very low level in the yolk syncytial layer (YSL) during the early gastrulation stage (50% epiboly and shield) (Fig. 1A, Suppl. Fig. 1A,B). At the end of gastrulation (tail bud), *enpp2* was notably expressed in the midline axis and its levels continued to increase during the segmentation period (10 somite stage, 15 somite stage) (Fig. 1A, Suppl. Fig. 1), suggesting a potential role in midline and axis formation. In order to assess the role of enpp2 during embryogenesis, we injected increasing amounts of capped *enpp2* mRNA into the zebrafish embryo (1–4 cell stage), determined the morphology (Fig. 1B) and measured the phenotype penetrance (Fig. 1C). The overexpression of enpp2 resulted in significant axis defects and in a kinked notochord in a dose-dependent manner (Fig. 1B–F). This was accompanied by aberrant somite shapes, highlighted by the lack of chevron-shaped somites and shortened body length (Fig. 1B). Embryos injected with *enpp2* mRNA at increasing concentrations exhibited dose-dependent penetrance of phenotypes (25 pg: 19.3 ± 6.8%; 50 pg: 25.6 ± 9.6%; 100 pg: 60.7 ± 2.7%; 200 pg: 60.5 ± 4.3%, Fig. 1D). These phenotypes suggest midline and axis defects during embryogenesis. To examine this further, we performed WISH to assess the expression patterns of midline markers *shha* and *ntl* to mark the notochord of the zebrafish embryo at 10 somite stage and 24 hours post fertilization, respectively (Fig. 1E,F). The majority of enpp2-overexpressing embryos displayed abnormal pattern of *shha* and *ntl* expression, characterized by kinked, patchy or expanded patterns, or expression in multiple buds (duplicated), which indicated a notochord defect. This finding suggests a role of enpp2 in regulating the midline formation and its impact on the expression of the midline axis genes *shha* and *ntl*.

**Enpp2 overexpression induces L-R patterning defect.** Establishment of the midline is important for L-R asymmetry and loss of enpp2/LPA signaling has been linked to asymmetry phenotypes<sup>31,35–37</sup>. To establish if increased enpp2/LPA signaling alters the L-R patterning of the zebrafish embryo, we examined the expression of the nodal-related asymmetric genes, *lefty1/2* and *southpaw* (*spaw*) in the lateral plate mesoderm (LPM) during somite stages in control wild type and *enpp2*-overexpressing zebrafish. As shown in Fig. 2A–D, *lefty1/2* and *spaw* were mainly expressed on the left side of the LPM during mid-late somitogenesis (15–21ss) in the control wild type zebrafish, which is consistent with previous reports<sup>38,39</sup>. However, following *enpp2* overexpression, the zebrafish displayed a more randomized expression pattern of these genes, either right-sided, absent, or bilaterally expressed in the LPM compared to control (Fig. 2A–D). In addition, the *spaw* expression domain was shifted posteriorly in some of the *enpp2* overexpression embryos suggesting that migration of the lateral plate mesoderm is delayed or perturbed (Fig. 2A–D). Taken together, this demonstrated that *enpp2* overexpression modulates L-R patterning and maintenance of the expression of the nodal-related asymmetric genes *lefty1/2* and *spaw*.

**Enpp2 modulates the organogenesis and ciliogenesis of the KV.** In the zebrafish, the Kupffer's vesicle (KV) plays an important role in the establishment of L-R patterning by maintaining the flow of nodal morphogens along the body axis<sup>40</sup>. Ciliated cells in the KV drive gradients of signalling molecules that shape the L-R asymmetry. We next investigated if the formation and morphology of the KV were altered by enpp2 overexpression. Firstly, we examined the lumen shape of the KV at the 6–8-somite stage, when the KV is formed and nodal morphogen flow occurs<sup>40</sup>. In control embryos, the KV appeared as a flattened sphere with a single round lumen, while in the overexpressed-*enpp2* fish, the lumen was smaller, misshapen or absent (Fig. 2E–G). Quantification of KV size showed that the lumen size was dramatically reduced in the enpp2 overexpressed embryos (45.00 ± 7.1%) compared to control (18.46 ± 6.80%) (Fig. 2F). To examine KV cilia, we performed live imaging using the axonemal marker *arl13b*-GFP<sup>41</sup>. Live imaging showed that the cilia lengths in the KV were significantly reduced following enpp2 overexpression (2.00 ± 0.04 units), compared to control (2.80 ± 1.44, *p* < 0.001, *n* > 200, Fig. 2H,I). This data indicates that enpp2 regulates cilia formation. *Foxj1* is a master regulator of ciliogenesis and labels cells in the KV<sup>42,43</sup>. Analysis of *foxj1* expression in enpp2 overexpressing embryos showed de-clustered pattern of *foxj1* expression pattern (Fig. 2J). In control embryos *foxj1* expression was clustered and compacted in an ovoid-like shape; in enpp2-overexpressing embryos *foxj1* was significantly de-clustered to a linear domain (arrow, Fig. 2J).



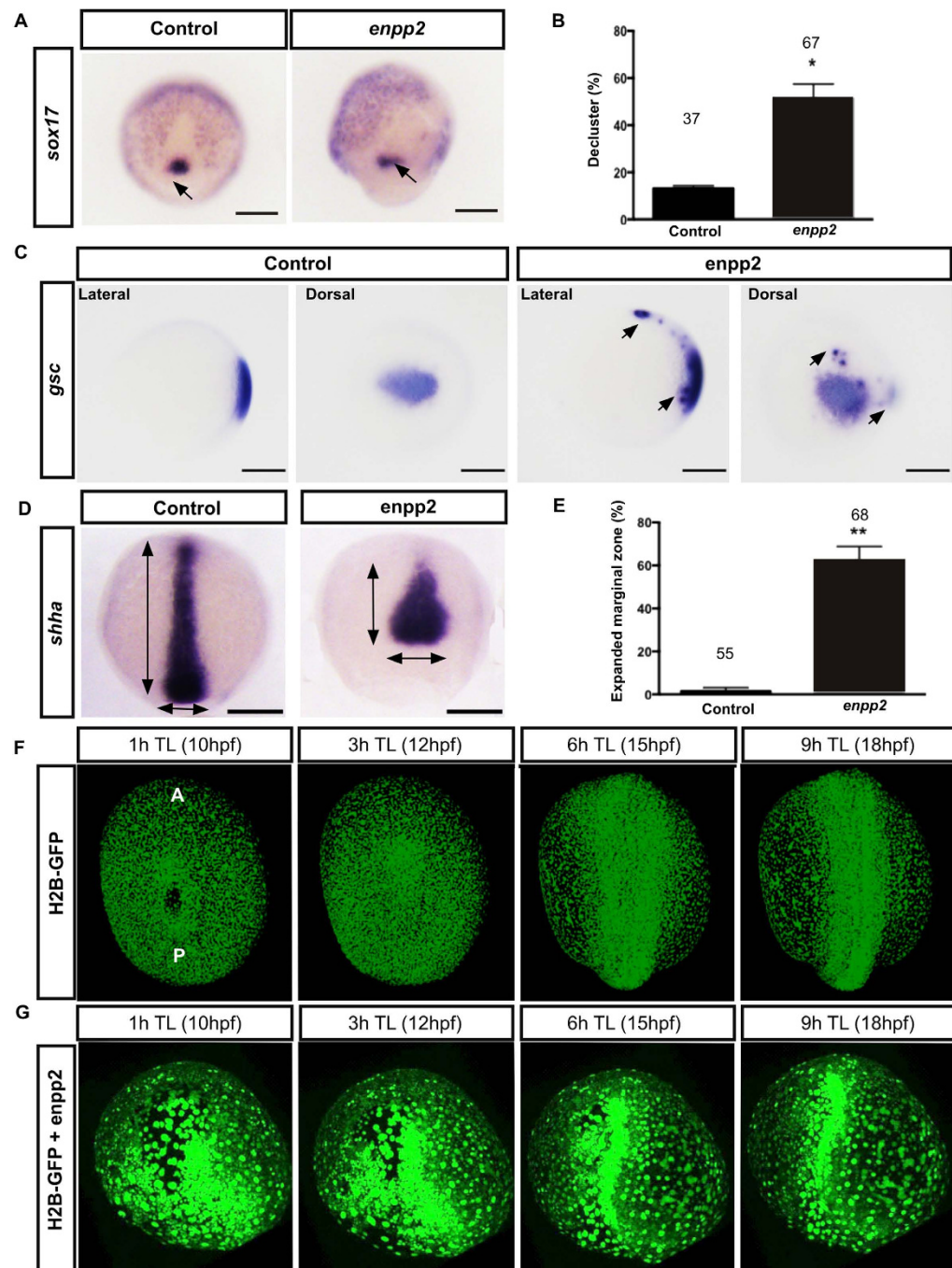
**Figure 1. *Enpp2* overexpression alters midline axis formation in the early embryogenesis of zebrafish.** Developmental series of WISH at designated stages were performed using *enpp2* antisense riboprobe. (A) Lateral view and animal/dorsal view of WISH early zebrafish embryos at designated stages. (B) Representative pictures of control, mild (slight delay and smooth somite borders) and severe (developmental delay and midline axial defect) phenotypes in *enpp2* injected embryos. (C,D) Quantification of phenotype variant and penetrance (detectable phenotype) following injection with different doses of *enpp2* RNA and normalized to control. The sample size (n) is stated as numerical value above each bar. Data are mean  $\pm$  SEM from at least four independent experiments. Statistical analysis was established by one-way ANOVA; \* $P < 0.05$ ; \*\*\* $P < 0.001$ . Dead embryos were normalised to uninjected embryos (E) Representative WISH pictures of control and *enpp2* injected embryos with midline axis gene probes *ntl* or *shha* riboprobes (F). >50 embryos used in each experiment. Scale bars: 200  $\mu$ m.



**Figure 2. Enpp2 modulates nodal-related asymmetry gene expressions, KV formation and ciliogenesis.**

Representative images of WISH of the control and *enpp2*-injected zebrafish embryos at 15–21ss with *lefty2* (A) and *spaw* (C) antisense riboprobes respectively. The midline/notochord is highlighted by *ntl* expression. The determination of left, bilateral, absence, and right groups was based on the location of *lefty2* or *spaw* expression compared to *ntl* (midline). The quantification of *lefty2* and *spaw* expression distribution embryos is shown in (B) and (D) respectively. (B) The proportion of *lefty2* expression on the left, bilateral, absence and right side are 67.3%, 4.5%, 28.2%, and 0% respectively in the control embryos and 54%, 12.6%, 28.7%, and 4.6% respectively in the *enpp2* overexpressed embryos. (D) The proportion of *spaw* expression on the left, bilateral, absence and right side are 86%, 4%, 8%, and 2% respectively in the control embryos and 61.3%, 21.8%, 12.7%, 4.2% respectively in the *enpp2*-overexpressed embryos. (E) Representative images of the KV morphology in control- and in *enpp2* overexpressing embryos (5–8ss). (F) Quantification KV size in *enpp2* overexpressed embryos compared to control. (G) Whole-mount antibody staining of zona occludens 1 (ZO-1) that labels cell junctions and outlines the KV in control *enpp2* injected embryos. (H) Representative confocal Z-stack images of cilia in the KV, marked with *arl13b* GFP, highlighting cilia morphology and distribution in control- and *enpp2* overexpressing embryos at 5–8ss. (I) Quantification of the cilia length between control- and *enpp2* overexpressing embryos. The sample size (n) is stated as numerical value above each bar. Data are mean  $\pm$  SEM. Statistical analysis was established by t-test; \*\*\* $P < 0.001$ . n indicated above bars. White circles highlight the KV lumen. (J) *Enpp2* overexpression altered the expression pattern of *foxj1*, a master regulator of ciliogenesis, observed at 90% epiboly embryo.

**Enpp2 overexpression induces defects in the midline axis and in the KV organogenesis.** The KV formation is dependent on the correct migration and clustering of distinct precursor cells, the dorsal fore runner cells (DFC), during mid-gastrulation (75% epiboly stage)<sup>44,45</sup>. The DFC express *sox17* during early embryogenesis<sup>45</sup>. In *enpp2* overexpressing embryos, the expression of *sox17* is readily observed, indicating that the specification of the DFC is not altered. However, the cluster formation of the DFC is significantly impaired in *enpp2* overexpressing embryos ( $51.2 \pm 6.2\%$ ) compared to control ( $13.1 \pm 1.1\%$ ). *Enpp2* overexpressing displayed an altered pattern of *sox17* expression (Fig. 3A,B), similar to the one observed with *foxj1* expression (Fig. 2F). The disorganisation of the DFC could lead to reduced or absent of KV formation. By measuring and observing the KV at stage 6–8ss, we detected a smaller or absent KV in *enpp2* overexpressing embryos (Fig. 2E). The de-clustered



**Figure 3. Enpp2 overexpression induces alteration in cellular migration, the expression pattern of KV precursors, dorsal organizer and midline genes (*gsc* and *shha*) during early embryogenesis.** (A) Representative images of WISH with *sox17* riboprobe to mark the integrity of the DFC cluster at 75% epiboly. (B) Quantification of KV defect and/alteration of DFC cluster, as highlighted by *sox17* WISH. (C) The *gsc* expression is observed in the dorsal organizer of *enpp2* overexpressing embryos. However, the expression is also detected in the axial mesoderm precursors suggesting that convergence and cell migration during midline formation is impaired. (D) Representative images of WISH of *shha* at 90% epiboly showing a truncated axis and laterally expanded expression of *shha* at the blastoderm margin. (E) Quantification of the broadened *shha* expression domain at the blastoderm margin in control and *enpp2* overexpressing embryos at 90% epiboly. (F,G) Representative images of time lapse series showing cell migration during midline formation (10 hpf- early somitogenesis) in wild type and *enpp2*-injected embryos. Image stacks were taken every 15 minutes. Time course for each image is indicated. The *enpp2* overexpressed embryo failed to undergo a proper CE (G) which results in bend and broadened midline compared to wild type (F). (B,E) The sample size (n) is stated as numerical value above each bar. Data are mean  $\pm$  SEM. Statistical analysis was established by t-test; \*P < 0.05; \*\*P < 0.01. n indicated above bars.

expression of *sox17* and *foxj1* expressions suggest the observed phenotypes could be caused by the impaired cell migration during gastrulation.

Since we found the convergence and extension movements (CE) during midline formation were impaired, we next assessed if the dorsal organizer is formed correctly following *enpp2* overexpression by performing WISH using *gooseoid* (*gsc*), a dorsal organizer marker<sup>46</sup>, during the mid epiboly stage (Fig. 3C). We observed *gsc* expression in the dorsal organizer but *gsc* expression was also detected in the axial mesoderm, suggesting that precursors do not properly converge into the dorsal organizer during midline formation (Fig. 3C). Consistent with this result, the expression of the axial mesoderm marker *shha* showed expanded expression at the blastoderm margin and a shortened anterior to posterior expression pattern in *enpp2* injected embryos ( $61.96 \pm 6.8\%$  in the *enpp2* overexpressed embryo compared to  $1.58 \pm 1.6\%$  in control, Fig. 3D,E) confirming an impaired convergence extension during gastrulation. We next performed time-lapse analysis of cell movement from the end of gastrulation until early somitogenesis to observe the midline formation. Cell migration defects were detected during epiboly in *enpp2* overexpressing embryos (Fig. 3F,G, Suppl. Movie 1–2). In wild type embryos, cells intercalated and initiated the elongation during mid-line formation, the cells in the *enpp2* overexpressing embryos failed to converge and extend, resulting in a broader midline area and a bent midline formation (Fig. 3F,G, Suppl. Movie 1–2). The detected cell migration phenotypes were in agreement with the altered gene expression patterns of *gsc*, *ntl*, *spaw* and *shha* in *enpp2* overexpressing embryos.

***enpp2* modulates cell migration via *lpa*<sub>1–3</sub> Rho/ROCK.** In order to dissect the signalling mechanisms underlying LPA's effect in development, we first examined the expression profile of its receptors (*lpa*<sub>1–3</sub>) in the early embryo by WISH. We detected expression of *lpa*<sub>1–3</sub> in the early embryo (Suppl. Fig. 2 and data not shown). We found that *lpa*<sub>1–3</sub> were ubiquitously expressed at the animal portion and at the margin between cells and yolk portion (yolk syncytial layer) during blastula until onset of gastrulation. During the late gastrulation and early segmentation stages, *lpa*<sub>1</sub> was expressed in the mesoderm tissue adjacent to the body axis, while *lpa*<sub>2–3</sub> were expressed along the midline during zebrafish (Suppl. Fig. 2). Taken together, the expression of *lpa*<sub>1–3</sub> suggests that these receptors may play an important role during gastrulation and the midline formation.

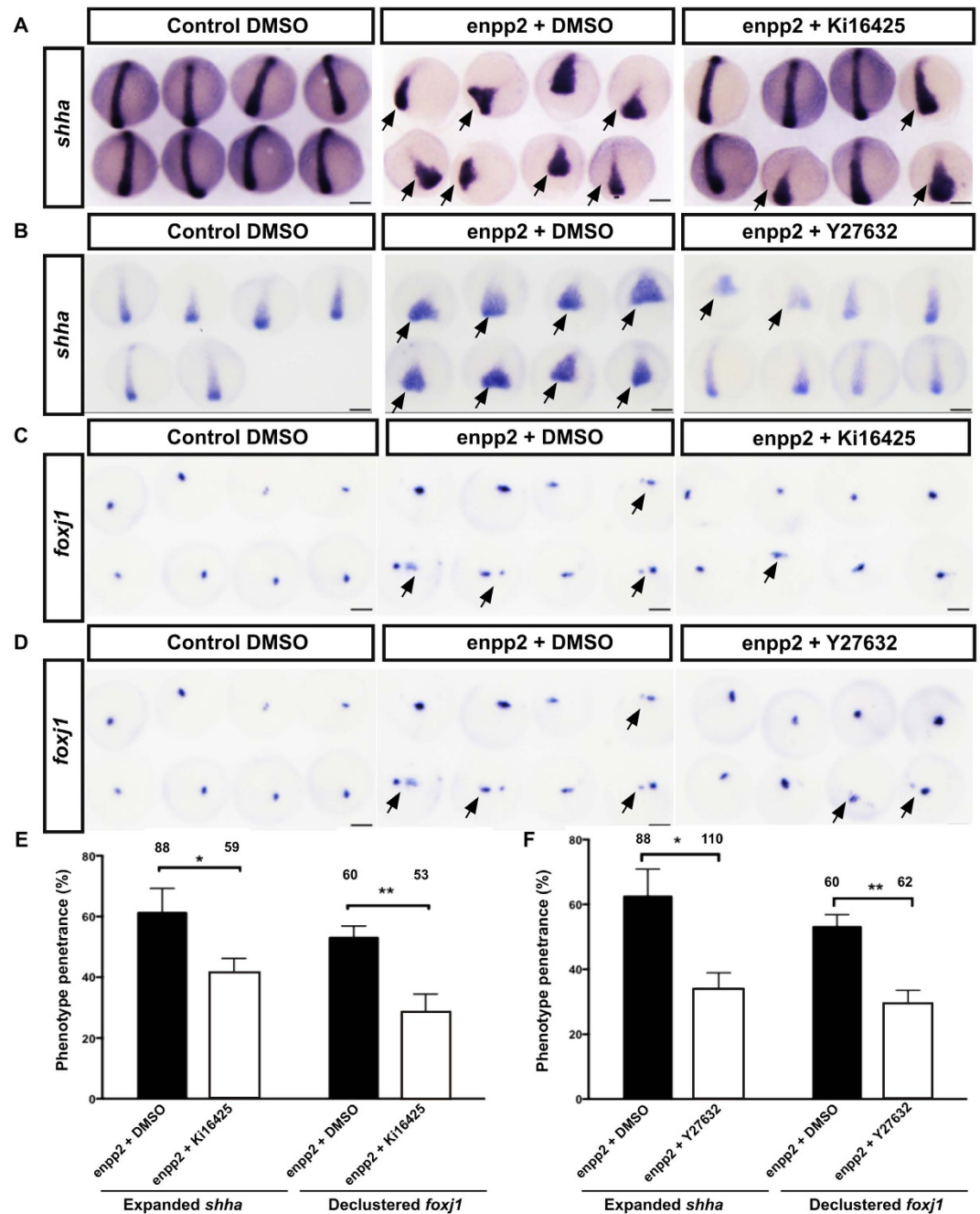
We next addressed the role played by LPA receptors in the phenotypes observed after *enpp2*-overexpression. To this end we performed rescue experiments using the well-established LPA receptor (*lpa*<sub>1–3</sub>) antagonist Ki16425<sup>47</sup>. Ki16425 is a specific *lpa* receptor antagonist that efficiently antagonize *lpa*<sub>1–3</sub> but not *lpa*<sub>4</sub>, *lpa*<sub>6a</sub>, and *lpa*<sub>6b</sub> and it has been used to antagonise LPA signalling in zebrafish previously<sup>34</sup>. We incubated *enpp2*-overexpressing embryos immediately after RNA injection with increasing concentrations of Ki16425 and assessed the impact on phenotypes by using *shha* and *foxj1* expression as a read-out at the end of gastrulation (Fig. 4). *enpp2* injected embryos treated with Ki16425 showed a significant reduction of *enpp2*-induced CE and ciliogenesis phenotypes in a dose-dependent manner (Fig. 4A–C). Following *enpp2* overexpression, the CE phenotype characterised by a short axis and a broadened *shha* expression was detected in  $66.5 \pm 7.8\%$  of the injected embryos (Fig. 4E). However, following and Ki16425 treatment, the number of embryos displaying this phenotype decreased to  $41.3 \pm 4.9\%$  (Fig. 4E). Similarly, declustered *foxj1* expression was observed in  $53 \pm 3.8\%$  of the vehicle control embryos but decreased to  $28.3 \pm 6.2\%$  in Ki16425-treated embryos (Fig. 4E). The rescue effect was maintained until later stages of development as we observed significantly reduced morphological phenotypes and almost normal expression levels of *ntl*, *shha* and its downstream gene *gli2* following Ki16425 treatments at 24 hours post fertilisation (hpf, Fig. 2, Suppl. Fig. 3).

Rho/ROCK is one of the common downstream pathways of *enpp2*/LPA signalling and it plays an essential role in regulating cell shape and migration through actin stabilization of the cytoskeleton<sup>16,48</sup>. Furthermore, the Rho/ROCK pathway is an important regulator of CE cell movement during midline formation<sup>49,50</sup>. We inhibited the Rho/ROCK pathway by using a selective inhibitor of the Rho-associated protein kinase p160 ROCK, Y27632<sup>51</sup>. Inhibiting Rho/ROCK after *enpp2* overexpression significantly reversed the observed phenotypes. The penetrance and severity of CE and clustering of DFC were significantly reduced after Y27632 treatment (Fig. 4B,D,F). Y27632 treatment reduced the expanded *shh* expression phenotype from  $62.4 \pm 8.5\%$  in the vehicle control to  $33.9 \pm 4.9\%$  in treated embryos (Fig. 4F). The *foxj1* expression pattern was  $53 \pm 3.8\%$  in the vehicle control and decreased to  $29.2 \pm 4.3\%$  in Y27632-treated embryos (Fig. 4F). The rescue effect of Y27632 in *enpp2* injected embryos was maintained until later stages of development as we observed significantly reduced morphological phenotypes and almost normal expression patterns of *ntl* and *spaw* at 24 hpf (Suppl. Fig. 3). Taken together this data indicates that *enpp2* induces the midline axis phenotype through activation of the Rho/ROCK pathway in a *lpa*<sub>1–3</sub> mediated manner.

## Discussion

The role of LPA signalling in early embryonic development is limited<sup>19,22–24,31</sup>. Here we found a role of *enpp2*/LPA signalling in cell migration during gastrulation, which interfered with the midline axis formation and the establishment of the L-R asymmetry.

The establishment of the axial/midline mesoderm and the L-R asymmetry are crucial to the formation of the body plan in the zebrafish embryo. The body axis arises from the dorsal organizer shield<sup>6,52,53</sup>. Midline formation involves a complex morphogenetic movement, called the convergence and extension movement. During this process, the axial mesoderm cells migrate together and intercalate (converge) towards the dorsal organizer and then migrate towards the anterior and posterior poles (elongation) to give rise to the future midline<sup>4,5,54</sup>. *Enpp2* is expressed at the blastula stage in the zebrafish and it is highly expressed in the margin and midline during gastrulation and somitogenesis respectively. During epiboly, *enpp2* is expressed in the YSL and it is later expressed in the dorsal organizer suggesting a role in tissue patterning. *lpa*<sub>1–3</sub> are also expressed in the embryo during gastrulation. During early gastrulation *lpa*<sub>1</sub> is expressed at the dorsal organizer shield, where the CE occurs to form the body axis<sup>54</sup>. At the end of gastrulation *lpa*<sub>1</sub> expression is found in the mesoderm adjacent to the midline. In



**Figure 4. Pharmacological blocking of  $lpa_{1-3}$  or Rho/ROCK signalling rescue enpp2 overexpression induced midline phenotype.** Representative WISH images of *shha* (A,B) and *foxj1* expression (C,D) at 90% epiboly of enpp2 overexpressing embryos treated with the  $lpa_{1-3}$  antagonist Ki16425 (A,C) and Rho/ROCK signalling inhibitor Y27632 (B,D). Enpp2 overexpressing embryos display expanded expression of *shha* (A,B) and *foxj1* (C,D) illustrated by pointed in arrows. Quantifications of the of *shha* expression domain in enpp2 overexpressing embryo following vehicle or Ki16425 (E) or Y27632 treatment (F). The phenotype penetrance following enpp2 injection and Ki16425 treatment and Y27632 were measured at 90% epiboly (E,F). (E,F) The sample size (n) is stated as numerical value above each bar from at least two independent experiments. Data are mean  $\pm$  SEM. Statistical analysis was established by t-test; \*P < 0.05; \*\*P < 0.01. n indicated above bars.

contrast,  $lpa_{2b-3}$  are expressed at the midline and at the end of gastrulation. These expression patterns suggest that enpp2/LPA signalling is involved in epiboly and gastrulation. To assess the role of enpp2 in the zebrafish embryogenesis, we overexpressed *enpp2* mRNA and assessed the embryonic phenotype at different stages of the zebrafish development. We observed an alteration of cell migration towards the midline during gastrulation. Cells failed to complete the CE e.g. to migrate towards the dorsal organizers and to elongate to form the midline, resulting in perturbed expression of midline and lateral plate mesoderm genes (*shha* *ntl* and *spaw*), bent body axis and shortened body length. The impaired cell migration during CE is likely responsible for the expanded

expression of midline genes, bend/kinked midline in the *enpp2*-overexpressing embryo and the shortened body plan at later stages of development. The Rho/ROCK pathway is a common and a likely downstream pathway of the *enpp2*/LPA signalling axis, and particularly through multiple LPA G-protein coupled receptors, including the *lpa*<sub>1-6</sub><sup>16,48</sup>. Rho/ROCK is a key regulator of cell shape and migration through its action on actin stabilization of cytoskeleton<sup>49</sup>. Rho/ROCK pathway is an important regulator of CE process during midline formation and regulates left-right asymmetry<sup>49,50</sup>. Indeed ROCK has been previously shown to play an obligatory role in the morphogenetic movement during embryonic organogenesis. Inhibiting ROCK with Y27632 in the early chick embryo blocked cell migration and fusion of the bilateral heart primordia, axis formation, movement of Hensen's node (similar to KV), and establishment of L-R asymmetry<sup>50</sup>. We demonstrated that the *enpp2* overexpression midline defect phenotype could be rescued by blocking ROCK, suggesting that the phenotype is mediated by Rho/ROCK. In agreement with our findings, the Rho/ROCK pathway regulates CE cell movement during gastrulation<sup>49,55</sup>. *rock2b* in zebrafish is specifically expressed in DFC and KV and later in midline<sup>56</sup>. Knock-down of *rock2b* function using morpholinos leads to altered anterior posterior patterning, random distribution of ciliated cells and loss of L-R symmetry in the KV<sup>56</sup>. Similarly, loss of *Rock2* function in mice alters organ laterality and asymmetry gene expression including the heart, hypochord and notochord<sup>50</sup>. Interestingly, over activation of sphingosine-1-phosphate (S1P) signalling via its cognate G protein-coupled receptor S1pr2 results in a very similar endoderm convergence phenotype as *enpp2*<sup>57</sup>. This phenotype is mediated via a RhoGEF-dependent pathway<sup>57</sup> and implies shared downstream mechanisms. Rho/ROCK is also downstream of the planar cell polarity (PCP) pathway that is a well-known regulator of CE during gastrulation. The vertebrate PCP pathway is a non-canonical Wnt signalling cascade that can modulate the actin cytoskeleton via small GTPases including Rho<sup>58</sup> to control polarized cell behaviours such as CE movements and orientation of stereocilia in the inner ear<sup>49,59-62</sup>. An intriguing possibility is that LPA signalling interacts with the PCP pathway via the extracellular matrix. *Enpp2* has several positively charged surface residues and it may interact with heparin sulphate proteoglycans like glypicans that are well known regulators of the PCP pathway<sup>63,64</sup>.

Furthermore, overexpression of *enpp2* altered the migration and clustering of dorsal fore runner cells. This leads to perturbed formation of the KV and ciliogenesis, and subsequently to a failure to establish a correct L-R asymmetry in the embryo. Overexpression of *enpp2* could result in effects independent of LPA signalling. However, the efficient pharmacological rescue with the specific LPA receptor (*Lpa*<sub>1-3</sub>) antagonist Ki16425 suggests that the phenotypes observed are mediated by LPA receptors. Furthermore, our findings are consistent with the previously reported roles of *enpp2*/LPA signalling during L-R patterning in mice and zebrafish. Loss of *enpp2* in mice leads to phenotypes such as: axial mesoderm/notochord defects, and abnormal organ positioning<sup>29,65</sup>, and are associated with abnormal L-R patterning<sup>66</sup>. Furthermore, knock-down of *enpp2* or *lpa*<sub>3</sub> in zebrafish using morpholinos results in altered KV formation, ciliogenesis and L-R asymmetry<sup>31</sup>. In addition, over activation of *enpp2*/LPA signalling induces cardiac bifida in zebrafish<sup>32</sup>. Cardiac bifida is caused by the failure of bilateral mesoderm to migrate and coalesce into a single central heart tube<sup>67</sup>. Similar to *enpp2*/LPA signaling, over activation of sphingosine-1-phosphate (S1P) signalling via its cognate G protein-coupled receptor S1pr2 impairs endoderm convergence and results in cardiac bifida<sup>57</sup>. Taken together, available data suggests that LPA signaling needs to be tightly regulated during multiple stages of embryonic development.

## Conclusion

Midline formation and establishment of L-R symmetry is crucial in the embryonic development. By overexpressing *enpp2* in the zebrafish embryo, our study for the first time shows the unique role of *enpp2*/LPA in regulating mesendoderm cell migration in the establishment of axial midline and L-R asymmetry in the embryogenesis through Rho/ROCK pathway via *lpa*<sub>1-3</sub>.

## Material and Methods

All experimental work performed in this study was approved by the Monash Animal Ethics Committee (MARF/2013/096), in accordance with the requirements of the National Health & Medical Research Council of Australia.

**Zebrafish.** Zebrafish embryos were collected immediately after fertilization, maintained at 28.5 °C, and staged by developmental time (hours post-fertilization, hpf) using morphological criteria<sup>68</sup>. The Tübingen was used as a wild type strain. To perform *in situ*, the embryos at the designated stages were fixed in 4% PFA (2 hrs RT (<24 hpf) or overnight 4 °C), washed with PBS-0.1% Tween (PBS-T, 5 min, twice), dechorionated, dehydrated with MeOH (5 min RT, twice) and stored in MeOH at -20 °C until processed for *in situ*.

**Microinjection and phenotypic analysis.** The embryos were injected with 1 nl of the corresponding RNA at different concentrations. Full-length *enpp2/atx* constructs (25, 50, 75, 100, 150 and 200 pg) or *Arl13b* GFP mRNA (50–100 pg) or H2B (75 pg) were injected into the 1–4 cell stage embryos. Phenotype penetrances and mortality rates were quantified within 9–24 hrs post injection. For assessment of Kupffer's vesicle (KV), embryos were examined at the 5–8 somite stages.

**Quantitative Polymerase Chain Reaction (qPCR).** qPCR was carried out using TaqMan Universal master mix (Applied Biosystems, Foster City, CA) and the 7900HT Fast Real-Time PCR system (Applied Biosystems) and SYBR Green (Sigma). Primer set targeting *enpp2*: forward (TTCTCCTTCATCCTTCCACAC) and reverse (GTAAGTCCACATCTCGCAGG) (NM\_200603) were used. A total of 10 µl reaction was prepared and repeated in triplicate per test. The relative quantitation was achieved by applying the comparative CT method ( $\Delta\Delta CT$ ) whereby the mRNA levels were normalized against the level of glyceraldehyde-3-phosphate dehydrogenase (GAPDH) mRNA with *enpp2* expression at the shield stage used as the reference.

**Whole Mount *In Situ* Hybridization (WISH).** WISH was performed following the established method with minor modifications<sup>69,70</sup>. The primer sequence used for PCR amplification and cloning the following genes, *lpa1-3*, *enpp2/atx*, *shha*, *foxj1* (Suppl. Table 1). The resulting DNA fragments were subcloned into pGEMT-easy vector or pCS2+ vector; of which were digested to be transcribed into sense (*enpp2*) and/or antisense mRNA sequence/probe.

**Embryo Immersion/Drug treatment.** Following *enpp2/atx* mRNA injection, embryos were immersed in Ringer's solution containing 0.5% DMSO and supplemented with the Rho/ROCK inhibitor, Y27632 (5 and 10  $\mu$ M, Sigma-Aldrich) or *lpar1-3* antagonist, Ki16425 at different concentrations (1, 5, 7.5 and 10  $\mu$ M Sigma-Aldrich). Y27632 was used at 10  $\mu$ M and Ki16425 was used at 7.5  $\mu$ M for the rescue experiments. WT control embryos immersed with the drugs from early gastrulation and embryos did not develop detectable phenotype at 24 hpf. The embryos were then collected and fixed with 4% PFA and processed for WISH.

**Live Imaging.** To visualize the KV, wild type or over-expressed zebrafish embryos were dechorionated at 5–8ss stage and mounted in 0.8–1% of Low Melting-point Agar (LMP, Sigma). The KV was then visualized using an Olympus Dissecting microscope MVX10 using the CellSens Standard software (v8.1, Olympus). In some experiments, cilia in embryos KV were visualized under a Leica SP5 confocal microscope and the Leica application suit advanced software (v3, Leica) using a 20  $\times$  1.0 NA water immersion lens. For imaging of cilia, Z-projections of the entire KV were captured with slow acquisition with some frame averaging using *arl13b-GFP*, a live axoneme marker of cilia, to investigate cilia in the living embryo as studied in ref. 41. To image cell migration during midline axis formation, the embryos were injected with H2B together with or without *enpp2/atx* and visualized starting at the tail bud stage (10 hpf) and recorded by time lapse for 8–10 hours. The embryo was visualized every 10–15 minutes.

**Statistical analysis.** All sets of experiments were performed at least three times in triplicates, unless specified (n refers to the number of independent experiments performed on different cell cultures). Data-sets were expressed as mean  $\pm$  standard error of the mean (SEM). Significance of the differences was evaluated using the unpaired t-test (two-tailed) or the one and two-way ANOVA followed by Dunnett's multiple comparisons test in the *in vivo* experiment. Statistical significance was established at \* $p < 0.05$ , \*\* $p < 0.01$  and \*\*\* $p < 0.001$ , \*\*\*\*= $< 0.0001$ .  $\alpha = 0.5$ , using Prism (v.6, GraphPad).

## References

- Cleaver, O., Seufert, D. & Krieg, P. Endoderm patterning by the notochord: development of the hypochord in *Xenopus*. *Development (Cambridge, England)* **127**, 869–879 (2000).
- Isogai, S., Lawson, N. D., Torrealday, S., Horiguchi, M. & Weinstein, B. M. Angiogenic network formation in the developing vertebrate trunk. *Development (Cambridge, England)* **130**, 5281–5290, doi: 10.1242/dev.00733 (2003).
- Lawson, N. D., Vogel, A. M. & Weinstein, B. M. *sonic hedgehog* and vascular endothelial growth factor Act Upstream of the Notch Pathway during Arterial Endothelial Differentiation. *Developmental cell* **3**, 127–136 (2002).
- Tada, M. & Heisenberg, C.-P. Convergent extension: using collective cell migration and cell intercalation to shape embryos. *Development (Cambridge, England)* **139**, 3897–3904, doi: 10.1242/dev.073007 (2012).
- Schier, A. & Talbot, W. Molecular genetics of axis formation in zebrafish. *Annu. Rev. Genet.* (2005).
- Saúde, L., Woolley, K., Martin, P., Driever, W. & Stemple, D. Axis-inducing activities and cell fates of the zebrafish organizer. *Development (Cambridge, England)* **127**, 3407–3417 (2000).
- Odenthal, J. *et al.* Mutations affecting the formation of the notochord in the zebrafish, *Danio rerio*. *Development (Cambridge, England)* **123**, 103–115 (1996).
- Talbot, W. *et al.* A homeobox gene essential for zebrafish notochord development. *Nature* **378**, 150–157, doi: 10.1038/378150a0 (1995).
- Lenhart, K. F., Lin, S. Y., Titus, T. A., Postlethwait, J. H. & Burdine, R. D. Two additional midline barriers function with midline lefty1 expression to maintain asymmetric Nodal signaling during left-right axis specification in zebrafish. *Development (Cambridge, England)* **138**, 4405–4410, doi: 10.1242/dev.071092 (2011).
- Macdonald, R. *et al.* Midline signalling is required for Pax gene regulation and patterning of the eyes. *Development (Cambridge, England)* **121**, 3267–3278 (1995).
- Montero, J.-A., Kilian, B., Chan, J., Bayliss, P. E. & Heisenberg, C.-P. Phosphoinositide 3-kinase is required for process outgrowth and cell polarization of gastrulating mesendodermal cells. *Current biology* **13**, 1279–1289 (2003).
- Kai, M., Heisenberg, C.-P. & Tada, M. Sphingosine-1-phosphate receptors regulate individual cell behaviours underlying the directed migration of prechordal plate progenitor cells during zebrafish gastrulation. *Development (Cambridge, England)* **135**, 3043–3051, doi: 10.1242/dev.020396 (2008).
- Ulrich, F., Krieg, M., Schötz, E., Link, V., Castanon, I. *et al.* Wnt11 functions in gastrulation by controlling cell cohesion through Rab5c and E-cadherin. *Developmental cell* (2005).
- Myers, D. C., Sepich, D. S. & Solnica-Krezel, L. Bmp activity gradient regulates convergent extension during zebrafish gastrulation. *Developmental biology* **243**, 81–98 (2002).
- Fürthauer, M., Thisse, C. & Thisse, B. A role for FGF-8 in the dorsoventral patterning of the zebrafish gastrula. *Development (Cambridge, England)* **124**, 4253–4264 (1997).
- Frisca, F., S. R., Goldshmit, Y. & Pébay, A. Biological effects of lysophosphatidic acid in the nervous system. *Int Rev Cell Mol Biol* **296**, 273–322 (2012).
- Choi, J. W. *et al.* LPA Receptors: Subtypes and Biological Actions. *Annual Review of Pharmacology and Toxicology* **50**, 157–186, doi: 10.1146/annurev.pharmtox.010909.105753 (2010).
- Kingsbury, M. A., Rehen, S. K., Contos, J. J., Higgins, C. M. & Chun, J. Non-proliferative effects of lysophosphatidic acid enhance cortical growth and folding. *Nat Neurosci* **6**, 1292–1299 (2003).
- Contos, J. J. A., Fukushima, N., Weiner, J. A., Kaushal, D. & Chun, J. Requirement for the *lpa1* lysophosphatidic acid receptor gene in normal suckling behavior. *Proceedings of the National Academy of Sciences of the United States of America* **97**, 13384–13389 (2000).
- Yung, Y. C. *et al.* Lysophosphatidic Acid Signaling May Initiate Fetal Hydrocephalus. *Science Translational Medicine* **3**, 99ra87, doi: 10.1126/scitranslmed.3002095 (2011).
- Estivill-Torres, G. *et al.* Absence of LPA1 signaling results in defective cortical development. *Cereb Cortex* **18**, 938–950, doi: bhm132 [pii]10.1093/cercor/bhm132 (2008).

22. Sumida, H. *et al.* LPA4 regulates blood and lymphatic vessel formation during mouse embryogenesis. *Blood* **116**, 5060–5070, doi: 10.1182/blood-2010-03-272443 (2010).
23. Contos, J. J. A. *et al.* Characterization of lpa2 (Edg4) and lpa1/lpa2 (Edg2/Edg4) Lysophosphatidic Acid Receptor Knockout Mice: Signaling Deficits without Obvious Phenotypic Abnormality Attributable to lpa2. *Mol. Cell. Biol.* **22**, 6921–6929, doi: 10.1128/mcb.22.19.6921-6929.2002 (2002).
24. Ye, X. *et al.* LPA3-mediated lysophosphatidic acid signalling in embryo implantation and spacing. *Nature* **435**, 104–108, doi: [http://www.nature.com/nature/journal/v435/n7038/supinfo/nature03505\\_S1.html](http://www.nature.com/nature/journal/v435/n7038/supinfo/nature03505_S1.html) (2005).
25. Tigyi, G. Aiming drug discovery at lysophosphatidic acid targets. *British Journal of Pharmacology* **161**, 241–270, doi: 10.1111/j.1476-5381.2010.00815.x (2010).
26. Geach, T. J. *et al.* An essential role for LPA signalling in telencephalon development. *Development (Cambridge, England)* **141**, 940–949, doi: 10.1242/dev.104901 (2014).
27. Tokumura, A. *et al.* Identification of Human Plasma Lysophospholipase D, a Lysophosphatidic Acid-producing Enzyme, as Autotaxin, a Multifunctional Phosphodiesterase. *Journal of Biological Chemistry* **277**, 39436–39442, doi: 10.1074/jbc.M205623200 (2002).
28. Umezū-Goto, M. *et al.* Autotaxin has lysophospholipase D activity leading to tumor cell growth and motility by lysophosphatidic acid production. *The Journal of Cell Biology* **158**, 227–233, doi: 10.1083/jcb.200204026 (2002).
29. van Meeteren, L. A. *et al.* Autotaxin, a secreted lysophospholipase D, is essential for blood vessel formation during development. *Mol Cell Biol* **26**, 5015–5022, doi: 10.1128/MCB.02419-05 (2006).
30. Tanaka, M. *et al.* Autotaxin stabilizes blood vessels and is required for embryonic vasculature by producing lysophosphatidic acid. *The Journal of biological chemistry* **281**, 25822–25830, doi: 10.1074/jbc.M605142200 (2006).
31. Lai, S.-L. *et al.* Autotaxin/Lpar3 signaling regulates Kupffer's vesicle formation and left-right asymmetry in zebrafish. *Development (Cambridge, England)* **139**, 4439–4448, doi: 10.1242/dev.081745 (2012).
32. Nakanaga, K. *et al.* Overexpression of autotaxin, a lysophosphatidic acid-producing enzyme, enhances cardia bifida induced by hypo-sphingosine-1-phosphate signaling in zebrafish embryo. *Journal of Biochemistry* **155**, 235–241, doi: 10.1093/jb/mvt114 (2014).
33. Yüelling, L., Waggner, C., Afshari, F., Lister, J. & Fuss, B. Autotaxin/ENPP2 regulates oligodendrocyte differentiation *in vivo* in the developing zebrafish hindbrain. *Glia* **60**, 1605–1618, doi: 10.1002/glia.22381 (2012).
34. Yukiura, H. *et al.* Autotaxin regulates vascular development via multiple lysophosphatidic acid (LPA) receptors in zebrafish. *The Journal of biological chemistry* **286**, 43972–43983, doi: 10.1074/jbc.M111.301093 (2011).
35. Bisgrove, B., Essner, J. & Yost, H. Multiple pathways in the midline regulate concordant brain, heart and gut left-right asymmetry. *Development (Cambridge, England)* **127**, 3567–3579 (2000).
36. Danos, M. & Yost, H. Role of notochord in specification of cardiac left-right orientation in zebrafish and *Xenopus*. *Developmental biology* **177**, 96–103, doi: 10.1006/dbio.1996.0148 (1996).
37. Meno, C. *et al.* lefty-1 is required for left-right determination as a regulator of lefty-2 and nodal. *Cell* **94**, 287–297, doi: 10.1016/S0092-8674(00)81472-5 (1998).
38. Hirokawa, N., Tanaka, Y., Okada, Y. & Takeda, S. Nodal flow and the generation of left-right asymmetry. *Cell* **125**, 33–45 (2006).
39. Mercola, M. & Levin, M. Left-right asymmetry determination in vertebrates. *Annual Review of cell and developmental Biology* **17**, 779–805 (2001).
40. Essner, J., Amack, J., Nyholm, M., Harris, E. & Yost, H. Kupffer's vesicle is a ciliated organ of asymmetry in the zebrafish embryo that initiates left-right development of the brain, heart and gut. *Development (Cambridge, England)* **132**, 1247–1260, doi: 10.1242/dev.01663 (2005).
41. Borovina, A., Superina, S., Voskas, D. & Ciruna, B. Vangl2 directs the posterior tilting and asymmetric localization of motile primary cilia. *Nature cell biology* **12**, 407–412, doi: 10.1038/ncb2042 (2010).
42. Stubbs, J., Oishi, I., Izpisua Belmonte, J. & Kintner, C. The forkhead protein Foxj1 specifies node-like cilia in *Xenopus* and zebrafish embryos. *Nature genetics* **40**, 1454–1460, doi: 10.1038/ng.267 (2008).
43. Yu, X., Ng, C., Habacher, H. & Roy, S. Foxj1 transcription factors are master regulators of the motile ciliogenic program. *Nature genetics* **40**, 1445–1453, doi: 10.1038/ng.263 (2008).
44. Cooper, M. & D'Amico, L. A cluster of noninvoluting endocytic cells at the margin of the zebrafish blastoderm marks the site of embryonic shield formation. *Developmental biology* **180**, 184–198, doi: 10.1006/dbio.1996.0294 (1996).
45. Aamar, E. & Dawid, I. Sox17 and chordin are required for formation of Kupffer's vesicle and left-right asymmetry determination in zebrafish. *Developmental Dynamics*, doi: 10.1002/dvdy.22431 (2010).
46. Schulte-Merker, S. *et al.* Expression of zebrafish goosecoid and no tail gene products in wild-type and mutant no tail embryos. *Development (Cambridge, England)* **120**, 843–852 (1994).
47. Ohta, H. *et al.* Ki16425, a subtype-selective antagonist for EDG-family lysophosphatidic acid receptors. *Mol Pharmacol* **64**, 994–1005, doi: 10.1124/mol.64.4.994 (2003).
48. Sriwai, W., Zhou, H. & Murthy, K. G(q)-dependent signalling by the lysophosphatidic acid receptor LPA(3) in gastric smooth muscle: reciprocal regulation of MYPT1 phosphorylation by Rho kinase and cAMP-independent PKA. *The Biochemical journal* **411**, 543–551, doi: 10.1042/bj20071299 (2008).
49. Marlow, F., Topczewski, J., Sepich, D. & Solnica-Krezel, L. Zebrafish Rho Kinase 2 Acts Downstream of Wnt11 to Mediate Cell Polarity and Effective Convergence and Extension Movements. *Current Biology* **12**, 876–884 (2002).
50. Wei, L. *et al.* Rho kinases play an obligatory role in vertebrate embryonic organogenesis. *Development (Cambridge, England)* **128**, 2953–2962 (2001).
51. Davies, S., Reddy, H., Caivano, M. & Cohen, P. Specificity and mechanism of action of some commonly used protein kinase inhibitors. *Biochem. J* **351**, 95–105 (2000).
52. Spemann, H. & Mangold, H. Induction of embryonic primordia by implantation of organizers from a different species. 1923. *The International journal of developmental biology* **45**, 13–38 (2000).
53. Shih, J. & Fraser, S. E. Characterizing the zebrafish organizer: microsurgical analysis at the early-shield stage. *Development (Cambridge, England)* **122**, 1313–1322 (1996).
54. Warga, R. & Kimmel, C. Cell movements during epiboly and gastrulation in zebrafish. *Development (Cambridge, England)* (1990).
55. Topczewski, J. *et al.* The zebrafish glypican knypek controls cell polarity during gastrulation movements of convergent extension. *Developmental cell* **1**, 251–264, doi: 10.1016/S1534-5807(01)00005-3 (2001).
56. Wang, G. *et al.* The Rho kinase Rock2b establishes anteroposterior asymmetry of the ciliated Kupffer's vesicle in zebrafish. *Development (Cambridge, England)* **138**, 45–54, doi: 10.1242/dev.052985 (2011).
57. Ye, D. & Lin, F. S1pr2/Galpha13 signaling controls myocardial migration by regulating endoderm convergence. *Development (Cambridge, England)* **140**, 789–799, doi: 10.1242/dev.085340 (2013).
58. Schlessinger, K., Hall, A. & Tolwinski, N. Wnt signaling pathways meet Rho GTPases. *Genes & development*, doi: 10.1101/gad.1760809 (2009).
59. Winter, C. G. *et al.* Drosophila Rho-associated kinase (Drok) links Frizzled-mediated planar cell polarity signaling to the actin cytoskeleton. *Cell* **105**, 81–91 (2001).
60. Zhu, S., Liu, L., Korzh, V., Gong, Z. & Low, B. RhoA acts downstream of Wnt5 and Wnt11 to regulate convergence and extension movements by involving effectors Rho kinase and Diaphanous: use of zebrafish as .... *Cellular signalling* (2006).

61. Wang, Y. & Nathans, J. Tissue/planar cell polarity in vertebrates: new insights and new questions. *Development (Cambridge, England)* **134**, 647–658, doi: 10.1242/dev.02772 (2007).
62. Kim, G.-H. & Han, J.-K. JNK and ROK $\alpha$  function in the noncanonical Wnt/RhoA signaling pathway to regulate *Xenopus* convergent extension movements. *Developmental Dynamics* **232**, 958–968, doi: 10.1002/dvdy.20262 (2005).
63. Moolenaar, W. & Perrakis, A. Insights into autotaxin: how to produce and present a lipid mediator. *Nature reviews. Molecular cell biology* **12**, 674–679, doi: 10.1038/nrm3188 (2011).
64. Ohkawara, B., Yamamoto, T. S., Tada, M. & Ueno, N. Role of glypican 4 in the regulation of convergent extension movements during gastrulation in *Xenopus laevis*. *Development (Cambridge, England)* **130**, 2129–2138 (2003).
65. Koike, S. *et al.* Autotaxin is required for the cranial neural tube closure and establishment of the midbrain-hindbrain boundary during mouse development. *Developmental dynamics: an official publication of the American Association of Anatomists* **240**, 413–421, doi: 10.1002/dvdy.22543 (2011).
66. Bisgrove, B., Morelli, S. & Yost, H. Genetics of human laterality disorders: insights from vertebrate model systems. ... and human genetics (2003).
67. Compernelle, V. *et al.* Cardia bifida, defective heart development and abnormal neural crest migration in embryos lacking hypoxia-inducible factor-1 $\alpha$ . *Cardiovascular Research* **60**, 569–579, doi: 10.1016/j.cardiores.2003.07.003 (2003).
68. Kimmel, C., Ballard, W., Kimmel, S., Ullmann, B. & Schilling, T. Stages of embryonic development of the zebrafish. *Developmental dynamics: an official publication of the American Association of Anatomists* **203**, 253–310, doi: 10.1002/aja.1002030302 (1995).
69. Reifers, F. *et al.* Fgf8 is mutated in zebrafish acerebellar (ace) mutants and is required for maintenance of midbrain-hindbrain boundary development and somitogenesis. *Development (Cambridge, England)* **125**, 2381–2395 (1998).
70. Naslin, J. *et al.* Stem cells in the adult zebrafish cerebellum: initiation and maintenance of a novel stem cell niche. *The Journal of neuroscience: the official journal of the Society for Neuroscience* **29**, 6142–6153, doi: 10.1523/JNEUROSCI.0072-09.2009 (2009).

## Acknowledgements

The authors would like to thank Karuna Sampath for providing the *spaw* and *lefty2* constructs, Brian Ciruna for providing the *arl13b-GFP* construct, Peter D Currie for providing the *gsc* construct, and Lee Miles for providing the *ntl* construct. This work was supported by an NHMRC project grant (GNT1068411), Monash University Faculty of Medicine and Nursing strategic grant and Operational Infrastructure Support from the Victorian Government. J.K. is supported by an Eva and Les Erdi fellowship. F.F. received an Australia Awards by the Australian government (AusAID). A.P. is supported by an Australian Research Council (ARC, FT140100047) Future Fellowship. We thank Monash Medical Micro Imaging Facility and Fish Core for excellent support.

## Author Contributions

F.F., Y.G., D.C., M.A., A.P., J.K.: concept and design, financial support, collection and/or assembly of data, data analysis and interpretation, manuscript writing, final approval of manuscript.

## Additional Information

**Supplementary information** accompanies this paper at <http://www.nature.com/srep>

**Competing financial interests:** The authors declare no competing financial interests.

**How to cite this article:** Frisca, F. *et al.* Role of ectonucleotide pyrophosphatase/phosphodiesterase 2 in the midline axis formation of zebrafish. *Sci. Rep.* **6**, 37678; doi: 10.1038/srep37678 (2016).

**Publisher's note:** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>

© The Author(s) 2016